

Medizinische Fakultät der Charité – Universitätsmedizin Berlin
Campus Benjamin Franklin
aus dem Institut für Virologie
Direktorin: Univ.-Prof. Dr. Regine Heilbronn

Dissertation

Die Rolle des Helicase-Primase-Komplexes von Herpes-simplex-Virus Typ 1 bei der DNA-Replikation des adenoassoziierten Virus

Inaugural-Dissertation
zur Erlangung der medizinischen Doktorwürde
der Charité – Universitätsmedizin Berlin
Campus Benjamin Franklin

von
Heiko Slanina
aus Witten

Referent: Univ.-Prof. Dr. Regine Heilbronn
Korreferent: Univ.-Prof. Dr. Christian Hagemeier

Gedruckt mit Genehmigung der Charité – Universitätsmedizin Berlin
Campus Benjamin Franklin

Promoviert am: 22. Mai 2007

Meiner Familie

INHALTSVERZEICHNIS**ABKÜRZUNGSVERZEICHNIS** **IX****1 EINLEITUNG** **1**

1.1 DAS ADENOASSOZIIERTE VIRUS – ERFOLG VERSPRECHENDES VEKTORSYSTEM FÜR DIE GENTHERAPIE	1
1.2 DAS ADENOASSOZIIERTE VIRUS (AAV)	3
1.2.1 AAV – VORKOMMEN UND RISIKOBEWERTUNG	3
1.2.2 DAS GENOM – STRUKTUR, TRANSKRIPTION UND TRANSLATION	4
1.2.2.1 GENOMAUFBAU	4
1.2.2.2 TRANSKRIPTION UND TRANSLATION	5
1.2.3 LATENTER UND PRODUKTIVER LEBENSZYKLUS VON AAV	6
1.2.3.1 ADSORPTION UND PENETRATION	7
1.2.3.2 LATENTE INFektION VON AAV	7
1.2.3.3 PRODUktIVE INFektION VON AAV	8
1.2.3.4 DNA-REPLIKATION VON AAV	8
1.2.3.5 VIRALE UND ZELLULÄRE HELFERFUNKTIONEN	10
1.2.4 HERSTELLUNG VON AAV-VEKTOREN	11
1.3 DAS HERPES-SIMPLEX-VIRUS TYP 1 (HSV-1) - HELFERVIRUS FÜR AAV	12
1.3.1 AUFBAU VON HSV-1	12
1.3.2 LEBENSZYKLUS VON HSV-1	13
1.3.2.1 PRODUktIV-LYTISCHER REPLIKATIONSZYKLUS VON HSV-1	13
1.3.2.2 DER HELICASE-PRIMASE-KOMPLEX	14
1.3.2.3 ENTSTEHUNG VON HSV-REPLIKATIONSKOMPARTIMENTEN	15
1.3.2.4 DIE HSV-DNA-REPLIKATION	17
1.3.3 HSV-1-HELFERFUNKTIONEN FÜR DIE AAV-REPLIKATION	18
1.4 ZIELSETZUNG	20

2 MATERIAlien UND GERÄTE	21
2.1 GERÄTE	21
2.2 CHEMIKALIEN UND REAGENZIEN	21
2.3 KITS	22
2.4 ANTIKÖRPER	23
2.5 OLIGONUKLEOTIDE	24
2.6 PLASMIDE	25
2.7 BAKTERIENSTÄMME	26
2.8 ZELLINIE	27
2.9 VIREN	27
2.10 COMPUTERSOFTWARE	27
3 METHODEN	28
3.1 MOLEKULARBIOLOGISCHE METHODEN	28
3.1.1 HÄUFIG VERWENDETE LÖSUNGEN UND PUFFER	28
3.1.2 HERSTELLUNG KOMPETENTER ZELLEN	29
3.1.3 TRANSFORMATION CHEMISCH- UND ELEKTROKOMPETENTER ZELLEN	29
3.1.4 AUSPLATTIEREN VON BAKTERIEN	30
3.1.5 MINIPRÄPARATIONEN VON PLASMID-DNA DURCH KOCHLYSE	30
3.1.6 PRÄPARATION VON HIGH UND LOW COPY PLASMIDS	31
3.1.7 KONZENTRATIONSBESTIMMUNG VON DNA	32
3.1.8 PHENOL-CHLOROFORM-EXTRAKTION VON DNA	32
3.1.9 ETHANOLFÄLLUNG VON DNA	32
3.1.10 RESTRIKTIONSVERDAU VON DNA	33
3.1.11 HORIZONTALE AGAROSEGELEKTROPHORESE ZUR AUFTRENNUNG VON DNA-FRAGMENTEN	33
3.1.12 ISOLIERUNG VON DNA-FRAGMENTEN AUS AGAROSEGELEN NACH DER FREEZE AND SQUEEZE-METHODE	35
3.1.13 LIGATION VON DNA	35

3.1.14 PCR-MUTAGENESE	36
3.1.14.1 SITE-DIRECTED MUTAGENESIS	36
3.1.14.2 QUIKCHANGE SITE-DIRECTED MUTAGENESIS KIT	37
3.1.15 SEQUENZIERUNG	38
3.1.16 SOUTHERN BLOT-ANALYSE	38
3.2 ZELLKULTURTECHNIK	42
3.2.1 FÜR DIE ZELLKULTUR VERWENDETE LÖSUNGEN, PUFFER UND MEDIEN	42
3.2.2 HALTUNG, PASSAGIEREN UND AUSSÄEN VON HE LA-ZELLEN	43
3.2.3 CALCIUM-PHOSPHAT-KOTRANSFEKTION NACH CHEN UND OKAYAMA	43
3.2.4 LIPOFEKTION	44
3.2.5 INFektION VON ZELLEN	45
3.2.6 TRANSFEKTION MIT ANSCHLIEßENDER INFektION	45
3.2.7 EXTRAKTION GENOMISCHER DNA	46
3.2.8 HIRT-EXTRAKTION (NACH HIRT, 1976)	47
3.2.9 PROTEINEXTRAKTION	48
3.3 PROTEINBIOCHEMISCHE METHODEN	49
3.3.1 DISKONTINUIERLICHE SDS-POLYACRYLAMIDGELEKTROPHORESE ZUR AUFTRENNUNG VON PROTEINEN (SDS-PAGE)	49
3.3.2 WESTERN BLOT MIT IMMUNDETEKTION	51
3.3.3 IMMUNFLUORESZENZ	53
4 ERGEBNISSE	55
4.1 VORBEREITENDE EXPERIMENTE	55
4.1.1 UNTERSUCHUNG DER FÜR DIE AAV-DNA-REPLIKATION MINIMAL ERFORDERLICHEN HSV-HELPERGENE	55
4.1.2 CHARAKTERISIERUNG VON MONOKLONALEN UL5- UND UL52-ANTIKÖRPERN	56
4.1.3 ANALYSE DER BENÖTIGTEN HSV-HELPERGENE FÜR EINE KOLONALISATION VON REP MIT ICP8	58
4.1.3.1 CHARAKTERISIERUNG DER IN DER IMMUNFLUORESZENZ BENÖTIGTEN ANTI- KÖRPER GEGEN ICP8 UND REP	58

