

## ORIGINAL RESEARCH

# Distal bile duct carcinomas and pancreatic ductal adenocarcinomas: postulating a common tumor entity

Rosa B. Schmuck, Cynthia V. de Carvalho-Fischer, Christopher Neumann, Johann Pratschke & Marcus Bahra

General, Visceral and Transplantation Surgery, Charité - Universitätsmedizin Berlin, Berlin, Germany

## Keywords

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## Correspondence

Rosa B. Schmuck, General, Visceral, and Transplantation Surgery, Charité - Universitätsmedizin Berlin, Augustenburger Platz 1, 13353 Berlin, Germany.  
Tel: +49 30 450 652184;  
Fax: +49 30 450 7652184;  
E-mail: rosa.schmuck@charite.de

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## Introduction

The defining feature of a tumor has hitherto been its organ of origin, thus determining the treatment strategy that is most beneficial for patients suffering from a malignant disease. It follows from this concept that carcinomas of the distal bile duct (dCC) and ductal adenocarcinomas of the pancreas (PDAC) are defined as two independent tumor entities. This is in accordance with the current WHO classification of tumors, which distinguishes between tumors of the liver and intrahepatic bile ducts, tumors of the gallbladder and extrahepatic bile ducts, and tumors of the exocrine pancreas. Here, dCC is defined by the location of the main tumor mass only, not by the microscopic aspect [1]. Given the spatial proximity of the duodenal curve, the pancreatic head, and the extrahepatic bile duct in the epigastric region, discriminating these single-organ structures with accuracy reveals constitutional limitations. The intertwined anatomy of the proximal pancreatic duct and the distal bile duct (pervading

## Abstract

The set definition of distal cholangiocarcinomas and adenocarcinomas of the pancreatic head is challenged by their close anatomical relation, similar growth pattern, and corresponding therapeutic outcome. They show a mutual development during embryologic organ formation and share phenotypic characteristics. This review will highlight the similarities with regard to the common origin of their primary organs, histopathological similarities, and modern clinical management. Thus, we propose to subsume those entities under a common superfamily.

the pancreatic head) gives ample reason to believe that both structures share more common features than previously believed. Moreover, both organs developed along similar embryological paths, and thus share numerous phenotypic characteristics. This hypothesis is corroborated by the fact that tumors of the distal bile duct and the pancreatic head also share the functional characteristics: a similar growth pattern, poor response to conventional chemotherapy, and a fairly unfavorable prognosis. We postulate that dCC and PDAC are a common tumor entity or should at least be subsumed under a common superfamily. Ampullary cancer, more particularly the pancreatobiliary subtype, should be considered as a part of this superfamily of tumors of the pancreatobiliary junction. The objective of this review is to define the common features as well as certain differences between these tumor entities, with regard to the embryonal development of the organ of origin, their diagnostic discrimination, histopathological and molecular similarities, and surgical and oncological treatment.

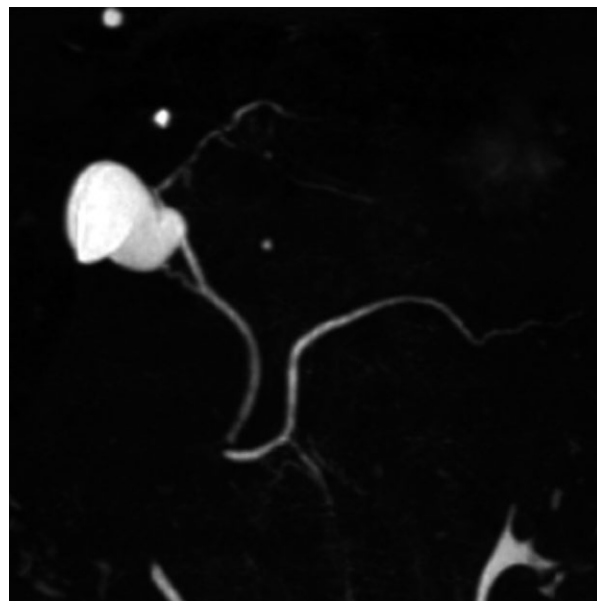
## The Embryologists View

Although biliary epithelial cells coat both the intrahepatic as well as the extrahepatic bile ducts, their developmental origins are distinct depending on their location in the biliary tree.

During the development of the gastrointestinal system, the endoderm forms the primitive gut. In the junction between the foregut and midgut that evolves into the duodenum, two outpunchings of the foregut form the so-called ventral and the dorsal bud, which serve as the basis for the pancreas and parts of the bile duct system. The larger bud, located on the dorsal aspect of the primitive gut forms the cranial part of the pancreatic head, the body, and the tail of the pancreas including their corresponding ductal tree. The smaller ventral bud forms the caudal section of the pancreatic head, the uncinate process and the proximal part of the main pancreatic duct (Duct of Wirsung). The distal bile duct also has its origin in the ventral bud and temporarily forms a common channel with the proximal part of the main pancreatic duct. As the embryo develops, the ventral bud, containing the primitive proximal pancreatic duct and distal bile duct, rotates dorsally and fuses with the dorsal bud. The proximal part of the pancreatic duct system in the dorsal bud obliterates completely. In some cases this obliteration is incomplete and the mostly nonfunctional, accessory pancreatic duct (duct of Santorini) persists. The remaining ductules of the dorsal bud fuse with the ductal system of the ventral duct. Meanwhile, the distal bile duct and the pancreatic duct separate and remain connected solely at the duodenal conjunction, the so-called ampulla of Vater [2, 3]. If the rotation, and particularly the fusion of the ventral and dorsal duct are impaired, two separate pancreatic ducts remain and a so-called *pancreas divisum* is formed (Fig. 1).

The hepatic diverticulum, which develops from the ventral foregut and remains attached to the above-mentioned ventral bud, forms the basis for liver development. Mesenchymal cells surrounding the septum transversum induce progenitor cell differentiation into hepatoblasts, and stimulate the creation of the liver's glandular structure. In the course of this development, biliary epithelial cells arise from hepatoblasts in the periportal layer. During the following phase of ductal plate remodeling, focal dilations of the aforementioned biliary epithelial cell precursors form bile ducts. Multiple defects in ductal plate remodeling lead to the formation of cysts rather than bile ducts, resulting in the so-called Alagille syndrome [4].

In conclusion, the proximal pancreatic and the distal bile duct arise from common endodermal structures, thus reveal mutual characteristics in organ development and formation. In contrast, the pancreatic duct leading from central and caudal part of the pancreas originates from a separate



**Figure 1.** Pancreas divisum with main pancreatic duct and duct of Santorini in endoscopic retrograde cholangiopancreatography.

endodermal portion. The intrahepatic bile duct system in contrast to the extrahepatic bile duct, takes its origin from periportal hepatoblasts in the hepatic diverticulum.

