



Complete High-Quality Genome Sequence of *Clostridium limosum* (*Hathewayia limosa*) Isolate 14S0207, Recovered from a Cow with Suspected Blackleg in Germany

Prasad Thomas,^{a*} Mostafa Y. Abdel-Gil,^a  Anne Busch,^a Lothar H. Wieler,^{b,c} Inga Eichhorn,^b Anne Bodenthin-Drauschke,^d Heinrich Neubauer,^a Christian Seyboldt^a

^aInstitute of Bacterial Infections and Zoonoses, Friedrich-Loeffler-Institut, Federal Research Institute for Animal Health, Jena, Germany

^bInstitute of Microbiology and Epizootics, Department of Veterinary Medicine, Freie Universität, Berlin, Germany

^cRobert Koch-Institut, Berlin, Germany

^dLandeslabor Schleswig-Holstein, Neumünster, Germany

ABSTRACT *Clostridium limosum* can be found in soil and the intestinal tract of animals. In 2014, *C. limosum* was isolated from a suspected blackleg outbreak in cattle in Schleswig-Holstein, Germany. We present a complete genome sequence of a *C. limosum* strain represented by a circular chromosome and three plasmids.

Clostridium is a genus of Gram-positive anaerobic bacteria within the phylum *Firmicutes*. The genus includes around 30 species that can cause clinical diseases in humans and animals, including birds. *Clostridium limosum* is a species that has received little attention in terms of its disease occurrence, prevalence, and virulence factors. The bacterium was found in various environments, including different animal and bird species (1). Recently, it was reported that the pathogen was the principal cause of metritis in farmed minks in Finland (2). In the current study, we isolated *C. limosum* from a suspected blackleg outbreak in cattle from Schleswig-Holstein, Germany. The organism was recovered from liver, spleen, and kidney tissues following anaerobic culture isolation methods. While the morphology on blood agar plates resembled *Clostridium chauvoei*, PCR results for the detection of *C. chauvoei* and *Clostridium septicum* (3) remained negative.

An initial categorization of the bacterial species using partial 16S rRNA gene sequencing (4) followed by a BLAST search (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>; blastn suite; 16S rRNA sequences for *Bacteria* and *Archaea* database) revealed homology to *C. limosum* (*Hathewayia limosa* Lawson and Rainey 2016 [5]) strain CECT 4329 (NCBI reference sequence number [NR_104825](https://.ncbi.nlm.nih.gov/nucl/NC_014825)).

The *C. limosum* isolate (14S0207) was cultured in 3 ml Selzer broth (6) under anaerobic conditions followed by genomic DNA extraction using a Genomic-tip 100/Q kit (Qiagen, Germany). GATC Biotech (Germany) carried out genome sequencing using a PacBio RS II sequencer (7), including the preceding library preparation to create a 10- to 20-kb insert size library. The total number of reads was 43,854 with an average length of 15,204 bp.

Additional sequencing using paired-end (2 × 300-bp) sequencing technology (MiSeq system) together with the Nextera XT library preparation protocol (Illumina, USA) was performed at the Institute of Bacterial Infections and Zoonoses in Jena, Germany. A total of 1,067,720 reads were received. The raw reads were checked for quality before and after read trimming using FastQC version 0.11.7 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>). Briefly, reads were trimmed using BBDuk (8) for adaptor removal (with the parameters $ktrim=r$, $k=23$, $mink=11$, $hdist=1$, tpe , and tbo) and Sickle version 1.33 (9) (with the parameter $-q 20$) for base quality.

Citation Thomas P, Abdel-Gil MY, Busch A, Wieler LH, Eichhorn I, Bodenthin-Drauschke A, Neubauer H, Seyboldt C. 2020. Complete high-quality genome sequence of *Clostridium limosum* (*Hathewayia limosa*) isolate 14S0207, recovered from a cow with suspected blackleg in Germany. *Microbiol Resour Announc* 9:e01487-19. <https://doi.org/10.1128/MRA.01487-19>.

Editor Steven R. Gill, University of Rochester School of Medicine and Dentistry

Copyright © 2020 Thomas et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Christian Seyboldt, Christian.Seyboldt@fli.de.

* Present address: Prasad Thomas, Division of Bacteriology and Mycology, ICAR-Indian Veterinary Research Institute, Izatnagar, India.