4.1.3.2 MINIMAL BENÖTIGTE KOMPONENTEN FÜR DIE KOLOKALISATION VON REP MIT ICP8	60
4.1.4 ZUSAMMENFASSUNG DER BISHERIGEN DATEN	61
4.2 UNTERSUCHUNG DER HELFERFUNKTION DER HSV-1-PRIMASE UL52	62
4.2.1 MUTAGENESE VON UL52	62
4.2.2 FUNKTIONELLE CHARAKTERISIERUNG DER UL52-PUNKTMUTANTEN IM HSV- REPLIKATIONSASSAY	63
4.2.3 EINFLUSS DER PUNKTMUTATIONEN IN DER PRIMASE AUF DIE KOLOKALISATIONS- FÄHIGKEIT VON ICP8 MIT REP	64
4.2.4 EINFLUSS DER HSV-UL52-PRIMASEAKTIVITÄT AUF DIE AAV-DNA-REPLIKATION NACH HSV-INFektion	66
4.2.5 EINFLUSS DER HSV-UL52-PRIMASEAKTIVITÄT AUF DIE AAV-DNA-REPLIKATION NACH TRANSFEKTION DER MINIMALEN HELFERFUNKTIONEN	67
4.2.6 VERGLEICH DER AAV-DNA-REPLIKATIONSRATE NACH INFektION MIT HSV UND NACH TRANSFEKTION DES MINIMALSATZES VON HSV-HELFERFUNKTIONEN	69
4.2.7 ZUSAMMENFASSUNG DER DATEN ZUR BEDEUTUNG DER PRIMASEAKTIVITÄT FÜR DIE AAV-DNA-REPLIKATION	70
4.3 ANALYSE DER HELFERFUNKTION DER HSV-1-HELICASE UL5	72
4.3.1 MUTAGENESE VON UL5	72
4.3.2 FUNKTIONELLE CHARAKTERISIERUNG DER HSV-HELICASEMUTANTEN IM HSV- REPLIKATIONSASSAY	73
4.3.3 UNTERSUCHUNG DER KOLOKALISATION ZWISCHEN REP UND ICP8 BEI VERWEN- DUNG DER HELICASE-PUNKTMUTANTEN	74
4.3.4 ANALYSE DES EINFLUSSES DER HSV-UL5-PUNKTMUTATIONEN AUF DIE AAV-DNA-REPLIKATION NACH HSV-INFektion	76
4.3.5 EINFLUSS DER HELICASE-PUNKTMUTATIONEN AUF DIE AAV-DNA-REPLIKATION NACH TRANSFEKTION MIT DEN MINIMALEN HSV-HELFERFUNKTIONEN	77
4.3.6 QUANTITATIVER VERGLEICH DER AAV-DNA-REPLIKATION ZWISCHEN DEN BEIDEN UNTERSUCHUNGSSYSTEMEN	79
4.3.7 ZUSAMMENFASSUNG DER ERGEBNISSE AUS DEN EXPERIMENTEN MIT DEN HELICASE-PUNKTMUTANTEN	80

5 DISKUSSION	81
5.1 BEDEUTUNG DER HSV-REPLIKATIONSKOMPARTIMENTE FÜR AAV	82
5.2 EINFLUSS DER HSV-PRIMASE UL52 AUF DIE AAV-DNA-REPLIKATION	85
5.2.1 MÖGLICHE FUNKTION DER HSV-PRIMASEAKTIVITÄT BEI DER AAV-DNA-REPLIKATION	87
5.2.2 MÖGLICHER EINFLUSS VIRALER UND ZELLULÄRER FUNKTIONEN IM TRANSFEKTIONS- UND INFektionsassay AUF DIE AAV-DNA-REPLIKATION	89
5.3 DIE ROLLE DER HSV-HELICASE UL5 BEI DER AAV-DNA-REPLIKATION	92
5.3.1 ERKLÄRUNGSMODELLE FÜR DEN EINFLUSS DER HSV-HELICASE AUF DIE AAV-DNA-REPLIKATION	93
5.3.1.1 STRUKTURELLE BEDEUTUNG DER HELICASE FÜR DIE AUSBILDUNG VON REPLIKATIONSKOMPLEXEN	94
5.3.1.2 DIE HSV-HELICASE UND IHRE BEDEUTUNG BEI REKOMBINATIONSEREIGNISSEN	94
5.3.1.3 MODELLE FÜR DIE FUNKTION DER UL5-HELICASE BEI DER AAV-DNA-REPLIKATION	95
6 LITERATURVERZEICHNIS	98
7 ZUSAMMENFASSUNG	110
8 ANHANG	112
8.1 MONOCLONAL ANTIBODIES AGAINST UL5	112
8.2 VERÖFFENTLICHUNGEN	113
8.3 SELBSTSTÄNDIGKEITSERKLÄRUNG	114
8.4 DANKSAGUNGEN	115
8.5 LEBENSLAUF	116

Abkürzungsverzeichnis

Nicht aufgenommen sind die Abkürzungen für chemische Elemente und internationale Standardeinheiten (SI-Einheiten).

A	Alanin
AAV, rAAV	(rekombinantes) adenoassoziiertes Virus
Abb.	Abbildung
bidest.	zweifach destilliertes Wasser
bp, kbp	Basenpaar, Kilobasenpaare
bzw.	beziehungsweise
Ci	Curie
CMV	Zytomegalievirus
D	Asparaginsäure
Da	Dalton
d.h.	das heißt
DNA, ssDNA	(einzelsträngige) Desoxyribonukleinsäure
EDTA	Ethyldiamintetraessigsäure
FCS	fetales Kälberserum
FITC	Fluoresceinthionylcarbonat
G	Glycin
GTP	Guanosin-5'-triphosphat
h	Stunde
HSV, rHSV	(rekombinantes) Herpes-simplex-Virus
ICP	<i>infected cell protein</i>
ITR	<i>inverted terminal repeat</i>
K	Lysin
mAk	monoklonaler Antikörper
min	Minuten
MOI	Multiplizität der Infektion
ND10	<i>nuclear domain 10, promyelotic leukemia nuclear body</i>
NTP, dNTP	(Desoxy)nukleosidtriphosphat

ORF	offenes Leseraster
<i>ori</i>	<i>origin of replication</i>
Page	Polyacrylamidgelektrophorese
PBS, PBS-T	phosphatgepufferte Salzlösung, mit Tween®
pCM	Plasmid unter CMV-Promotorkontrolle
PCR	Polymerase-Kettenreaktion
pfu	<i>plaque forming units</i>
PML	<i>promyelocytic leukemia protein</i>
Q	Glutamin
R	Arginin
RBE	<i>rep-binding element</i>
RF1, RF2	Replikationsform 1, Replikationsform 2 von AAV
RNA, mRNA	(<i>messenger</i> -)Ribonukleinsäure
s.	siehe
SDS	Natriumdodekylsulfat
Tab.	Tabelle
TNF	<i>tumor necrosis factor</i>
Tris	Tris(hydroxymethyl)-Aminomethan
TRITC	Tetramethylrhodaminisothiocyanat
trs	<i>terminal resolution site</i>
u.a.	unter anderem
U _L , UL	<i>unique long</i> von HSV
V	Valin
v.a.	vor allem
vgl.	vergleiche
% [v/v]	Volumenprozent
% [w/v]	Gewichtsprozent
wt	Wildtyp
z.B.	zum Beispiel