## PDAC and dCC: Analogies in Tumor Genesis

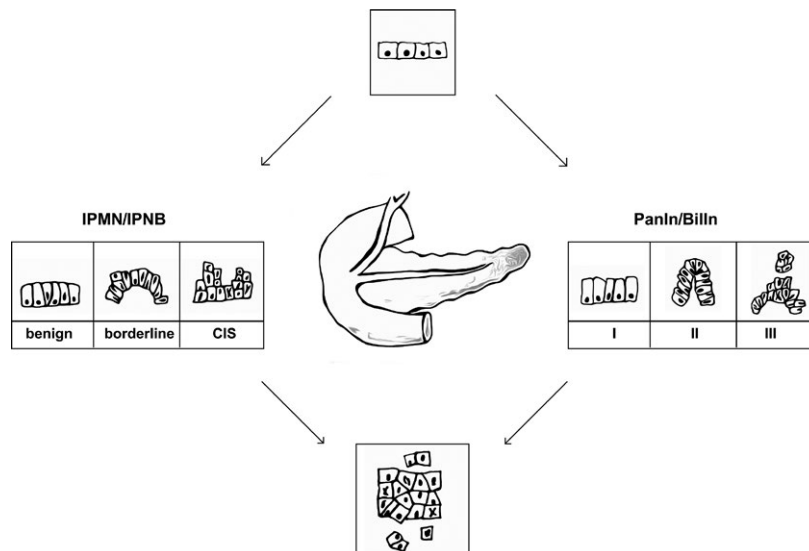
Intrahepatic cholangiocarcinomas (IH-CC) are especially known for their histological diversity. Until recently, it was thought that this neoplasm is mostly likely derived from biliary epithelial cells, whereas it is now thought that this cancer can alternatively take its origin from hepatic progenitor cells. In opposition, extrahepatic cholangiocarcinoma (EH-CC) arises from the biliary epithelium and the peribiliary glands [5]. It is therefore of particular interest that a subset of pancreatic exocrine acini are physiologically interspersed within these same peribiliary glands [6]. Three cell types have been identified in these acini: acinar cells with eosinophilic zymogen-like granules, clear cells resembling centroacinar cells and ductular elements. Additionally, peribiliary glands have been found to contain trypsin and amylase enzymes [7]. A recent comparative study in transgenic mice as well as human tissue, focusing on precursors of PDAC underlined that PDACs originate from the ductal system and adjacent structures [8]. It has been suggested that PDAC develops in the centroacinar–acinar compartment by acinar–ductal metaplasia. Pancreatic intraepithelial neoplasia (PanIN) lesions have been shown to be the precursor lesions of PDAC [9]. In transgenic mouse models, mucin-producing

ductular structures are frequently observed within the acinar parenchyma. These structures could represent an intermediate stage preceding the development of mouse PanIN. Another study analyzed the incidence and distribution of these ductular structures with and without mucinous metaplasia (mucinous tubular complexes/MCTs or tubular complexes/TCs). It could be demonstrated that the MCTs and TCs are common lesions in human pancreas. Furthermore, the expression of acinar markers, for example, trypsin was upheld in PanIN lesions [10]. In conclusion, these results indicate that the origin of PDAC lies in the periphery of the ductal system. Therein lies a strong parallelism to the origin of EH-CC, which also arises from the peribiliary and therefore periductal glands.

Intraepithelial neoplasia have been identified as premalignant lesions in several organs such as the pancreas (PanIN) or the prostate (PIN). Biliary intraepithelial neoplasia (BilIN) is assumed to be premalignant lesions of the bile duct system [11]. BilINs are located both in the extrahepatic bile ducts and in the major intrahepatic bile ducts. By analogy to ductal carcinomas of the pancreas (PanIN 1 to PanIN 3), BilINs are classified into three grades [12]. BilIN-1 describes low-grade dysplasia with mild cellular/nuclear atypia, BilIN-2 is defined as intermediate grade whereas BilIN-3 stands for high-grade dysplasia and contains significant cellular and nuclear atypia. On account of the fact that BilIN-3 corresponds to a carcinoma in situ, it is considered as an overt cancer of the bile ducts. A comparison between BilIN and PanIN reveals impressive similarities. In conclusion, BilIN should be seen as a biliary counterpart of PanIN (Fig. 2).

Recently, intraductal papillary mucinous neoplasms (IPMN) of the pancreas were identified as precursor lesions with high malignant potential [13]. Especially main duct IPMNs are associated with a high risk for transformation into intraductal papillary mucinous carcinomas (IPMC) [14]. It is current consensus that two different precursor lesions in the pancreas have to be distinguished, namely PanINs and IPMNs. If one assumes that BilINs are the biliary counterpart of PanINs, is there also a biliary counterpart for IPMN? Biliary papillomas/papillomatosis may present suitable candidates, due to their role as premalignant lesions for invasive biliary carcinomas. Furthermore, biliary papillomas/papillomatosis and IPMN possess similar clinicopathological features [15]. By virtue of this analogy, an alternative name was proposed for biliary papillomas/papillomatosis, namely the intraductal papillary neoplasm of the biliary tract (IPNB).

IPNBs are located in the extrahepatic bile duct (primarily hilar and distal), as well as in larger intrahepatic bile ducts. Furthermore, invasive IPNB lesions show elements from both tubular adenocarcinoma and mucinous carcinoma, which is also a well-studied invasion pattern in IPMNs. In conclusion, biliary preneoplastic lesions and pancreatic precursor lesions share common characteristics. Both, PanINs and IPMNs have a corresponding counterpart in the biliary tract: BilINs and IPNBs (Fig. 2). Premalignant lesions congruent with those concepts have not yet been described for ampullary cancer. The Ampulla Vateri is the epithelial junction of the main pancreatic duct and the distal bile duct. Ampullary cancer may arise either from the intestinal epithelium or the pancreatobiliary ducts [16]. The pancreatobiliary subtype



**Figure 2.** Biliary preneoplastic lesions and pancreatic precursor lesions share common characteristics. Both, PanINs (Intraepithelial neoplasia of the pancreas) and IPMNs (intraductal papillary mucinous neoplasms) have a corresponding counterpart in the biliary tract: BilINs (Biliary intraepithelial neoplasia) and IPNBs (intraductal papillary neoplasm of the biliary tract).

shows a positive correlation with a higher rate of tissue invasion, lymph node metastasis, and a worse outcome, than the intestinal type [17]. Thus, it has been hypothesized that the pancreatobiliary subtype should be seen as a very distal PDAC or dCC. In accordance with the aforementioned concept, PDAC and dCC and ampullary adenocarcinoma should be considered as a common tumor entity [16]. Never the less, the focus of this review remains the comparison of PDAC and dCC subentities.

## Molecular Pattern: Differences and Analogies Between PDAC and EH-CC

The two most commonly mutated genes in PDAC are p53 and KRAS. Activating KRAS mutations can be found in up to 95% of PDACs [18]. It is of particular interest that KRAS mutations are present in 90% of early stage PDACs, as well as in premalignant lesions such as PanINs [19, 20]. Through the analysis of KRAS mutations in biliary tract neoplasms and their corresponding premalignant lesions, Hsu et al. proved a similar process of carcinogenesis in the progression of BilINs to CCs [21]. Moreover, a recent study reports that KRAS mutations are present in 61.1% of the ampullary cancers, but only in 15.2% of bile duct and 2.7% of gallbladder cancers [22].

One of the key players of physiological apoptotic response, tumor suppressor gene p53, is frequently subject to mutations in CC as well as PDAC. Khan et al. reported mutation rates of 20% to 61% for dCC [23], and another study determined that a low (0–30%) p53 expression showed a favorable prognostic factor in patients with resected dCC [24]. In PDAC, p53 mutations can be found in up to 70% of primary, and in more than 50% of metastatic pancreatic cancers [25–27].

Unlike KRAS, p53 mutations seem to be a late event in the development of PanINs to PDAC [20, 28]. Nevertheless, p53 has already been the subject of therapeutic approaches in PDAC. Transfer of the p53 gene via vectors has led to an inhibition of tumor cells in vitro as well as in vivo [29, 30]. It is of further note that mutant p53 stimulates chemoresistance against gemcitabine, and the reintroduction (*reactivation*) of p53 increases the cytotoxicity of gemcitabine [31, 32]. Despite the aforementioned results, no attempt to evaluate p53 as a therapeutic target has been made in CC.

In recent years, miRNAs have emerged as diagnostic, as well as therapeutic targets in multiple cancers. miRNAs are small, noncoding RNAs that influence multiple physiological and pathological processes on the post-transcriptional level. Furthermore, they have been identified as crucial players in carcinogenesis due to their role as both oncogenes and tumor suppressors. miR-21 as well as let-7a undergo an upregulation in PDAC and CC, [33] and may additionally modulate gemcitabine-induced apoptosis, thus lending these

molecules a potential therapeutic relevance [34]. In a similar study, the targeted inhibition of miR-21 via lentoviruses leads to a significant regression of PDAC tumor growth in vivo [35].