Received 3 December 2019

Accepted 4 December 2019

Published 9 January 2020

TABLE 1 Annotation features of *Clostridium limosum* 14S0207

Type ^a	NCBI RefSeq no. ^b	GenBank accession no.	Genome size (Mb)	GC content (%)	No. of proteins	No. of rRNAs	No. of tRNAs	No. of other RNAs	No. of genes	No. of pseudogenes
Chr	NZ_CP026600	CP026600	2.95	28.0	2,527	33	92	4	2,718	62
Plsm	NZ_CP026601	CP026601	0.14	25.5	125				132	7
Plsm	NZ_CP026602	CP026602	0.04	27.5	54				58	4
Plsm	NZ_CP026603	CP026603	0.03	26.4	30				31	1

^a Chr, chromosome; Plsm, plasmid.

^b RefSeq, reference sequence.

Genome assembly was done using the Hierarchical Genome Assembly Process algorithm version 3 (HGAP3) with default parameters (10) implemented in PacBio SMRT portal version 2.3.0. The seed length obtained during HGAP3 assembly was 10,613 bp (pre-assembled read length). HGAP3 assembly generated one contig representing the chromosome and three contigs representing the plasmids for the *C. limosum* isolate. The circularization of the received contig to a bacterial chromosome was carried out using a protocol recommended by PacBio for merging and circularization (<https://github.com/PacificBiosciences/Bioinformatics-Training/wiki/Circularizing-and-trimming>). The overlapping regions of circular sequences were determined using Gepard software version 1.40 with default parameters (11). For the circularization of contigs, Circlator version 1.5.0 was used with default parameters, as described before (12). The circular contigs representing the chromosome and plasmids were initially polished using PacBio long reads with the RS_Resequencing.1 protocol in SMRT portal version 2.3.0 followed by short Illumina reads using Pilon version 1.22 with default parameters (13). As a result, a complete genome is now available that meets high-quality standards. For the PacBio sequence data, we submitted the bax.h5 files and the methylation profiles to NCBI under BioProject number [PRJNA432648](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA432648) and SRA number [SRP216188](https://www.ncbi.nlm.nih.gov/sra/SRR216188).

The final assembly contained one circular chromosome and three plasmids (Table 1). The annotation was performed with the NCBI Prokaryotic Genome Annotation Pipeline (see Table 1). The chromosome carries a streptolysin-associated gene cluster encoding a streptolysin S (SLS) homolog (locus tag [C3495_10690](https://www.ncbi.nlm.nih.gov/locustags/C3495_10690), encoding 52 amino acids), a bacteriocin, and a virulence factor of group A *Streptococcus* known to possess hemolytic/cytolytic activity (14). Studies based on genomic analysis have identified a similar SLS-type gene cluster in *Clostridium* species, including *C. botulinum* and *C. sporogenes*, reported as the clostridiolysin S gene cluster (15).

Data availability. This whole-genome sequencing project has been deposited at DDBJ/EMBL/GenBank under the accession numbers [CP026600](https://www.ncbi.nlm.nih.gov/locustags/CP026600) (chromosome), [CP026601](https://www.ncbi.nlm.nih.gov/locustags/CP026601) (plasmid 1), [CP026602](https://www.ncbi.nlm.nih.gov/locustags/CP026602) (plasmid 2), and [CP026603](https://www.ncbi.nlm.nih.gov/locustags/CP026603) (plasmid 3). The raw sequence data are available under SRA accession numbers [SRR9822081](https://www.ncbi.nlm.nih.gov/sra/SRR9822081) (PacBio RS II) and [SRR9822082](https://www.ncbi.nlm.nih.gov/sra/SRR9822082) (Illumina MiSeq). The associated BioProject and BioSample accession numbers are [PRJNA432648](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA432648) and [SAMN08456352](https://www.ncbi.nlm.nih.gov/biosample/SAMN08456352), respectively. The versions described in this paper are the first versions.

ACKNOWLEDGMENTS

The ICAR-International Fellowship from the Indian Council of Agricultural Research (ICAR), New Delhi, India, for Prasad Thomas is gratefully acknowledged. Anne Busch was supported by a grant from the German Federal Ministry of Education and Research within the framework of the project Ess-B.A.R. (FKZ 13N13983). Mostafa Y. Abdel-Gliil received a DAAD (GERLS) Ph.D. scholarship.