6 LITERATURVERZEICHNIS

- Archetti, I., and Bocciarelli, D. S. (1965). Structure and biological characteristics of a small, still unclassified virus. *Ann. Ist. Super Sanita* **1**, 103-106.
- Argos, P. (1988). A sequence motif in many polymerases. *Nuc. Acids Res.* **16**, 9909-9916.
- Asubel, F. M., Brent, R., Kingston, R. E., Moore, D. D., Seidmann, J. G., Smith, J. A., and Struhl, K. (1987). "Current protocols in molecular biology." John Wiley & Sons.
- Atchinson, R. W., Casto, B. C., and Hammon, W. M. C. D. (1965). Adenovirus-associated defective virus particles. *Science* **164**, 754-756.
- Bartlett, J. S., Wilcher, R., and Samulski, R. J. (2000) Infectious entry pathway of adeno-associated virus and adeno-associated virus vectors. *J. Virol.* **74**, 2777-2785.
- Batchu, R. B., Kotin, R. M., and Hermonat, P. L. (1994). The regulatory *rep* protein of adeno-associated virus binds to sequences within the c-H-ras promoter. *Cancer Letters* **86**, 23-31.
- Berns, K. I., and Linden, R. M. (1995). The cryptic life style of adeno-associated virus. *BioEssays* **17**, 237-245.
- Betz, U. A., Fischer, R., Kleymann, G., Hendrix, M., and Rubsam-Waigmann, H. (2002). Potent in vivo antiviral activity of the herpes simplex virus primase-helicase inhibitor BAY 57-1293. *Antimicrob. Agents Chemother.* **46**, 1766-1772.
- Biswas, N., and Weller, S. K. (1999). A mutation in the c-terminal putative Zn²⁺ finger motif of UL52 severely affects the biochemical activities of the HSV-1 helicase-primase subcomplex. *J. Biol. Chem.* **274**, 8068-8076.
- Biswas, N., and Weller S. K. (2001). The UL5 and UL52 subunits of the herpes simplex virus type 1 helicase-primase subcomplex exhibit a complex interdependence for DNA binding. *J. Biol. Chem.* **276**, 17610-17619.
- Blacklow, N. R., Hoggan, M. D., Kapikian, A. Z., Austin, J. B., and Rowe, W. P. (1968a). Epidemiology of adenovirus-associated virus infection in a nursery population. *Am. J. Epidemiol.* **88**, 368-378.
- Blacklow, N. R., Hoggan, M. D., and Rowe, W. P. (1968b). Serologic evidence for human infection with adenovirus-associated viruses. *J. Natl. Cancer Inst.* **40**, 319-327.
- Blumel, J., and Matz, J. (1995). Thermosensitive UL9 gene function is required for early stages of herpes simplex virus type 1 DNA synthesis. *J. Gen. Virol.* **76**, 3119-3124.
- Boehmer, P. E., and Lehman, I. R. (1993a). Herpes simplex virus type 1 ICP8: helix-destabilizing properties. *J. Virol.* **67**, 711-715.
- Boehmer, P. E., Dodson, M. S., and Lehman, I. R. (1993b). The herpes simplex virus type-1 origin binding protein. *J. Biol. Chem.* **268**, 1213-1219.
- Boehmer, P. E., and Lehman, I. R. (1997). Herpes simplex virus DNA replication. *Annuv. Rev. Biochem.* **66**, 347-384.
- Boehmer, P. E. (1998). The herpes simplex virus type-1 single-strand DNA-binding protein, ICP8, increases the processivity of the UL9 protein DNA helicase. *J. Biol. Chem.* **273**, 2676-2683.
- Borden, K. L. (2002). Pondering the promyelocytic leukaemia protein (PML) puzzle: possible functions for PML nuclear bodies. *Mol. Cell. Biol.* **22**, 5259-5269.
- Boyer, H. W., and Roulland-Dussoix, D. (1969). A complementation analysis of restriction and modification of DNA in *Escherichia coli*. *J. Mol. Biol.* **41**, 459-472.
- Brister, J. R., and Muzyczka, N. (1999): Rep-mediated nicking of the adeno-associated virus origin requires two biochemical activities, DNA helicase activity and transesterification. *J. Virol.* **73**, 9325-9336.
- Buller, R. M., and Rose, J. A. (1978). Characterization of adeno-associated virus-induced polypeptides in KB cells. *J. Virol.* **25**, 331-338.
- Buller, R. M., Janik, J. E., Sebring, E. D., and Rose, J. A. (1981). Herpes simplex virus types 1 and 2 completely help adenovirus-associated virus replication. *J. Virol.* **40**, 241-247.

- Burger, C., Gorbatyuk, O. S., Velardo, M. J., Peden, C. S., Williams, P., Zolotukhin, S., Reier, P. J., Mandel, R. J., and Muzyczka, N. (2004). Recombinant AAV viral vectors pseudotyped with viral capsids from serotypes 1, 2, and 5 display differential efficiency and cell tropism after delivery to different regions of the central nervous system. *Mol. Therapy* **10**, 302-317.
- Burkham, J., Coen, D. M., and Weller, S. K. (1998). ND10 protein PML is recruited to herpes simplex virus type 1 prereplicative sites and replication compartments in the presence of viral DNA polymerase. *J. Virol.* **72**, 10100-10107.
- Burkham, J., Coen, D. M., Hwang, C. B. C., and Weller, S. K. (2001). Interactions of herpes simplex virus type 1 with ND10 and recruitment of PML to replication compartments. *J. Virol.* **75**, 2353-2367.
- Bush, M., Yager, D. R., Gao, M., Weisshart, K., Marcy, A. L., Coen, D. M., and Knipe, D. M. (1991). Correct intranuclear localization of herpes simplex virus DNA polymerase requires the viral ICP8 DNA-binding protein. *J. Virol.* **65**, 1082-1089.
- Cai, W., Astor, T. L., Liptak, L. M., Cho, C., Coen, D. M., and Schaffer, P. M. (1993). The herpes simplex virus type 1 regulatory protein ICP0 enhances virus replication during acute infection and reactivation from latency. *J. Virol.* **67**, 7501-7512.
- Calder, J. M., Stow, E. C., and Stow, N. D. (1992). On the cellular localization of the components of the herpes simplex virus type 1 helicase-primase complex and the viral origin-binding protein. *J. Gen. Virol.* **73**, 531-538.
- Campbell, M.E., Palfreyman, J.W., and Preston C.M. (1984). Identification of herpes simplex virus DNA sequences which encode a trans-acting polypeptide responsible for stimulation of immediate early transcription. *J. Mol. Biol.* **180**, 1-19.
- Carrington-Lawrence, S. D., and Weller, S. K. (2003). Recruitment of polymerase to herpes simplex virus type 1 replication foci in cells expressing mutant primase (UL52) proteins. *J. Virol.* **77**, 4237-4247.
- Carter, B. J., Antoni, B. A., and Klessig, D. F. (1992). Adenovirus containing a deletion of the early region 2A gene allows growth of adeno-associated virus with decreased efficiency. *Virology* **191**, 473-476.
- Carter, B. J. (2005). Adeno-associated virus vectors in clinical trials. *Hum. Gene Ther.* **16**, 541-550.
- Chakrabarti, R., and Schutt, C. E. (2001). The enhancement of PCR amplification by low molecular-weight sulfones. *Gene* **274**, 293-298.
- Chen, A., and Okayama, H. (1987). High-efficiency transformation of mammalian cells by plasmid DNA. *Mol. Cell. Biol.* **7**, 2745-2752.
- Chen, A., and Okayama, H. (1988). Calcium phosphate-mediated gene transfer: a highly efficient transfection system for stably transforming cells with plasmid DNA. *BioTechniques* **6**, 632-638.
- Chen, Y., Carrington-Lawrence, S. D., Bai, P., and Weller, S. K. (2005). Mutations in the putative zinc-binding motif of UL52 demonstrate a complex interdependence between the UL5 and UL52 subunits of the human herpes simplex virus type 1 helicase/primase complex. *J. Virol.* **79**, 9088-9096.
- Chiornini, J. A., Wiener, S. M., Owens, R. A., Kyöstiö, S. R. M., Kotin, R. M., and Safer, B. (1994). Sequence requirements for stable binding and function of Rep68 on the adeno-associated virus type 2 inverted terminal repeats. *J. Virol.* **68**, 7448-7457.
- Cocchi, F., Menotti, L., Dubreuil, P., Lopez, M., and Campadelli-Fiume, G. (2000). Cell-to-cell spread of wild-type herpes simplex virus type 1, but not of syncytial strains, is mediated by the immunoglobulin-like receptors that mediate virion entry, nectin 1 (PRR1/HveC/HlgR) and nectin 2 (PRR2/HveB). *J. Virol.* **74**, 3909-3917.
- Constantin, N., and Dodson, M. S. (1999). Two-hybrid analysis of the interaction between the UL52 and UL8 subunits of the herpes simplex virus type 1 helicase-primase. *J. Gen. Virol.* **80**, 2411-2415.
- Constanzo, F., Campadelli-Fiume, G., Foa-Tomasi, L., and Cassai, E. (1977). Evidence that herpes simplex virus DNA is transcribed by cellular RNA polymerase B. *J. Virol.* **21**, 996-1001.