Despite these promising results, current literature does not enable a direct comparison of miRNA expression levels in PDAC and CCs. Furthermore, many projects do not distinguish between IH-CCs and EH-CCs, rendering a scientifically based conclusion on the similarities and difference between PDAC and dCCs impossible (Table 1) [36].

## The Pathologists View: Are There Phenotypical Similarities Between PDAC and dCC?

The diagnosis of PDAC has hitherto been based on the identification of tumor markers such as CEA, CK19-9, CK7, and CK20, in conjunction with its histological and clinical characteristics.

Cytokeratin (CK) 7 is a pancreatobiliary marker expressed in IH-CC, EH-CC, and pancreatic neoplasms [37–42]. Similarly, CEA and CK19-9 are also expressed by all three tumor entities [16]. Cytokeratin 20 shows a more nuanced expression, in that it revealed high expression in EH-CCs and variable negative expression in IH-CCs [39]. In PDAC, CK20 expression was highest in tumors with a high number of reactive cells [40], despite an overall more common CK7+/CK20-phenotype [40–42]. Nevertheless, other studies suggest higher expression rates (65%) for CK20 in pancreatic cancer, and a plausible negative prognostic significance of the marker [43]. Interestingly, the pancreatobiliary subtype of ampullary cancer lacks CK20 in majority of cases whereas the intestinal subtype shows a high CK20 expression [16, 44].

In conclusion, common tumor markers such as CK7, CEA, and CK19-9 are unable to differentiate IH-, EH-CCs, and PDAC. Moreover, although CK20 shows a subtler expression profile (low in IH-CC, high in EH-CC, and variable expression in PDAC), it demonstrates a high biological and methodical variability, rendering it an unreliable marker for pancreatobiliary cancers.

The inadequacies of current tumor marker panels have led to the exploration of alternative indicator molecules, among which proteins from the mucin and cadherin family have taken on a distinctive role. Such research could be valuable for identifying potential tumor subentities and further describing their clinical behavior, that is, drug response, overall survival, and rate of metastasis/invasion. Specific targeting of strongly expressed antigens could also lead to personalized therapeutic interventions.

Mucins (MUC) are high-molecular, highly glycosylated proteins that form biological gels as a means of protecting epithelia from their external conditions. Furthermore,

**Table 1.** Summary of similarities and differences in IH-CC, EH-CC, dCC, and PDAC.

	IH-CC	EH-CC and dCC	PDAC	References
Developmental origin	Mesenchymal cells surrounding the septum transversum form small intrahepatic bile ducts	Ventral part of forgut forms main extrahepatic bile ducts	Ventral part of forgut forms mainpancreatic duct	[2–4]
Tumor genesis	Hepatocytes, hepatic progenitor cells or BECs	Ductal system or periductal glands	Ductal system or periductal glands	[5–8]
Premalignant lesions	No corresponding lesions in IH-CC	Biliary intraepithelial neoplasia ( <i>BilIN</i> )	Pancreatic intraepithelial neoplasia ( <i>PanIn</i> )	[9–12]
Precursor lesions with malignant potential	No corresponding lesions in IH-CC	Intraductal papillary mucinous neoplasms ( <i>IPMN</i> )	Intraductal papillary neoplasm of the biliary tract ( <i>IPNB</i> )	[13–15]
Molecular pattern	KRAS+/- p53+/- CK20-	KRAS+ p53+ CK20+	KRAS+ p53+ CK20+	[18–27]
Phenotype	MUC1+/- MUC4+ (only larger bile ducts) S100P+ (only larger bile ducts)	MUC1+ MUC4 + S100P +	MUC1+ MUC4+ S100P+	[39–43, 46–52, 55–57]
Surgery	Hemihepatectomy	PPPD/Kausch-Whipple; extended hemihepatectomy for Klatskin-Tumors	PPPD/Kausch-Whipple	[72–75]
Response to Chemotherapy	5-FU+ Gemcitabine- CapOx- Gemcitabine+ cisplatin+ nab-Paclitaxel+/?	5-FU+ Gemcitabine+ CapOx+ Gemcitabine+ cisplatin+ nab-Paclitaxel+/?	5-FU+/- Gemcitabine+ CapOx+ Gemcitabine+ cisplatin- nab-Paclitaxel+	[78–97]

mucins play an important role in cell-to-cell signaling, and inducing immune reactions [45]. A set of twenty different mucins has been identified in human epithelia, each glycosylated differently in order to carry out a specific function [46]. Despite this biological variety, this comparative analysis will focus on solely MUC1 and MUC4 as potential tumor markers.

MUC1 has been identified in both pancreatic and so-called “mucin-producing” cholangiocarcinomas derived from the large bile ducts of the liver [47–50]. Tamada et al. revealed that while MUC1 is not expressed in normal epithelia of the bile duct, it can be found in 87% of EH-CC tumors analyzed ( $n = 70$ ) [51]. More importantly, MUC1 showed an analogous expression pattern in biliary intraepithelial and pancreatic intraepithelial neoplasms [52]. Furthermore, MUC1 may promote cell adhesion in foreign tissues, and thus allows for PDAC invasion [53]. In comparison to the above mentioned MUC1-expressing tumors, mixed-type CCs, and cholangiocarcinomas (deriving from hepatic progenitor cells rather than cholangiocytes) only produce negligible amounts of MUC1 [47].

Interestingly Remmers et al. showed that MUC1's expression and glycosylation pattern transformed from normal tissue to premalignant and cancerous tissues of the pancreas [46]. This study detailed that unglycosylated MUC1 was highest in metastatic tissue, whereas “T-MUC1” (a specifically glycosylated form of the protein) expression was

lowest in normal tissue. Hence, MUC1 could potentially be used to map the progression of normal to malignant tissue in both pancreatic and extrahepatic biliary cancer.

Although MUC1 functions as a membrane receptor, MUC4 may bind to the ErbB2 receptor, possibly increasing p27 Expression [54]. Several studies have shown that while MUC4 is not present in normal liver or pancreatic tissue, it is present in PDAC, EH-CC, and IH-CC derived from the larger bile ducts [46, 51, 55–57]. More importantly, it has been identified as a negative prognostic factor for survival in all three tumor types correlating with higher metastatic and lymphatic invasion [55–57]. In IPMN's of the pancreas, MUC4 may enable a differentiation between malignancies of the intestinal type and the gastric type [56]. Unfortunately, data on MUC4 expression in mixed-type CCs is scant.

Proteins from the cadherin family have also recently come into the spotlight as potential indicator substances for gastrointestinal neoplasms. Cadherins play an important role in cell adhesion and cell communication, both as receptors and ligands within signaling pathways. Additionally, it has been postulated that during epithelial–mesenchymal transition (EMT) a so-called “cadherin switch” comes into effect, in which E-Cadherin, typically expressed in epithelia, is replaced by N-Cadherin, found mostly in mesenchymal tissue [58]. E-Cadherin has proved

to be a protective factor in both CC and PDAC [59]. Nitta and Mitsuhashi et al. recently showed that N-Cadherin was a negative prognostic factor for survival in EH-CC, a finding confirmed by Araki et al. [60, 61]. Elevated expression of N-Cadherin in IH-CC was also linked to aggressive behavior with increased metastasis and invasion [62]. However, a recent study was able to show that N-Cadherin expression was significantly lower in metastatic liver PDAC than in CCs, possibly enabling a differentiation of these tumor subtypes in conjunction with other markers [63]. Unfortunately, this study did not discriminate between subentities of cholangiocarcinoma.