REFERENCES

- Cato EP, Cummins CS, Smith LD. 1970. *Clostridium limosum* André in Prévot 1948, 165 amended description and pathogenic characteristics. *Int J Syst Evol Microbiol* 20:305–316. <https://doi.org/10.1099/00207173-20-3-305>.
- Bistrom M, Moisander-Jylha AM, Heinikainen S, Pelkola K, Raunio-Saarnisto M. 2016. Isolation of *Clostridium limosum* from an outbreak of metritis in farmed mink. *Acta Vet Scand* 58:49. <https://doi.org/10.1186/s13028-016-0230-7>.
- Sasaki Y, Yamamoto K, Amimoto K, Kojima A, Ogikubo Y, Norimatsu M, Ogata H, Tamura Y. 2001. Amplification of the 16S-23S rDNA spacer

- region for rapid detection of *Clostridium chauvoei* and *Clostridium septicum*. Res Vet Sci 71:227–229. <https://doi.org/10.1053/rvsc.2001.0495>.
4. Kuhnert P, Capaul SE, Nicolet J, Frey J. 1996. Phylogenetic position of *Clostridium chauvoei* and *Clostridium septicum* based on 16S rRNA gene sequences. Int J Syst Bacteriol 46:1174–1176. <https://doi.org/10.1099/00207713-46-4-1174>.
 5. Lawson PA, Rainey FA. 2016. Proposal to restrict the genus *Clostridium* Prazmowski to *Clostridium butyricum* and related species. Int J Syst Evol Microbiol 66:1009–1016. <https://doi.org/10.1099/ijsem.0.000824>.
 6. Selzer J, Hofmann F, Rex G, Wilm M, Mann M, Just I, Aktories K. 1996. *Clostridium novyi* alpha-toxin-catalyzed incorporation of GlcNAc into Rho subfamily proteins. J Biol Chem 271:25173–25177. <https://doi.org/10.1074/jbc.271.41.25173>.
 7. Eid J, Fehr A, Gray J, Luong K, Lyle J, Otto G, Peluso P, Rank D, Baybayan P, Bettman B, Bibillo A, Bjornson K, Chaudhuri B, Christians F, Cicero R, Clark S, Dalal R, Dewinter A, Dixon J, Foquet M, Gaertner A, Hardenbol P, Heiner C, Hester K, Holden D, Kearns G, Kong X, Kuse R, Lacroix Y, Lin S, Lundquist P, Ma C, Marks P, Maxham M, Murphy D, Park I, Pham T, Phillips M, Roy J, Sebra R, Shen G, Sorenson J, Tomaney A, Travers K, Trulson M, Veceli J, Wegener J, Wu D, Yang A, Zaccarin D, Zhao P, Zhong F, Korlach J, Turner S. 2009. Real-time DNA sequencing from single polymerase molecules. Science 323:133–138. <https://doi.org/10.1126/science.1162986>.
 8. Bushnell B. 2017. BMAP software package. <http://sourceforge.net/projects/bbmap/>.
 9. Joshi NA, Fass JN. 2011. Sickle: a sliding-window, adaptive, quality-based trimming tool for FastQ files. Version 1.33. <https://github.com/najoshi/sickle>.
 10. Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Non-hybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
 11. Krumsiek J, Arnold R, Rattei T. 2007. Gepard: a rapid and sensitive tool for creating dotplots on genome scale. Bioinformatics 23:1026–1028. <https://doi.org/10.1093/bioinformatics/btm039>.
 12. Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. Genome Biol 16:1–10. <https://doi.org/10.1186/s13059-015-0849-0>.
 13. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
 14. Nizet V, Beall B, Bast DJ, Datta V, Kilburn L, Low DE, De Azavedo J. 2000. Genetic locus for streptolysin S production by group A *Streptococcus*. Infect Immun 68:4245–4254. <https://doi.org/10.1128/iai.68.7.4245-4254.2000>.
 15. Gonzalez DJ, Lee SW, Hensler ME, Markley AL, Dahesh S, Mitchell DA, Bandeira N, Nizet V, Dixon JE, Dorrestein PC. 2010. Clostridiolysin S, a post-translationally modified biotoxin from *Clostridium botulinum*. J Biol Chem 285:28220–28228. <https://doi.org/10.1074/jbc.M110.118554>.