- Conway, J., Rhys, C., Zolotukhin, I., Zolotukhin, S., Muzycka, N., Hayward, G., and Byrne, B. (1999). High-titer recombinant adeno-associated virus production utilizing a recombinant herpes simplex virus type I vector expressing AAV-2 rep and cap. *Gene Therapy* **6**, 986-993.
- Costello, E., Saudan, P., Winocour, E., Pizer, L., and Beard, P. (1997). High mobility group chromosomal protein 1 binds to the adeno-associated virus replication protein (Rep) and promotes Rep-mediated site-specific cleavage of DNA, ATPase activity and transcriptional repression. *EMBO J.* **16**, 5943-5954.
- Crute, J. J., Tsurumi, T., Zhu, L. A., Weller, S. K., Olivo, P. D., Challberg, M. D., Mocarski, E. S., and Lehman, I. R. (1989a). Herpes simplex virus 1 helicase-primase: a complex of three herpes-encoded gene products. *Proc. Natl. Acad. Sci. U.S.A.* **86**, 2186-2189.
- Crute, J. J., and Lehman, I. R. (1989b). Herpes simplex-1 DNA polymerase. Identification of an intrinsic 5'-3' exonuclease with ribonuclease H activity. *J. Biol. Chem.* **64**, 19266-19270.
- Davidson, B. L., Stein, C. S., Heth, J. A., Martins, I., Kotin, R. M., Derksen, T. A., Zabner, J., Ghodsi, A., and Chiorini, J. A. (2000). Recombinant adeno-associated virus type 2, 4 and 5 vectors: transduction of variant cell types and regions in the mammalian central nervous system. *Proc. Natl. Acad. Sci. U.S.A.* **97**, 3428-3432.
- Davis, M. D., Wu, J., and Owens, R. A. (2000). Mutational analysis of adeno-associated virus type 2 Rep68 protein endonuclease activity on partially single-stranded substrates. *J. Virol.* **74**, 2936-2942.
- de Bruyn Kops, A., and Knipe, D. M. (1988). Formation of DNA replication structures in herpes simplex virus-infected cells requires a viral DNA binding protein. *Cell* **55**, 857-868.
- de Bruyn Kops, A., and Knipe, D. M. (1994). Preexisting nuclear architecture defines the intranuclear location of herpesvirus DNA replication structures. *J. Virol.* **68**, 3512-2526.
- Delius H., and Clements, J. B. (1976). A partial denaturation map of herpes simplex virus type 1 DNA: evidence for inversions of the unique DNA regions. *J. Gen. Virol.* **33**, 125-133.
- Dodson, M. S., and Lehman, I. R. (1991). Association of DNA helicase and primase activities with a subassembly of the herpes simplex virus 1 helicase-primase composed of the UL5 and UL52 gene products. *Proc. Natl. Acad. Sci. U.S.A.* **88**, 1105-1109.
- Dracheva, S., Koonin, E. V., and Crute, J. J. (1995). Identification of the primase active site of the herpes simplex virus type 1 helicase-primase. *J. Biol. Chem.* **270**, 14148-14153.
- Dubielzig, R. King, J. A., Weger, S., Kern, A., and Kleinschmidt, J. A. (1999). Adeno-associated virus type 2 protein interactions: formation of pre-encapsidation complexes. *J. Virol.* **73**, 8989-8998.
- Ebert, S. N., Subramanian, D., Shtrom, S. S., Chung, I. K., Parris, D. S., and Muller, M. T. (1994). Association between the p170 form of human topoisomerase II and progeny viral DNA in cells infected with herpes simplex virus type 1. *J. Virol.* **68**, 1010-1020.
- Everett, R. D., Earnshaw, W. C., Findlay, J., and Lomonte, P. (1999). Specific destruction of kinetochore protein CENP-C and disruption of cell division by herpes simplex virus immediate-early protein Vmw110. *EMBO J.* **18**, 1526-1538.
- Everett, R. D., and Murray, J. (2005). ND10 components relocate to sites associated with herpes simplex virus Type 1 nucleoprotein complexes during virus Infection. *J. Virol.* **79**, 5078-5089.
- Everett, R. D. (2006a). Interactions between DNA viruses, ND10 and the DNA damage response. *Cell Microbiol.* **8**, 365-374.
- Everett, R. D., Rechter, S., Papier, P., Tavalai, N., Stamminger, T., and Orr, A. (2006b). PML contributes to a cellular mechanism of repression of herpes simplex virus type 1 infection that is inactivated by ICP0. *J. Virol.* **80**, 7995-8005.
- Falkenberg, M., Bushnell, D. A., Elias, P., and Lehman, I. R. (1997). The UL8 subunit of the heterotrimeric herpes simplex virus type 1 helicase-primase is required for the unwinding of single strand DNA-binding protein (ICP8)-coated DNA substrates. *J. Biol. Chem.* **272**, 22766-22770.

- Feinberg, A. P., and Vogelstein, B. (1983). A technique for radiolabeling DNA restriction endonuclease fragments to high specific activity. *Anal. Biochem.* **132**, 6-13.
- Fisher, K. J., Jooss, K., Alston, J., Yang, Y., Haecker, S. E., High, K., Pathak, R., Raper, S. E., and Wilson, J. M. (1997). Recombinant adeno-associated virus for muscle directed gene therapy. *Nature Medicine* **3**, 306-312.
- Flotte, T. R., Afione, S. A., and Zeitlin, P. L. (1994). Adeno-associated virus vector gene expression occurs in nondividing cells in the absence of vector DNA integration. *Am. J. Respir. Cell Mol. Biol.* **11**, 517-521.
- Frenkel, N., Locker, H., Batterson, W., Hayward, G.S. and Roizman, B. (1976). Anatomy of herpes simplex virus DNA. VI. Defective DNA originates from the S component. *J Virol* **20**, 527-531.
- Friedman-Einat, M., Grossman, Z., Mileguir, F., Smetana, Z., Ashkenazi, M., Barkai, G., Varsano, N., Glick, E., and Mendelson, E. (1997). Detection of adeno-associated virus type 2 sequences in human genital tract. *J. Clin. Microbiol.* **35**, 71-78.
- Gac, N. T. L., Villani, G., Hoffmann, J. S., and Boehmer, P. E. (1996). The UL8 subunit of the herpes simplex virus type-1 DNA helicase-primase optimizes utilization of DNA templates covered by the homologous single-strand DNA-binding protein ICP8. *J. Biol. Chem.* **271**, 21645-21651.
- Gao, G. P., Alvira, M. R., Wang, L., Calcedo, R., Johnston, J., and Wilson, J. M. (2002). Novel adeno-associated viruses from rhesus monkeys as vectors for human gene therapy. *Proc. Natl. Acad. Sci. U S A* **99**, 11854-11859.
- Gao, G., Vandenberghe, L.H., Alvira, M.R., Lu, Y., Calcedo, R., Zhou, X., and Wilson, J.M. (2004). Clades of Adeno-associated virus are widely disseminated in human tissues. *J. Virol.* **78**, 6381-6388.
- Garber, D. A., Beverley, S. M., and Coen, D. M. (1993). Demonstration of circularisation of herpes simplex virus DNA following infection using pulsed field gel electrophoresis. *Virology* **197**, 459-462.
- Ge, H., and Roeder, R. G. (1994a). Purification, cloning, and characterization of a human coactivator, PC4, that mediates transcriptional activation of class II genes. *Cell* **78**, 513-523.
- Ge, H., Zhao, Y., Chait, B. T., and Roeder, R. G. (1994b). Phosphorylation negatively regulates the function of coactivator PC4. *Proc. Natl. Acad. Sci. U S A* **91**, 12691-12695.
- Geoffroy, M.-C., Epstein, A. L., Toublanc, E., Moullier, P., and Salvetti, A. (2004). Herpes simplex virus type 1 ICP0 protein mediates activation of adeno-associated virus type 2 *rep* gene expression from a latent integrated form. *J. Virol.* **78**, 10977-10986.
- Georg-Fries, B., Biederlack, S., Wolf, J., and zur Hausen, H. (1984). Analysis of proteins, helper dependence, and seroepidemiology of a new human parvovirus. *Virology* **134**, 64-71.
- Geraghty, R. J., Krummenacher, C., Cohen, G. H., Eisenberg, G. J., and Spear, P. G. (1998). Entry of alphaherpesviruses mediated by poliovirus receptor-related protein 1 and poliovirus receptor. *Science* **280**, 1618-1620.
- Gey, G. O., Coffman, W. D., and Kubicek, M. T. (1952). Tissue culture studies of the proliferative capacity of cervical carcinoma and normal epithelium. *Cancer Res.* **12**, 264-265.
- Girod, A., Ried, M., Wobus, C., Lahm, H., Leike, K., Kleinschmidt, J., Deleage, G., and Hallek, M. (1999). Genetic capsid modifications allow efficient retargeting of adeno-associated virus type 2. *Nat. Med.* **5**, 1052-1056.
- Goldstein, D. J., and Weller, S. K. (1988). An ICP6::*lacZ* insertional mutagen is used to demonstrate that the UL52 gene of herpes simplex virus type 1 is required for virus growth and DNA synthesis. *J. Virol.* **62**, 2970-2977.
- Gorbalenya, A. E., Koonin, E. V., and Wolf, Y. I. (1990). A new superfamily of putative NTP-binding domains encoded by genomes of small DNA and RNA viruses. *FEBS Lett.* **262**, 145-148.
- Gottlieb, J., Marcy, A. I., Coen, D. M., and Challberg, D. M. (1990). The herpes simplex virus type 1 UL42 gene product: a subunit of DNA polymerase that functions to increase processivity. *J. Virol.* **64**, 5976-5987.