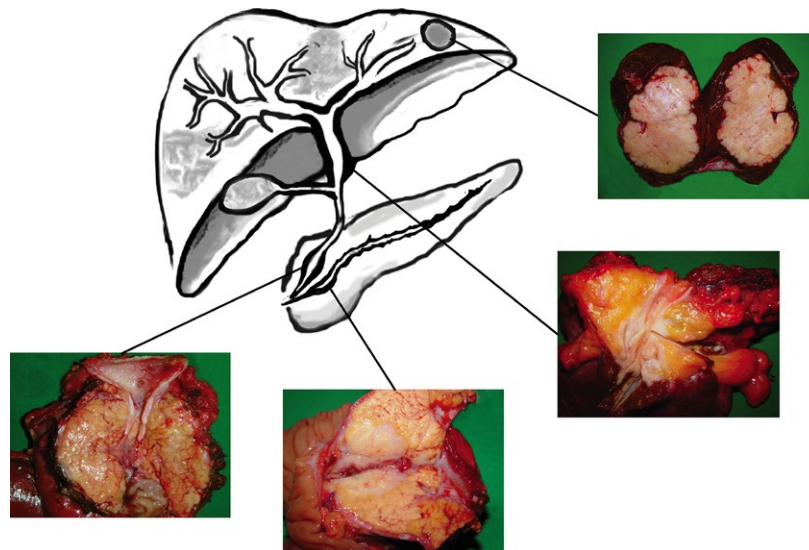
Finally, S100p encodes a family of proteins responsible for regulating differentiation and proliferation. Ali et al. have identified this protein as a highly specific and sensitive marker for pancreatobiliary tumors, achieving 100% specificity and 83% sensitivity on its own [64]. By means of a histological panel including S100p, MUC1, KOC, and mesothelin, the diagnosis of pancreatobiliary tumors was achieved to nearly 100% specificity and sensitivity, in cytological specimens sampled through endoscopic retrograde cholangiopancreatography (ERCP). Other studies confirm high S100p expression in PDAC and EH-CC, in contrast to infrequent expression in normal pancreatobiliary tissue [65]. A recent study showed that S100P staining identified IH-CCs with bile duct morphology (a.k.a derived from the larger ducts of the biliary tree) to have a similar expression profile (CEA+, CK19-9+, MUC2+, and more likely N-Cadherin negative) to EH-CC and PDAC [66]. S100p negative tumors showed less of a bile duct morphology and were more likely to be N-Cadherin negative, Liau et al. confirmed a higher expression of S100p in IH-CC of bile duct morphology.

Although EH-CC and PDAC show many similarities concerning the expression of MUC1 and S100P, IH-CC derived from the small bile ducts seem to have a different phenotype. MUC4, N-Cadherin may also prove to be useful markers, given further study. In conclusion, standardized immunohistochemical panels including atypical marker proteins should be further evaluated in order to improve the accuracy of IH-CC and EH-CC diagnosis.

### Assigning a Diagnosis: Imaging Strategies for PDACs and dCCs in Light of Common Features and Anatomical Difficulties

Diagnostic methods for discriminating cancers of the pancreatic head and the distal bile duct often overlap. In this chapter, we aim to highlight common features and challenges in the discernment of the abovementioned entities.

The early detection of the tumor is crucial for the prognosis of PDAC and dCC patients. Both neoplasms arise from ductal epithelia, and therefore tend to grow in longitudinal alignment with the ductal system during the early stages of tumor formation. It is due to this mechanism that mass formation occurs mostly in the later stages of the tumor development, impeding an early, and thus potentially curative, image-based diagnosis. In contrast, IH-CCs show a distinct growth pattern (including concentric growth and mass formation), thus facilitating tumor detection in an abdominal ultrasound (US), computed tomography (CT), and/or magnetic resonance imaging (MRI) (Fig. 3). Therefore, the longitudinal rather than concentric growth pattern of PDAC and dCC severely



**Figure 3.** Tumors are classified according to their location (from top to bottom): intrahepatic cholangiocarcinoma (IH-CC), hilar cholangiocarcinoma, distal cholangiocarcinoma (dCC), and adenocarcinoma of the pancreas (PDAC).

restricts image-based screening, due to their limited sensitivity. Through a comparison of CT, MRI, magnetic resonance cholangiopancreatography (MRCP), and the endoscopic ultrasound (EUS), the CASPS3 study showed that the best visualization of small pancreatic lesions could be achieved by EUS and MRCP [67]. This result may be explained by the occurrence of neoplastic obstruction of the ductal system, which can occur in early stages of PDAC and dCC progression when the tumor itself is not yet detectable via radiological assessment. An additional benefit of the EUS is the possibility to take samples via fine needle aspiration or brush cytology. A sufficiently large tumor sample can discriminate between benign and malignant lesions in selected cases. Studies on intraductal endoscopic ultrasound (IEUS) took the EUS approach to dCC and PDAC identification a step further, and demonstrated that the diagnosis of bile duct strictures and lesions of the pancreatic head can be improved by the assessment of the ductal wall and its surrounding structures [68–70]. In comparison to IH-CCs and lesions of the caudal pancreas, PDACs and dCCs can be easily assessed via ERCP, EUS, and IEUS. Furthermore, the early diagnosis of the latter tumor entities is mostly based on the visualization of the concomitant cholestasis. A large portion of PDACs, dCCs as well as ampullary cancers clinical symptoms, result from cholestasis due to neoplastic obstruction. Therefore, tumors with a high cholestatic effect are often diagnosed at an earlier stage due to their prompt onset of symptoms. The same explanation may clarify the slightly higher survival of dCC as compared to PDAC: tumors located in the corpus or cauda of the pancreas will not cause cholestasis in the early stages of the condition. Another reason for this finding might be the fact that some studies include ampullary cancer (a neoplasm with an overall more favorable prognosis) into prognosis estimation of dCC [71].

Due to the contiguity of PDACs and dCCs, the discrimination of these neoplasms can be particularly challenging. A clear anatomical attribution can be especially difficult in later tumor stages, when malignant growth involves neighboring structures. Applying the hypothesis of a common superfamily for those tumor entities, a distinction according to the organ of origin would be obsolete. The common tumor entity approach becomes a depiction of clinical reality in that the only possibility for curative treatment, that is a radical surgical resection, is congruent concerning the surgical technique for PDAC and dCC.

### The Surgeons Point of View: Surgical Treatment Options in PDAC and CCs

Radical surgical resection with a microscopic tumor-free resection margin (R0), is the only curative option for PDAC and CC [72]. The guiding principle behind radical

resection applies to the whole range of tumors of the pancreatobiliary system, regardless of their location. Thus, the procedure of choice for hilar cholangiocarcinoma (Klatskin tumor) is an extended hemihepatectomy, with en bloc extrahepatic bile duct resection. This procedure has a clear advantage regarding the local recurrence rate and overall survival time, as compared to alternative strategies. Neuhaus et al. described a 1-, 3-, and 5-year survival rate after hilar en bloc resection, which included the removal of the portal vein bifurcation, of 87%, 70%, and 58%, respectively. Survival rates after conventional major hepatectomy were significantly lower with 1-, 3-, and 5-year survival rates of 79%, 40%, and 29%, respectively [73]. Due to the spatial proximity of the duodenum, the pancreatic head, the extrahepatic bile duct, and their overlapping blood supply, curative resections of both PDAC and dCC require a pancreatoduodenectomy, preferably under preservation of the pylorus (PPPD). In selected cases of ampullary cancer, a local resection via transduodenal ampullectomy or an endoscopic resection can be considered [74]. As a result of the longitudinal spread of the tumor in the bile duct wall, an additional hemihepatectomy has to be considered in selected cases in order to achieve an R0 resection [75]. From a surgeon's point of view, a suspected malignant lesion located in the pancreatic head – regardless of whether it arises from the pancreatic or the bile duct – needs to be removed in toto using an identical surgical technique.

### The Oncologists Perspective: How Do Pancreatobiliary Tumors Respond to Conventional Adjuvant Chemotherapy?

There are various therapeutic options for pancreatobiliary cancers, namely PDAC, IH-CC, and EH-CC [76, 77]. Nevertheless, most patients diagnosed with these types of cancer have a low survival rate and die within the first one to three years of diagnosis with or without chemotherapeutic treatment [78–80].

Common therapeutic agents for pancreatobiliary cancers include antimetabolites such as 5-fluorouracil (5-FU) [80, 81] and capecitabine [82–84], nucleoside analogs such as gemcitabine [85–90], platinum analogs such as oxaliplatin, [82–84] and cisplatin [91, 92]. In recent studies, chemicals known as taxenes in form of *nab*-paclitaxel, a 130-nm albumin-bound formulation of paclitaxel particles, have been evaluated to this end [93].

While 5-FU is not the first preference for adjuvant chemotherapy of PDAC and is only reported to have minimal effects on overall survival [81], it has a much bigger impact on IH-CC and EH-CC. A retrospective study by Yoshitda et al. including 26 patients with distal

bile duct cancer after pancreaticoduodenectomy, reports a 5-year-survival rate of 56% with adjuvant chemotherapy compared to 27% for untreated patients [80]. Similarly, another study carried out with a collective of 35 patients with intraductal papillary peripheral cholangiocarcinoma, accounts for a 5-year-survival rate of 33.3% for treated and 10.8% for untreated patients [79].

Gemcitabine, on the other hand, has been used very successfully in adjuvant chemotherapy to increase the overall survival rates of patients suffering from PDAC, exemplary shown within the scope of the CONKO Study by Sinn et al. [94, 95].

A multicentered, randomized, phase III, clinical trial compared the effects of gemcitabine with 5-FU for pancreatic carcinoma in first-line palliative setting. The study concluded that the clinical benefits (23.8% vs. 4.8%) as well as the 12-month survival rate (18% vs. 2%) were both significantly greater for gemcitabine than for 5-FU, respectively [90].

A similar therapeutic effect has been reported for Gemcitabine in EH-CC. According to Murakami et al. the 5-year-survival rate of patients who received adjuvant chemotherapy with gemcitabine after extended surgery for hilar cholangiocarcinoma, was noticeably higher (57%) compared to those who did not receive any postoperative treatment (23%) [85]. For IH-CC, however, Gemcitabine is reported to have minimal effect [86]. Thus, the clinical response to gemcitabine is similar for PDAC and EH-CC, in contrast to IH-CC.

Several alternative treatment protocols for pancreatobiliary cancers involve platinum-based therapeutic agents. The combination therapy of capecitabine and oxaliplatin (CapOx), for example, offers a different treatment for PDAC with similar outcomes to gemcitabine-related regimes [84]. A prospective, multicentre, phase II trial by Nehls et al. analyzed the response of 47 patients with EH-CC with respect to CapOx treatment, and compared the results with 18 patients with IH-CC who received the same chemotherapy. The response rate of the EH-CC collective was reported to be 27%, while there were no objective responses among the IH-CC patients. Additionally, 49% of the EH-CC had a stable disease after treatment compared to 33% for the IH-CC [83].

On the other hand, reports suggest that a gemcitabine and cisplatin combination therapy leads to different clinical responses. According to an extensive phase II study by Valle et al. both EH-CC and IH-CC have a better clinical response to the above mentioned combination therapy, as compared to a gemcitabine monotherapy [78]. In PDAC, however, this combination therapy only shows moderate activity, highlighted by a low response rate of 11% [88]. Hence, while PDAC and E-CC respond to CapOx analogically, the response is quite different for a gemcitabine and cisplatin combination therapy.

Recently, the use of nab-Paclitaxel in conjunction with gemcitabine for treating PDAC has/come into the spotlight. A contemporary phase III study compared a collective of 430 patients with PDAC on a gemcitabine monotherapy with 431 patients who received a combination of gemcitabine and nab-Paclitaxel. The treatment resulted in a higher median overall survival for the combination therapy collective (8.5 vs. 6.7 months), despite higher rates of peripheral neuropathy and myelosuppression [96]. However, there have been no extensive clinical trials in cholangiocarcinoma populations so far. A recent study by Kolinsky et al. examined the therapeutic effect of nab-Paclitaxel monotherapy for two patients with cholangiocarcinoma and suggests that nab-Paclitaxel does indeed have therapeutic activity [97]. Considering the similarities between PDAC and EH-CC relative to their therapeutic response, as outlined in this section, future studies examining the therapeutic activity of a nab-Paclitaxel and gemcitabine combination in EH-CC are more than justified. This is of particular significance as chemotherapy algorithms of dCC and PDAC currently diverge, particularly for advanced disease.

## Conclusion: New Thoughts and Concepts on Defining Pancreatobiliary Cancers

Carcinomas of the pancreatic head and dCCs share a wide range of common features. Both tumors possess a common embryologic origin, a marked anatomical overlap of the primary organ tissue, and several phenotypic analogies. All of these factors explain the difficulties in discriminating both tumor entities through imaging studies, and the exigency for radical surgical resection. Finally, similarities in the therapeutic outcomes of PDAC and extrahepatic bile duct cancer suggest strong behavioral analogies. Summarily, a new and unified superfamily for those tumor entities of the pancreatobiliary junction should be considered to more accurately define these neoplasms. We consider the definition of such a superfamily more adequate than the simple categorization of IH-CC and dCC as cholangiocarcinomas.

## Conflict of Interest

None declared.

## References

1. Hamilton, S. R., and L. A. Aaltonen, eds. 2000. World health organization classification of tumors. Pathology and genetics of tumors of the digestive system. IARC Press, Lyon.