- Gourves, A. S., Gac, N. T. L., Villani, G., Boehmer, P. E., and Johnson, N. P. (2000). Equilibrium binding of single-stranded DNA with herpes simplex virus type I-coded single-stranded DNA-binding protein, ICP8. *J. Biol. Chem.* **275**, 10864-10869.
- Graves-Woodward, K. L., Gottlieb, J., Challberg, M. D., and Weller, S. K. (1997). Biochemical analyses of mutations in the HSV-1 helicase-primase that alter ATP hydrolysis, DNA unwinding, and coupling between hydrolysis and unwinding. *J. Biol. Chem.* **272**, 4623-4630.
- Grimm, D., Kern, A., Rittner, K., and Kleinschmidt J.A. (1998). Novel tools for production and purification of recombinant adenoassociated virus vectors. *Hum. Gene Ther.* **10**, 2745-2760.
- Grimm, D. (2002). Production methods for gene transfer vectors based on adeno-associated virus serotypes. *Methods* **28**, 146-157.
- Grossman, Z., Mendelson, E., Brok-Simoni, F., Mileguir, F., Leitner, Y., Rechavi, G., and Ramot, B. (1992). Detection of adeno-associated virus type 2 in human peripheral blood cells. *J. Gen. Virol.* **73**, 961-966.
- Hamatake, R. K., Bifano, M., Hurlburt, W. W., and Tenney, D. J. (1997). A functional interaction of ICP8, the herpes simplex virus single-stranded DNA-binding protein, and the helicase-primase complex that is dependent on the presence of the UL8 subunit. *J. Gen. Virol.* **78**, 857-865.
- Hauswirth, W. W., and Berns, K. I. (1977). Origin and termination of adeno-associated virus DNA replication. *Virology* **78**, 488-499.
- Hauswirth, W. W., and Berns, K. I. (1979). Adeno-associated virus DNA replication: nonunit-length molecules. *Virology* **93**, 57-68.
- Hayward, G. S., Jacob, R. J., Wadsworth, S. C., and Roizman, B. (1975). Anatomy of herpes simplex virus DNA: evidence for four populations of molecules that differ in the relative orientations of their long and short components. *Proc. Natl. Acad. Sci. U S A* **72**, 4243-4247.
- Heilbronn, R., and zur Hausen, H. (1989). A subset of herpes simplex virus replication genes induces DNA amplification within the host cell genome. *J. Virol.* **63**, 3683-3692.
- Heilbronn, R., Bürkle, A., Stephan, S., and zur Hausen, H. (1990). The adeno associated virus *rep* gene suppresses herpes simplex virus-induced DNA amplification. *J. Virol.* **64**, 3012-3018.
- Heilbronn, R., Engstler, M., Weger, S., Krahn, A., Schetter, C., and Boshart, M. (2003). ssDNA-dependent colocalization of adeno-associated virus Rep and herpes simplex virus ICP8 in nuclear replication domains. *Nuc. Acids Res.* **31**, 6206-6213.
- Hermonat, P. L., and Muzyczka, N. (1984a). Use of adeno-associated virus as mammalian DNA cloning vector: transduction of neomycin resistance into mammalian tissue culture cells. *Proc. Natl. Acad. Sci. U S A* **81**, 6466-6470.
- Hermonat, P. L., Labow, M. A., Wright, R., Berns, K. I., and Muzyczka, N. (1984b). Genetics of adeno-associated virus: isolation and preliminary characterization of adeno-associated virus type 2 mutants. *J. Virol.* **51**, 329-339.
- Hermonat, P. L. (1994a). Adeno-associated virus inhibits human papillomavirus type 16: a viral interaction implicated in cervical cancer. *Cancer Res.* **54**, 2278-2281.
- Hermonat, P. L. (1994b). Down-regulation of the human *c-fos* and *c-myc* proto-oncogene promoters by adeno-associated virus Rep78. *Cancer Lett.* **81**, 129-136.
- Hernandez, T. R., and Lehman, I. R. (1990). Functional interaction between the herpes simplex-1 DNA polymerase and UL42 protein. *J. Biol. Chem.* **265**, 11227-11232.
- Hernandez, Y. J., Wang, J., and Kearns, W. G. (1999). Latent adeno-associated virus infection elicits humoral but not cell-mediated immune responses in a nonhuman primate model. *J. Virol.* **73**, 8549-8558.
- Hickman, A. B., and Dyda, F. (2005). Binding and unwinding: SF3 viral helicases. *Cur. opinion in struct. Biol.* **15**, 77-85.
- Hirt, B. (1967). Selective extraction of polyoma DNA from infected mouse cell cultures. *J. Mol. Biol.* **26**, 365-369.

- Hoggan, M. D., Blacklow, N. R., and Rowe, W. P. (1966). Studies of a small DNA viruses found in various adenovirus preparations: physical, biological, and immunological characteristics. *Proc. Natl. Acad. Sci. U S A* **55**, 1467-1474.
- Honess, R. W., and Roizman, B. (1974). Regulation of herpesvirus macro-molecular synthesis. I. Cascade regulation of the synthesis of three groups of viral proteins. *J. Virol.* **14**, 8-9.
- Hüser, D., and Heilbronn, R. (2003). Adeno-associated virus integrates site-specifically into human chromosome 19 in either orientation and with equal kinetics and frequency. *J. Gen. Virol.* **84**, 133-137.
- Ilyina, T. V., Gorbaleyna, A. E., and Koonin, E. V. (1992). Organization and evolution of bacterial and bacteriophage primase-helicase systems. *J. Mol. Evol.* **34**, 351–357.
- Im, D.-S., and Muzyczka, N. (1990). The AAV origin-binding protein Rep68 is an ATP-dependent site-specific endonuclease with helicase activity. *Cell* **6**, 447-457.
- Janik, J. E., Huston, M. M., and Rose, J. A. (1981). Locations of adenovirus genes required for the replication of adenovirus-associated virus. *Proc. Natl. Acad. Sci. U S A* **78**, 1925-1929.
- Jooss, K., Yang, Y., Fisher, K. J., and Wilson, J. M. (1998). Transduction of dendritic cells by DNA viral vectors directs the immune response to transgene products in muscle fibers. *J. Virol.* **72**, 4212-4223.
- Kashiwakura, Y., Tamayose, K., Iwabuchi, K., Hirai, Y., Shimada, T., Matsumoto, K., Nakamura, T., Oshimi, K., and Daida, H. (2005). Hepatocyte growth factor receptor is a coreceptor for adeno-associated virus type 2 infection. *J. Virol.* **79**, 609-614.
- King, J. A., Dubielzig, R., Grimm, D., and Kleinschmidt, J. A. (2001). DNA helicase-mediated packaging of adeno-associated virus type 2 genomes into preformed capsids. *EMBO J.* **20**, 3282-3291.
- Kleymann, G., Fischer, R., Betz, U. A., Hendrix, M., Bender, W., Schneider, U., Handke, G., Eckenberg, P., Hewlett, G., Pevzner, V., Baumeister, J., Weber, O., Henninger, K., Keldenich, J., Jensen, A., Kolb, J., Bach, U., Popp, A., Maben, J., Frappa, I., Haebich, D., Lockhoff, O., and Rubsam-Waigmann H. (2002). New helicase-primase inhibitors as drug candidates for the treatment of herpes simplex disease. *Nat. Med.* **8**, 392–398.
- Klinedinst, D. K., and Challberg, M. D. (1994). Helicase-primase complex of herpes simplex virus type 1: a mutation in the UL52 subunit abolishes primase activity. *J. Virol.* **68**, 3693-3701.
- Knopf, K. W. (1979). Properties of herpes simplex virus DNA polymerase and characterization of its associated exonuclease activity. *Eur. J. Biochem.* **98**, 231-244.
- Korolev, S., Yao, N., Lohman, T. M., Weber, P. C., and Waksman, G. (1998). Comparisons between the structures of HCV and Rep helicases reveal structural similarities between SF1 and SF2 super-families of helicases. *Protein Sci.* **7**, 605-610.
- Kotin, R. M., Siniscalco, M., Samulski, R. J., Zhu, X. D., Hunter, L., Laughlin, C. A., McLaughlin, S., Muzyczka, N., Rocchi, M., and Berns, K. I. (1990). Site-specific integration by adeno-associated virus. *Proc. Natl. Acad. Sci. U S A* **87**, 2211-2215.
- Kotin, R. M., Menninger, J. C., Ward, D. C., and Berns, K. I. (1991). Mapping and direct visualization of a region-specific viral DNA integration site on chromosome 19q13-qter. *Genomics* **10**, 831-834.
- Kotin, R. M., Linden, R. M., and Berns, K. I. (1992). Characterization of a preferred site on human chromosome 19q for integration of adeno-associated virus DNA by non-homologous recombination. *EMBO J.* **11**, 5071-5078.
- Kyöstiö, S. R. M., Owens, R. A., Weitzman, M. D., Antoni, B. A., Chejanovsky, N., and Carter, B. J. (1994). Analysis of adeno-associated virus (AAV) wild-type and mutant Rep proteins for their abilities to negatively regulate AAV p5 and p19 mRNA levels. *J. Virol.* **68**, 2947-2957.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* **277**, 680-685.
- Lamberti, C., and Weller, S. K. (1998). The herpes simplex virus type 1 cleavage/packaging protein, UL32, is involved in efficient localization of capsids to replication compartments. *J. Virol.* **72**, 2463-2473.

- Laughlin, C. A., Cardellichio, C. B., and Coon, H. C. (1986). Latent infection of KB cells with adeno-associated virus type 2. *J. Virol.* **60**, 515-524.
- Lee, C. K., and Knipe, D. M. (1985). An immunoassay for the study of DNA-binding activities of herpes simplex virus protein ICP8. *J. Virol.* **54**, 731-738.
- Lee, S. S., and Lehman, I. R. (1997). Unwinding of the box I element of a herpes simplex virus type 1 origin by a complex of the viral origin binding protein, single-strand DNA binding protein, and single-stranded DNA. *Proc. Natl. Acad. Sci. U S A* **94**, 2383-2842.
- Liptak, L., Uprichard, S. L., and Knipe, D. M. (1996). Functional order of assembly of herpes simplex virus DNA replication proteins into prereplicative site structures. *J. Virol.* **70**, 1759-1767.
- Liuzzi, M., Kibler, P., Bousquet, C., Harji, F., Bolger, G., Garneau, M., Lapeyre, N., McCollum, R. S., Faucher, A. M., Simoneau, B., and Cordingley, M. G. (2004). Isolation and characterization of herpes simplex virus type 1 resistant to aminothiazolylphenyl-based inhibitors of the viral helicase-primase. *Antiviral Res.* **64**, 161-170.
- Locker, H., Frenkel, N. and Halliburton, I. (1982). Structure and expression of class II defective herpes simplex virus genomes encoding infected cell polypeptide number 8. *J Virol* **43**, 574-593.
- Lukonis, C. J., and Weller, S. K. (1996). Characterization of nuclear structures in cells infected with herpes simplex virus type 1 in the absence of viral DNA replication. *J. Virol.* **70**, 1751-1758.
- Lukonis, C. J., Weller, S. K. (1997). Formation of herpes simplex virus type 1 replication compartments by transfection: requirements and localization to nuclear domain 10. *J. Virol.* **71**, 2390-2399.
- Lusby, E. W., Fife, K. H., and Berns, K. I. (1980). Nucleotide sequences of the inverted terminal repetition in adeno-associated virus DNA. *J. Virol.* **34**, 402-409.
- Lusby, E. W., Bohensky, R. A., and Berns, K. I. (1981). Inverted terminal repetition in adeno-associated virus DNA: independence of the orientation of either end of the genome. *J. Virol.* **37**, 1083-1086.
- Lusby, E. W., and Berns, K. I. (1982). Mapping of the 5' termini of two adeno-associated virus 2 RNAs in the left half of the genome. *J. Virol.* **41**, 518-526.
- Makhov, A. M., Lee, S. S., Lehman, I. R., and Griffith, J. D. (2003). Origin-specific unwinding of herpes simplex virus 1 DNA by the viral UL9 and ICP8 proteins: visualization of a specific preunwinding complex. *Proc. Natl. Acad. Sci. U S A* **100**, 898-903.
- Marintcheva, B., and Weller, S. K. (2001). A tale of two HSV-1 helicases: roles of phage and animal virus helicases in DNA replication and recombination. *Prog. In Nuc. Acid Research and Mol. Biol.* **70**, 79-118.
- Maul, G. G., Guldner, H. H., and Spivack, J. G. (1993). Modification of discrete nuclear domains induced by herpes simplex virus type 1 immediate early gene 1 product (ICP0). *J. Gen. Virol.* **74**, 2679-2690.
- Maul, G. G., and Everett, R.D. (1994). The nuclear location of PML, a cellular member of the C3HC4 zinc-binding domain protein family, is rearranged during herpes simplex virus infection by the C3CH4 viral protein ICP0. *J. Gen. Virol.* **75**, 1223-1233.
- Mayor, H. D., Drake, S., Stahmann, J., and Mumford, D. M. (1976). Antibodies to adeno-associated satellite virus and herpes simplex in sera from cancer patients and normal adults. *Am. J. Obstet. Gynecol.* **126**, 100-104.
- McCarty, D. M., Christensen, M., and Muzyczka, N. (1991). Sequences required for coordinate induction of adeno-associated virus p19 and p40 promoters by Rep protein. *J. Virol.* **65**, 2936-2945.
- McCarty, D. M., Ryan, J. H., Zolotukhin, S., Zhou, X., and Muzyczka, N. (1994). Interaction of the adeno-associated virus Rep protein with a sequence within the A palindrome of the viral terminal repeat. *J. Virol.* **68**, 4998-5006.
- McGeoch, D. J., Dalrymple, M. A., Dolan, A., McNab, D., Perry, L. J., Taylor, P., and Challberg, M. D. (1988). Structures of herpes simplex virus type 1 genes required for replication of virus DNA. *J. Virol.* **62**, 444-453.

- McLaughlin, S. K., Collis, P., Hermonat, P. L., and Muzyczka, N. (1988). Adeno-associated virus general transduction vectors: analysis of proviral structures. *J. Virol.* **62**, 1963-1973.
- McLean, G. W., Abbotts, A. P., Parry, M. E., Marsden, H. S., and Stow, N. D. (1994). The herpes simplex virus type 1 origin-binding protein interacts specifically with the viral UL8 protein. *J. Gen. Virol.* **75**, 2699-2706.
- McPherson, R. A., Rosenthal, L. J., and Rose, J. A. (1985). Human cytomegalievirus completely helps adeno-associated virus replication. *Virology* **174**, 217-222.
- Mendelman, L. V., Beauchamp, B. B., and Richardson, C. C. (1994). Requirement for a zinc motif for template recognition by the bacteriophage T7 primase. *EMBO J.* **13**, 3909-3916.
- Mendelson, E., Trempe, J. P., and Carter, B. J. (1986). Identification of the *trans*-acting rep proteins of adeno-associated virus by antibodies to a synthetic oligopeptide. *J. Virol.* **60**, 823-832.
- Mizuno, M., and Yoshida, J. (1998). Improvement of transduction efficiency of recombinant adeno-associated virus vector by entrapment in multilamellar liposomes. *Jpn. J. Cancer Res.* **89**, 352-354.
- Monahan, P. E., Jooss, K., and Sands, M. S. (2002). Safety of adeno-associated virus gene therapy vectors: a current evaluation. *Expert Opin Drug Saf.* **1**, 79-91.
- Montgomery, R. I., Warner, M. S., Lum, B. J., and Spear, P. G. (1996). Herpes simplex virus-1 entry into cells mediated by a novel member of the TNF/NGF receptor family. *Cell* **87**, 427-436.
- Morgan, C., Rose, H. M., and Mednis, B. (1968). Electron microscopy of herpes simplex virus. I. Entry. *J. Virol.* **2**, 507-516.
- Mori, S., Wang, L., Takeuchi, T., and Kanda, T. (2004). Two novel adeno-associated viruses from cynomolgus monkey: pseudotyping characterization of capsid protein. *Virol.* **330**, 375-383.
- Muzyczka, N. (1992). Use of adeno-associated virus as a general transduction vector for mammalian cells. *Curr. Top. Microbiol. Immunol.* **158**, 97-129.
- Muzyczka, N., and Berns, K. I. (2001). Parvoviridae: the viruses and their replication. In "Fields Virology" (P. M. Howley, Ed.), Vol. 2, pp. 2327-2359. 2 vols. Lippincott, Philadelphia.
- Nakai, H., Storm, T. A., and Kay, M. A. (2000). Recruitment of single-stranded recombinant adeno-associated virus vector genomes and intermolecular recombination are responsible for stable transduction of liver in vivo. *J. Virol.* **74**, 9451-9463.
- Negorev, D., and Maul G.G. (2001). Cellular proteins localized at and interacting with ND10/PML nuclear bodies/Pods suggest functions of a nuclear depot. *Oncogene* **20**, 7234-7242.
- Ni, T. H., McDonald, W. F., Zolotukhin, I., Melendy, T., Waga, S., Stillman, B., and Muzyczka, N. (1998). Cellular proteins required for adeno-associated virus DNA replication in the absence of adenovirus coinfection. *J. Virol.* **72**, 2777-2787.
- Nimonkar, A. V., and Boehmer, P. E. (2002). *In vitro* strand exchange promoted by the herpes simplex virus type-1 single strand DNA-binding protein (ICP8) and helicase-primase. *J. Biol. Chem.* **277**, 15182-15189.
- Nimonkar, A. V., and Boehmer, P. E. (2003). The herpes simplex virus type-1 single-strand DNA-binding protein (ICP8) promotes strand invasion. *J. Biol. Chem.* **278**, 9678-9682.
- O'Donnell, M. E., Elias, P., Funnel, B. E., and Lehman, I. R. (1987). Interaction between the DNA polymerase and single stranded DNA-binding protein (infected cell protein 8) of herpes simplex virus 1. *J. Biol. Chem.* **262**, 4260-4266.
- Ogston, P., Raj, K., and Beard, P. (2000). Productive replication of adeno-associated virus can occur in human papillomavirus type 16 (HPV-16) episome containing keratinocytes and is augmented by the HPV-16 E2 protein. *J. Virol.* **74**, 3494-3504.
- Ojala, P. M., Sodeik, P., Ebersold, M. W., Kutay, U., and Helenius A. (2000). Herpes simplex virus type 1 entry into host cells: reconstitution of capsid binding and uncoating at the nuclear pore complex in vitro. *Mol. Cell Biol.* **20**, 4922-4931.
- Owens, R. A., Weitzman, M. D., Kyöstiö, S. R. M., and Carter B. J. (1993). Identification of a DNA-binding domain in the amino terminus of adeno-associated virus rep proteins. *J. Virol.* **67**, 997-1005.