2. Gray, H. 2000. *Anatomy of the Human Body*. Lea & Febiger, 1918, Philadelphia. Bartleby.com.
3. Ando, H. 2010. Embryology of the biliary tract. *Dig. Surg.* 27:87–89.
4. Zorn, A. M. 2008. Liver development. *StemBook*. The Stem Cell Research Community.
5. Cardinale, V., G. Carpino, L. Reid, E. Gaudio, and D. Alvaro. 2012. Multiple cells of origin in CCC underlie biological, epidemiological and clinical heterogeneity. *World J. Gastrointest. Oncol.* 4:94–102.
6. Terada, T., Y. Nakanuma, and A. Kakita. 1990. Pathologic observations of intrahepatic peribiliary glands in 1000 consecutive autopsy livers. Heterotopic pancreas in the liver. *Gastroenterology* 98:1333–1337.
7. Terada, T., and Y. Nakanuma. 1993. Development of human intrahepatic peribiliary glands. Histological, keratin immunohistochemical, and mucus histochemical analyses. *Lab. Invest.* 68:261–269.
8. Aichler, M., C. Seiler, M. Tost, J. Siveke, P. K. Mazur, P. Da Silva-Buttkus, et al. 2012. Origin of pancreatic ductal adenocarcinoma from atypical flat lesions: a comparative study in transgenic mice and human tissue. *J. Pathol.* 226:723–734.
9. Brat, D. J., K. D. Lillemo, C. J. Yeo, P. B. Warfield, and R. H. Hruban. 1998. Progression of pancreatic intraductal neoplasias to infiltrating adenocarcinoma of the pancreas. *Am. J. Surg. Pathol.* 22:163–169.
10. Zhu, L., G. Shi, C. M. Schmidt, R. H. Hruban, and S. F., Konieczny. 2007. Acinar cells contribute to the molecular heterogeneity of pancreatic intraepithelial neoplasia. *Am. J. Pathol.* 171:263–273.
11. Zen, Y., N. V. Adsay, K. Bardadin, R. Colombari, L. Ferrell, H. Haga, et al. 2007. Biliary intraepithelial neoplasia: an international interobserver agreement study and proposal for diagnostic criteria. *Mod. Pathol.* 20:701–709.
12. Zen, Y., S. Aishima, Y. Ajioka, J. Haratake, M. Kage, F. Kondo, et al. 2005. Proposal of histological criteria for intraepithelial atypical/proliferative biliary epithelial lesions of the bile duct in hepatolithiasis with respect to cholangiocarcinoma: preliminary report based on interobserver agreement. *Pathol. Int.* 55:180–188.
13. Hruban, R. H., K. Takaori, D. S. Klimstra, N. V. Adsay, J. Albores-Saavedra, A. V. Biankin, et al. 2004. An illustrated consensus on the classification of pancreatic intraepithelial neoplasia and intraductal papillary mucinous neoplasms. *Am. J. Surg. Pathol.* 28:977–987.
14. Tanaka, M., C. Fernandez-del Castillo, V. Adsay, S. Chari, M. Falconi, J. Y. Jang, et al. 2012. International consensus guidelines for the management of IPMN and MCN of the pancreas. *Pancreatology* 2012:183–197.
15. Cha, J. M., M. H. Kim, S. K. Lee, D. W. Seo, S. S. Lee, J. H. Lee, et al. 2006. Clinicopathological review of 61 patients with early bile duct cancer. *Clin. Oncol.* 18:669–677.
16. Perysinakis, I., I. Margaritis, and G. Kouraklis. 2014. Ampullary cancer—a separate clinical entity? *Histopathology* 64:759–768.
17. Kimura, W., N. Futakawa, S. Yamagata, Y. Wada, A. Kuroda, T. Muto, et al. 1994. Different clinicopathologic findings in two histologic types of carcinoma of papilla of Vater. *Jpn. J. Cancer Res.* 85:161–166.
18. Almoguera, C., D. Shibata, K. Forrester, J. Martin, N. Arnheim, and M. Perucho. 1988. Most human carcinomas of the exocrine pancreas contain mutant c-K-ras genes. *Cell* 53:549–554.
19. Deramaudt, T., and A. K. Rustgi. 2005. Mutant KRAS in the initiation of pancreatic cancer. *Biochim. Biophys. Acta* 1756:97–101.
20. Murphy, S. J., S. N. Hart, J. F. Lima, B. R. Kipp, M. Klebig, J. L. Winters, et al. 2013. Genetic alterations associated with progression from pancreatic intraepithelial neoplasia to invasive pancreatic tumor. *Gastroenterology* 145:1098–1109.
21. Hsu, M., M. Sasaki, S. Igarashi, Y. Sato, and Y. Nakanuma. 2013. KRAS and GNAS mutations and p53 overexpression in biliary intraepithelial neoplasia and intrahepatic cholangiocarcinomas. *Cancer* 119:1669–1674.
22. Rashid, A., T. Ueki, Y. T. Gao, P. S. Houlihan, C. Wallace, B. S. Wang, et al. 2002. K-ras mutation, p53 overexpression, and microsatellite instability in biliary tract cancers: a population-based study in China. *Clin. Cancer Res.* 8:3156–3163.
23. Khan, S. A., H. C. Thomas, M. B. Toledano, and I. J. Cox. 2005. Taylor- Robinson SD: p53 Mutations in human cholangiocarcinoma: a review. *Liver Int.* 25:704–716.
24. Rijken, A. M., G. J. Offerhaus, M. M. Polak, D. J. Gouma, and T. M. van Gulik. 1999. p53 expression as a prognostic determinant in resected distal bile duct carcinoma. *Eur. J. Surg. Oncol.* 25:297–301.
25. Scarpa, A., P. Capelli, K. Mukai, G. Zamboni, T. Oda, C. Iacono, et al. 1993. Pancreatic adenocarcinomas frequently show p53 gene mutations. *Am. J. Pathol.* 142:1534–1543.
26. Berrozpe, G., J. Schaeffer, M. A. Peinado, F. X. Real, and M. Perucho. 1994. Comparative analysis of mutations in the p53 and K-ras genes in pancreatic cancer. *Int. J. Cancer* 58:185–191.
27. Moore, P. S., S. Beghelli, G. Zamboni, and A. Scarpa. 2003. Genetic abnormalities in pancreatic cancer. *Mol. Cancer* 2:7.
28. Sasaki, S., H. Yamamoto, H. Kaneto, I. Ozeki, Y. Adachi, H. Takagi, et al. 2003. Differential roles of alterations of p53, p16, and SMAD4 expression in the progression of intraductal papillary-mucinous tumors of the pancreas. *Oncol. Rep.* 10:21–25.

29. Bouvet, M., R. J. Bold, J. Lee, D. B. Evans, J. L. Abbruzzese, P. J. Chiao, et al. 1998. Adenovirus-mediated wild-type p53 tumor suppressor gene therapy induces apoptosis and suppresses growth of human pancreatic cancer. *Ann. Surg. Oncol.* 5:681–688.
30. Hwang, R. F., E. M. Gordon, W. F. Anderson, and D. Parekh. 1998. Gene therapy for primary and metastatic pancreatic cancer with intraperitoneal retroviral vector bearing the wild-type p53 gene. *Surgery* 124:143–150.
31. Fiorini, C., M. Cordani, C. Padroni, G. Blandino, S. Di Agostino, and M. Donadelli. 2015. Mutant p53 stimulates chemoresistance of pancreatic adenocarcinoma cells to gemcitabine. *Biochim. Biophys. Acta* 1853:89–100.
32. Hill, R., M. Rabb, P. A. Madureira, D. Clements, S. A. Gujar, D. M. Waisman, et al. 2013. Gemcitabine-mediated tumor regression and p53-dependent gene expression: implications for colon and pancreatic cancer therapy. *Cell Death Dis.* 4:791.
33. Nair, V. S., L. S. Maeda, and J. P. Ioannidis. 2012. Clinical outcome prediction by microRNAs in human cancer: a systematic review. *J. Natl Cancer Inst.* 104:528–540.
34. Meng, F., R. Henson, M. Lang, H. Wehbe, S. Maheshwari, J. T. Mendell, et al. 2006. Involvement of human micro-RNA in growth and response to chemotherapy in human cholangiocarcinoma cell lines. *Gastroenterology* 130:2113–2129.
35. Sicard, F., M. Gayral, H. Lulka, L. Buscail, and P. Cordelier. 2013. Targeting miR-21 for the therapy of pancreatic cancer. *Mol. Ther.* 21:986–994.
36. Collins, A. L., S. Wojcik, J. Liu, W. L. Frankel, H. Alder, L. Yu, et al. 2014. A differential microRNA profile distinguishes cholangiocarcinoma from pancreatic adenocarcinoma. *Ann. Surg. Oncol.* 21:133–138.
37. Gyure, K. A., and A. L. Morrison. 2000. Cytokeratin 7 and 20 expression in choroid plexus tumors: utility in differentiating these neoplasms from metastatic carcinomas. *Mod. Pathol.* 13:638–643.
38. Lau, S. K., S. Prakash, S. A. Geller, and R. Alsabeh. 2002. Comparative immunohistochemical profile of hepatocellular carcinoma, cholangiocarcinoma, and metastatic adenocarcinoma. *Hum. Pathol.* 33:1175–1181.
39. Rullier, A., B. Le Bail, R. Fawaz, J. F. Blanc, J. Saric, and P. Bioulac-Sage. 2000. Cytokeratin 7 and 20 expression in cholangiocarcinomas varies along the biliary tract but still differs from that in colorectal carcinoma metastasis. *Am. J. Surg. Pathol.* 24:870–876.
40. Goldstein, N. S., and D. Bassi. 2001. Cytokeratins 7, 17, and 20 reactivity in pancreatic and ampulla of Vater adenocarcinomas. Percentage of positivity and distribution is affected by the cut-point threshold. *Am. J. Clin. Pathol.* 115:695–702.
41. Bayrak, R., H. Haltas, and S. Yenidunya. 2012. The value of CDX2 and cytokeratins 7 and 20 expression in differentiating colorectal adenocarcinomas from extraintestinal gastrointestinal adenocarcinomas: cytokeratin 7-/20+ phenotype is more specific than CDX2 antibody. *Diagn. Pathol.* 7:9.
42. Chu, P., E. Wu, and L. M. Weiss. 2000. Cytokeratin 7 and cytokeratin 20 expression in epithelial neoplasms: a survey of 435 cases. *Mod. Pathol.* 13:962–972.
43. Matros, E., G. Bailey, T. Clancy, M. Zinner, S. Ashley, E. Whang, et al. 2006. Cytokeratin 20 expression identifies a subtype of pancreatic adenocarcinoma with decreased overall survival. *Cancer* 106:693–702.
44. Zhou, H., N. Schaefer, M. Wolff, and H. P. Fischer. 2004. Carcinoma of the ampulla of Vater: comparative histologic/immunohistochemical classification and follow-up. *Am. J. Surg. Pathol.* 28:875–882.
45. Hollingsworth, M. A., and B. J. Swanson. 2004. Mucins in cancer: protection and control of the cell surface. *Nat. Rev. Cancer* 4:45–60.
46. Remmers, N., J. M. Anderson, E. M. Linde, D. J. DiMaio, A. J. Lazenby, H. H. Wandall, et al. 2013. Aberrant expression of mucin core proteins and o-linked glycans associated with progression of pancreatic cancer. *Clin. Cancer Res.* 19:1981–1993.
47. Komuta, M., O. Govaere, V. Vandecaveye, J. Akiba, W. Van Steenberghe, C. Verslype, et al. 2012. Histological diversity in cholangiocellular carcinoma reflects the different cholangiocyte phenotypes. *Hepatology* 55:1876–1888.
48. Chaika, N. V., T. Gebregiorgis, M. E. Lewallen, V. Purohit, P. Radhakrishnan, X. Liu, et al. 2012. MUC1 mucin stabilizes and activates hypoxia-inducible factor 1 alpha to regulate metabolism in pancreatic cancer. *Proc. Natl Acad. Sci. USA* 109:13787–13792.
49. Skrypek, N., B. Duchêne, M. Hebbbar, E. Leteurtre, I. van Seuning, and N. Jonckheere. 2013. The MUC4 mucin mediates gemcitabine resistance of human pancreatic cancer cells via the Concentrative Nucleoside Transporter family. *Oncogene* 32:1714–1723.
50. Norris, A. M., A. Gore, A. Balboni, A. Young, D. S. Longnecker, and M. Korc. 2013. AGR2 is a SMAD4-suppressible gene that modulates MUC1 levels and promotes the initiation and progression of pancreatic intraepithelial neoplasia. *Oncogene* 32:3867–3876.
51. Tamada, S., H. Shibahara, M. Higashi, M. Goto, S. K. Batra, K. Imai, et al. 2006. MUC4 is a novel prognostic factor of extrahepatic bile duct carcinoma. *Clin. Cancer Res.* 12:4257–4264.
52. Sato, Y., K. Harada, M. Sasaki, and Y. Nakanuma. 2014. Histological characterization of biliary intraepithelial neoplasia with respect to pancreatic intraepithelial neoplasia. *Int. J. Hepatol.* 2014:678260.
53. Swanson, B. J., K. M. McDermott, P. K. Singh, J. P. Eggers, P. R. Crocker, and M. A. Hollingsworth. 2007. MUC1 is a counter-receptor for myelin-associated