- Parris, D. S., Cross, A., Haarr, L., Orr, A., Frame, M. C., Murphy, M., McGeoch, D. J., and Marsden, H. S. (1988). Identification of the gene encoding the 65-kilodalton DNA-binding protein of herpes simplex virus type 1. *J. Virol.* **62**, 818-825.
- Parry, M. E., Stow, N. D., and Marsden, H. S. (1993). Purification and properties of the herpes simplex virus type 1 UL8 protein. *J. Gen. Virol.* **74**, 607-612.
- Pereira, D. J., McCarty, D. M., and Muzyczka, N. (1997). The adeno-associated virus (AAV) Rep protein acts as both a repressor and an activator to regulate AAV transcription during a productive infection. *J. Virol.* **71**, 1079-1088.
- Poffenberger, K. L., and Roizman, B. (1985). Studies on non-inverting genome of a viable herpes simplex virus 1: presence of head-to-tail linkages in packaged genomes and requirements for circulation after infection. *J. Virol.* **53**, 589-595.
- Qing, K., Mah, C., Hansen, J., Zhou, S., Dwarki, V., and Srivastava, A. (1999). Human fibroblast growth factor receptor 1 is a co-receptor for infection by adeno-associated virus. *Nat. Med.* **5**, 71-77.
- Rabinowitz, J. E., and Samulski, R. J. (2000). Building a better vector: the manipulation of AAV virions. *Virology* **278**, 301-308.
- Regad, T., and Chelbi-Alex, M. K. (2001). Role and fate of PML nuclear bodies in response to interferon and viral infections. *Oncogene* **20**, 7274-7286.
- Richardson, W. D., and Anderson, C. W. (1984). Translation of adenovirus 2 late mRNA microinjected into cultured African green monkey kidney cells. *J. Virol.* **51**, 559-562.
- Rixon, F. J., and McLauchlan, J. (1990). Insertion of DNA sequences at a unique restriction enzyme site engineered for vector purposes into the genome of herpes simplex virus type 1. *J. Gen. Virol.* **71**, 2931-2939.
- Roizman, B., and Knipe, D. M. (2001). Herpes simplex viruses and their replication. In "Fields Virology" (P. M. Howley, Ed.), Vol. 2, pp. 2399-2459. 2 vols. Lippincott, Philadelphia.
- Ruffing, M., Zentgraf, H., and Kleinschmidt, J. A. (1992). Assembly of viruslike particles by recombinant structural protein of adeno-associated virus type 2 in insect cells. *J. Virol.* **66**, 6922-6930.
- Russell, D. W. (2003). AAV loves an active genome. *Nat. Genet.* **34**, 241-242.
- Ruyechan, W. T. (1983). The major herpes simplex virus DNA-binding protein holds single-stranded DNA in an extended configuration. *J. Virol.* **46**, 661-666.
- Sambrook, J., Fritsch, E. F., and Maniatis, T. (1989). Molecular cloning: a laboratory manual. 2nd ed. 3 vol. Cold Spring Harbor Laboratory Press, New York.
- Samulski, R. J., Berns, K. I., Tan, M., and Muzyczka, N. (1982). Cloning of adeno-associated virus into pBR322: rescue of intact virus from the recombinant plasmid in human cells. *Proc. Natl. Acad. Sci. U S A* **79**, 2077-2081.
- Samulski, R. J., Srivastava, A., Berns, K. I., and Muzyczka, N. (1983). Rescue of adeno-associated virus from recombinant plasmids: gene correction within the terminal repeats of AAV. *Cell* **33**, 135-143.
- Samulski, R. J., Zhu, X., Xiao, X., Brook, J. D., Housman, D. E., Epstein, N., and Hunter, L. A. (1991). Targeted integration of adeno-associated virus (AAV) into human chromosome 19. *EMBO J.* **10**, 3941-3950. (Erratum, **11**, 1228, 1992)
- Severini, A., Scraba, D. G., and Tyrrell, D. L. (1996). Branched structures in the intracellular DNA of herpes simplex virus type 1. *J. Virol.* **70**, 3169-3175.
- Sherman, G., Gottlieb, J., and Challberg, M. D. (1992). The UL8 subunit of herpes simplex virus helicase-primase complex is required for efficient primer utilization. *J. Virol.* **66**, 4884-4892.
- Shieh, M. T., WuDunn, D., Montgomery, R. I., Esko, J. D., and Spear, P. G. (1992). Cell surface receptors for herpes simplex virus are heparan sulfate proteoglycans. *J. Cell. Biol.* **116**, 1273-1281.
- Showalter, S. D., Zweig, M., and Hampar, B. (1981). Monoclonal antibodies to herpes simplex virus type 1 proteins, including the immediate-early protein ICP4. *Infection and Immunity* **34**, 684-692.