- glycoprotein (Siglec-4a) and their interaction contributes to adhesion in pancreatic cancer perineural invasion. *Cancer Res.* 67:10222–10229.
54. Jepson, S., M. Komatsu, B. Haq, M. E. Arango, D. Huang, C. A. Carraway, et al. 2002. Muc4/sialomucin complex, the intramembrane ErbB2 ligand, induces specific phosphorylation of ErbB2 and enhances expression of p27(kip), but does not activate mitogen-activated kinase or protein kinaseB/Akt pathways. *Oncogene* 21:7524–7532.
  55. Yeh, C. N., S. T. Pang, R. C. Wu, T. W. Chen, Y. Y. Jan, and M. F. Chen. 2009. Prognostic value of MUC4 for mass-forming intrahepatic cholangiocarcinoma after hepatectomy. *Oncol. Rep.* 21:49–56.
  56. Saitou, M., M. Goto, M. Horinouchi, S. Tamada, K. Nagata, T. Hamada, et al. 2005. MUC4 expression is a novel prognostic factor in patients with invasive ductal carcinoma of the pancreas. *J. Clin. Pathol.* 58:845–852.
  57. Kitazono, I., M. Higashi, S. Kitamoto, S. Yokoyama, M. Horinouchi, M. Osako, et al. 2013. Expression of MUC4 mucin is observed mainly in the intestinal type of intraductal papillary mucinous neoplasm of the pancreas. *Pancreas* 42:1120–1128.
  58. Mosnier, J. F., C. Kandel, D. Cazals-Hatem, C. Bou-Hanna, J. Gournay, A. Jarry, et al. 2009. N-cadherin serves as diagnostic biomarker in intrahepatic and perihilar cholangiocarcinomas. *Mod. Pathol.* 22:182–190.
  59. Mao, X., D. Chen, J. Wu, J. Li, H. Zhou, Y. Wu, et al. 2013. Differential expression of fascin, E-cadherin and vimentin: proteins associated with survival of cholangiocarcinoma patients. *Am. J. Med. Sci.* 346:261–268.
  60. Nitta, T., T. Mitsunashi, Y. Hatanaka, M. Miyamoto, K. Oba, T. Tsuchikawa, et al. 2014. Prognostic significance of epithelial-mesenchymal transition-related markers in extrahepatic cholangiocarcinoma: comprehensive immunohistochemical study using a tissue microarray. *Br. J. Cancer* 111:1363–1372.
  61. Araki, K., T. Shimura, H. Suzuki, S. Tsutsumi, W. Wada, T. Yajima, et al. 2011. E/N-cadherin switch mediates cancer progression via TGF- $\beta$ -induced epithelial-to-mesenchymal transition in extrahepatic cholangiocarcinoma. *Br. J. Cancer* 105:1885–1893.
  62. Yao, X., X. Wang, Z. Wang, L. Dai, G. Zhang, Q. Yan, et al. 2012. Clinicopathological and prognostic significance of epithelial mesenchymal transition-related protein expression in intrahepatic cholangiocarcinoma. *Onco. Targets Ther.* 5:255–261.
  63. Hooper, J. E., T. K. Morgan, M. Grompe, B. C. Sheppard, M. L. Troxell, C. L. Corless, et al. 2012. The novel monoclonal antibody HPC2 and N-cadherin distinguish pancreatic ductal adenocarcinoma from cholangiocarcinoma. *Hum. Pathol.* 43:1583–1589.
  64. Ali, A., V. Brown, S. Denley, N. B. Jamieson, J. P. Morton, C. Nixon, et al. 2014. Expression of KOC, S100P, mesothelin and MUC1 in pancreatico-biliary adenocarcinomas: development and utility of a potential diagnostic immunohistochemistry panel. *BMC Clin. Pathol.* 14:35.
  65. Gandou, C., K. Harada, Y. Sato, S. Igarashi, M. Sasaki, H. Ikeda, et al. 2013. Hilar cholangiocarcinoma and pancreatic ductal adenocarcinoma share similar histopathologies, immunophenotypes, and development-related molecules. *Hum. Pathol.* 44:811–821.
  66. Tsai, J. H., W. C. Huang, K. T. Kuo, R. H. Yuan, Y. L. Chen, and Y. M. Jeng. 2012. S100P immunostaining identifies a subset of peripheral-type intrahepatic cholangiocarcinomas with morphological and molecular features similar to those of perihilar and extrahepatic cholangiocarcinomas. *Histopathology* 61:1106–1116.
  67. Canto, M. I., R. H. Hruban, E. K. Fishman, I. R. Kamel, R. Schulick, Z. Zhang, et al. 2012. Frequent detection of pancreatic lesions in asymptomatic high-risk individuals. *Gastroenterology* 142:796–804.
  68. Meister, T., H. S. Heinzow, C. Woestmeyer, P. Lenz, J. Menzel, T. Kucharzik, et al. 2013. Intraductal ultrasound substantiates diagnostics of bile duct strictures of uncertain etiology. *World J. Gastroenterol.* 19:874–881.
  69. Fujita, N., Y. Noda, G. Kobayashi, K. Ito, J. Horaguchi, S. Koshita, et al. 2009. Intraductal ultrasonography (IDUS) for the diagnosis of biliopancreatic diseases. *Best Pract. Res. Clin. Gastroenterol.* 23:729–742.
  70. Domagk, D., P. Lenz, and L. Menzel. 2013. Impact of intraductal ultrasound (miniprobe-endoscopic ultrasound) in diagnosing and staging of pancreatobiliary tumors. *Video J. Encyclopedia GI Endoscopy* 1:491–493.
  71. Bahra, M., J. M. Langrehr, and P. Neuhaus. 2006. Carcinomas of the distal bile duct. *Chirurg* 77:335–340.
  72. Witkowski, E. R., J. K. Smith, and J. F. Tseng. 2013. Outcomes following resection of pancreatic cancer. *J. Surg. Oncol.* 107:97–103.
  73. Neuhaus, P., A. Thelen, S. Jonas, G. Puhl, T. Denecke, W. Veltzke-Schlieker, et al. 2012. Oncological superiority of hilar en bloc resection for the treatment of hilar cholangiocarcinoma. *Ann. Surg. Oncol.* 19:1602–1608.
  74. Song, J., H. Liu, Z. Li, C. Yang, Y. Sun, and C. Wang. 2015. Long-term prognosis of surgical treatment for early ampullary cancers and implications for local ampullectomy. *BMC Surg.* 22:15–32.
  75. Wakai, T., Y. Shirai, Y. Tsuchiya, T. Nomura, K. Akazawa, and K. Hatakeyama. 2008. Combined major hepatectomy and pancreatoduodenectomy for locally advanced biliary carcinoma: long-term results. *World J. Surg.* 32:1067–1074.
  76. Neoptolemos, J. P., D. Cunningham, H. Friess, C. Bassi, D. D. Stocken, D. M. Tait, et al. 2003. Adjuvant

- therapy in pancreatic cancer: historical and current perspectives. *Ann. Oncol.* 14:675–692.
77. Hejna, M., M. Pruckmayer, and M. Raderer. 1998. The role of chemotherapy and radiation in the management of biliary cancer: a review of the literature. *Eur. J. Cancer* 34:977–986.
  78. Valle, J., H. Wasan, D. H. Palmer, D. Cunningham, A. Anthony, A. Maraveyas, et al. 2010. Cisplatin plus gemcitabine versus gemcitabine for biliary tract cancer. *N. Engl. J. Med.* 362:1273–1281.
  79. Yeh, C. N., Y. Y. Jan, T. S. Yeh, T. L. Hwang, and M. F. Chen. 2004. Hepatic resection of the intraductal papillary type of peripheral cholangiocarcinoma. *Ann. Surg. Oncol.* 11:606–611.
  80. Yoshida, T., T. Matsumoto, A. Sasaki, Y. Morii, M. Aramaki, and S. Kitano. 2002. Prognostic factors after pancreatoduodenectomy with extended lymphadenectomy for distal bile duct cancer. *Arch. Surg.* 137:69–73.
  81. Hansen, R., E. Quebbeman, P. Ritch, C. Chitambar, and T. Anderson. 1988. Continuous 5-fluorouracil (5FU) infusion in carcinoma of the pancreas: a phase II study. *Am. J. Med. Sci.* 295:91–93.
  82. Eckmann, K. R., D. K. Patel, A. Landgraf, J. H. Slade, E. Lin, H. Kaur, et al. 2011. Chemotherapy outcomes for the treatment of unresectable intrahepatic and hilar cholangiocarcinoma: a retrospective analysis. *Gastrointest. Cancer Res.* 4:155–160.
  83. Nehls, O., H. Oettle, J. T. Hartmann, R. D. Hofheinz, H. G. Hass, M. S. Horgner, et al. 2008. Capecitabine plus oxaliplatin as first-line treatment in patients with advanced biliary system adenocarcinoma: a prospective multicentre phase II trial. *Br. J. Cancer* 98:309–315.
  84. Boeck, S., T. Hoehler, G. Seipelt, R. Mahlberg, A. Wein, A. Hochhaus, et al. 2008. Capecitabine plus oxaliplatin versus capecitabine plus gemcitabine versus gemcitabine plus oxaliplatin: final results of a multicenter randomized phase II trial in advanced pancreatic cancer. *Ann. Oncol.* 19:340–347.
  85. Murakami, Y., K. Uemura, T. Sudo, Y. Hayashidani, Y. Hashimoto, H. Nakamura, et al. 2009. Gemcitabine-based adjuvant chemotherapy improves survival after aggressive surgery for hilar cholangiocarcinoma. *J. Gastrointest. Surg.* 13:1470–1479.
  86. Inaba, Y., Y. Arai, H. Yamaura, Y. Sato, M. Najima, T. Aramaki, et al. 2011. Phase I/II study of hepatic arterial infusion chemotherapy with gemcitabine in patients with unresectable intrahepatic cholangiocarcinoma (JIVROSG-0301). *Am. J. Clin. Oncol.* 34:58–62.
  87. Park, J.-S., S.-Y. Oh, S.-H. Kim, H. C. Kwon, J. S. Kim, H. Jin-Kim, et al. 2005. Single-agent gemcitabine in the treatment of advanced biliary tract cancers: a Phase II study. *Jpn. J. Clin. Oncol.* 35:68–73.
  88. Heinemann, V. 2002. Gemcitabine in the treatment of advanced pancreatic cancer: a comparative analysis of randomized trials. *Semin. Oncol.* 29:9–16.
  89. Tempero, M., W. Plunkett, V. R. Haperen, J. Hainsworth, H. Hochster, R. Lenzi, et al. 2003. Randomized Phase II comparison of dose-intense gemcitabine: thirty-minute infusion and fixed dose rate infusion in patients with pancreatic adenocarcinoma. *J. Clin. Oncol.* 21:3402–3408.
  90. Burris, H. A., M. J. Moore, J. Andersen, M. R. Green, M. L. Rothenberg, M. R. Modiano, et al. 1997. Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. *J. Clin. Oncol.* 15:2403–2413.
  91. Lee, M. A., I. S. Woo, J.-H. Kang, Y. S. Hong, and K. S. Lee. 2004. Gemcitabine and cisplatin combination chemotherapy in intrahepatic cholangiocarcinoma as second-line treatment: report of four cases. *Jpn. J. Clin. Oncol.* 34:547–550.
  92. Heinemann, V., H. Wilke, H.-G. Mergenthaler, M. Clemens, H. König, H. J. Illiger, et al. 2000. Gemcitabine and cisplatin in the treatment of advanced or metastatic pancreatic cancer. *Ann. Oncol.* 11:1399–1403.
  93. Hoff, D. D. V., R. K. Ramanathan, M. J. Borad, D. A. Laheru, L. S. Smith, T. E. Wood, et al. 2011. Gemcitabine plus nab-paclitaxel is an active regimen in patients with advanced pancreatic cancer: a phase I/II trial. *J. Clin. Oncol.* 29:4548–4554.
  94. Oettle, H., P. Neuhaus, A. Hochhaus, J. T. Hartmann, K. Gellert, K. Ridwelski, et al. 2013. Adjuvant chemotherapy with gemcitabine and long-term outcomes among patients with resected pancreatic cancer: the CONKO-001 randomized trial. *JAMA* 310:1473–1481.
  95. Sinn, M., J. K. Striefler, B. V. Sinn, D. Sallmon, S. Bischoff, J. M. Stieler, et al. 2013. Does long-term survival in patients with pancreatic cancer really exist? Results from the CONKO-001 study. *J. Surg. Oncol.* 108:398–402.
  96. Von Hoff, D. D., T. Ervin, F. P. Arena, E. G. Chiorean, J. Infante, M. Moore, et al. 2013. Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. *N. Engl. J. Med.* 369:1691–1703.
  97. Kolinsky, M. P., M. B. Saywer, and J. L. Spratlin. 2014. A case series of patients with pancreatic cancer and cholangiocarcinoma treated with nab-paclitaxel at a single institution. *J. Cancer Ther.* 05:605–610.