- Shukla, D., Liu, J., Blaiklock, P., Shworak, N. W., Bai, X., Esko, J. D., Cohen, G. H., Eisenberg, R. J., Rosenberg, R. D., and Spear, P. G. (1999). A novel role for 3-O-sulfated heparan sulfate in herpes simplex virus 1 entry. *Cell* **99**, 13-22.
- Sivaraja, M., Giordano, H., and Peterson, M. G. (1998). High-throughput screening assay for helicase enzymes. *Analyt. Biochem.* **265**, 22-27.
- Skaliter, R., Makhov, A. M., Griffith, J. D., and Lehman, I. R. (1996). Rolling circle DNA replication by extracts of herpes simplex virus type 1-infected cells. *J. Virol.* **70**, 1132-1136.
- Snyder, R. O., Im, D.-S., Ni, T., Xiao, X., Samulski, R. J., and Muzyczka, N. (1993). Features of the adeno-associated virus origin involved in substrate recognition by the viral Rep protein. *J. Virol.* **67**, 6096-6104.
- Sodeik, B., Ebersold, M., and Helenius, A. (1997). Dynein mediated transport of incoming herpes simplex virus 1 capsids to the nucleus. *J. Cell Biol.* **136**, 1007-1021.
- Southern, E. M. (1975). Detection of specific sequences among DNA fragments separated by gel electrophoresis. *J. Mol. Biol.* **98**, 503-517.
- Sprecher-Goldberger, S., Thiry, L., Lefèvre, N., Dekegel, D., and de Halleux, F. (1971). Complement-fixation antibodies to adenovirus-associated viruses, adenoviruses, cytomegaloviruses and herpes simplex viruses in patients with tumors and in control individuals. *Am. J. Epidemiol.* **94**, 351-358.
- Starke, G., and Hlinak, P. (1974). Grundriß der allgemeinen Virologie. *Gustav Fischer Verlag*, Jena.
- Stracker, T. H., Cassell, G. D., Ward, P., Loo, Y. M., van Breukelen, B., Carrington-Lawrence, S. D., Hamatake, R. K., van der Vliet, P. C., Weller, S. K., Melendy, T., and Weitzman, M. D. (2004). The Rep protein of adeno-associated virus type 2 interacts with single-stranded DNA-binding proteins that enhance viral replication. *J. Virol.* **78**, 441-453.
- Straus, S. E., Ginsberg, H. S., and Rose, J. A. (1976). DNA-minus temperature-sensitive mutants of adenovirus type 5 help adenovirus-associated virus replication. *J. Virol.* **17**, 140-148.
- Summerford, C., and Samulski, R. J. (1998). Membrane-associated heparan sulfate proteoglycan is a receptor for adeno-associated virus type 2 virions. *J. Virol.* **72**, 1438-1445.
- Summerford, C., Bartlett, J. S., and Samulski, R. J. (1999). AlphavBeta5 integrin: a co-receptor for adeno-associated virus type 2 infection. *Nat. Med.* **5**, 78-82.
- Sun, L., Li, J., and Xiao, X. (2000). Overcoming adeno-associated virus vector size limitation through viral DNA heterodimerization. *Nat. Med.* **6**, 599-602.
- Tattersall, P., and Ward, D. C. (1976). Rolling hairpin model for replication of parvovirus and linear chromosomal DNA. *Nature* **263**, 106-109.
- Taylor, T. J., and Knipe, D. M. (2004). Proteomics of herpes simplex virus replication compartments: association of cellular DNA replication, repair, recombination, and chromatin remodeling proteins with ICP8. *J. Virol.* **78**, 5856-5866.
- Tenney, D. J., Hurlburt, W. M., Micheletti, P. A., Bifano, M., and Hamatake, R. K. (1994). The UL8 component of the herpes simplex virus helicase-primase complex. *J. Biol. Chem.* **270**, 9129-9136.
- Tenney, D. J., Sheaffer, A. K., Hurlburt, W. W., Bifano, M., and Hamatake, R. K. (1995). Sequence-dependent primer synthesis by the herpes simplex virus helicase-primase complex. *J. Biol. Chem.* **270**, 9129-9136.
- Thomson, B. J., Weindler, F. W., Gray, D., Schwaab, V., and Heilbronn, R. (1994). Human herpesvirus 6 (HHV6) is a helper virus for adeno-associated virus type 2 (AAV2) and the rep gene homologue in HHV6 can mediate AAV-2 DNA replication and regulate gene expression. *Virology* **204**, 304-311.
- Towbin, H., Stachelin, T., and Gordon, J. (1979). Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedures and some applications. *Proc. Natl. Acad. Sci. U S A* **76**, 4350-4353.
- Tratschin, J. D., Miller, I. L., and Carter, B. J. (1984). Genetic analysis of adeno-associated virus: properties of deletion mutants constructed in vitro and evidence for an adeno-associated virus replication function. *J. Virol.* **51**, 611-619.

- Trego, K. S., and Parris, D. S. (2003). Functional interaction between the herpes simplex virus type 1 polymerase processivity factor and origin-binding proteins: enhancement of UL9 helicase activity. *J. Virol.* **77**, 12646-12659.
- Urabe, M., Ding, C., and Kotin, R. M. (2002). Insect cells as a factory to produce adeno-associated virus type 2 vectors. *Hum. Gene Ther.* **13**, 1935-1943.
- Vlazny, D. A., Kwong, A., and Frenkel, N. (1991). Site-specific cleavage/packaging of herpes simplex virus DNA and the selective maturation of nucleocapsids containing full-length viral DNA. *Proc. Natl. Acad. Sci. U.S.A.* **79**, 1423-1427.
- Wadsworth, S., Jacob, R. J., and Roizman, B. (1975). Anatomy of herpes simplex virus DNA. II. Size, composition, and arrangement of terminal repetitions. *J. Virol.* **15**, 1487-1497.
- Walz, C., Deprez, A., Dupressoir, T., Dürst, M., and Schlehofer, J. R. (1997). Interaction of human papillomavirus type 16 and adeno-associated virus type 2 co-infection human cervical epithelium. *J. Gen. Virol.* **78**, 1441-1452.
- Ward, P., Falkenberg, M., Elias, P., Weitzmann, M., and Linden, R. M. (2001). Rep-dependent initiation of adeno-associated virus type 2 DNA replication by a herpes simplex virus type 1 replication complex in a reconstituted system. *J. Virol.* **75**, 10250-10258.
- Warren, S., and Chute, R. N. (1972). Pheochromocytoma. *Cancer* **29**, 327-331.
- Weger, S., Wistuba, A., Grimm, D., and Kleinschmidt, J. A. (1997). Control of adeno-associated virus type 2 Cap gene expression: relative influence of helper virus, terminal repeats, and Rep proteins. *J. Virol.* **71**, 8437-8447.
- Weger, S., Wendland, M., Kleinschmidt, J. A., and Heilbronn, R. (1999). The adeno-associated virus type 2 regulatory Rep78/Rep68 interact with the transcriptional coactivator PC4. *J. Virol.* **73**, 260-269.
- Weindler, F. W., and Heilbronn, R. (1991). A subset of herpes simplex virus replication genes provides helper functions for productive adeno-associated virus replication. *J. Virol.* **65**, 2476-2483.
- Weir, H. M., Calder, J. M., and Stow, N. D. (1989). Binding of the herpes simplex virus type 1 UL9 gene product to an origin of viral DNA replication. *Nuc. Acid Res.* **17**, 1409-1425.
- Weitzman, M. D., Kyöstiö, S. R. M., Kotin, R. M., and Owens, R. A. (1994). Adeno-associated virus (AAV) Rep proteins mediate complex formation between AAV DNA and its integration site in human DNA. *Proc Natl Acad Sci U.S.A.* **91**, 5808-5812.
- Wilcock, D., and Lane, D. P. (1991). Localization of p53, retinoblastoma and host replication proteins at sites of viral replication in herpes-infected cells. *Nature* **349**, 429-431.
- Wilkinson, D.E., Weller, S.K. (2003). The role of DNA recombination in herpes simplex virus DNA replication. *IUBMB Life* **55**, 451-458.
- Wu, C. A., Nelson, N. J., McGeoch, D. J., and Challberg, M. D. (1988). Identification of herpes simplex virus type 1 genes required for origin-dependent DNA synthesis. *J. Virol.* **62**, 435-443.
- Wonderling, R. S., Kyösti, S. R. M., and Owens, R. A. (1995). A maltose-binding protein/adeno-associated virus Rep68 fusion protein has DNA-RNA helicase and ATPase activities. *J. Virol.* **69**, 3542-3548.
- Wonderling, R. S., and Owens, R. A. (1997). Binding sites for adeno-associated virus Rep proteins within the human genome. *J. Virol.* **71**, 2528-2534.
- Xiao, X., Li, J., and Samulski, R.J. (1998). Production of high-titer recombinant adeno-associated virus vectors in the absence of helper adenovirus. *J. Virol.* **72**, 2224-2232.
- Xiao, W., Chirmule, N., Berta, S. C., McCullough, B., Gao, G., and Wilson, J. M. (1999). Gene therapy vectors based on adeno-associated virus type 1. *J. Virol.* **73**, 3994-4003.
- Xiao, W., Warrington, K. H., Jr., Hearing, P., Hughes, J., and Muzyczka, N. (2002). Adenovirus-facilitated nuclear translocation of adeno-associated virus type 2. *J. Virol.* **76**, 11505-11517.
- Yakobson, B., Koch, T., and Windocour, E. (1987) Replication of adeno-associated virus in synchronized cells without the addition of a helper virus. *J. Virol.* **61**, 972-981.

- Yakobson, B., Hryntko, T. A., Peak, M. J., and Winocour, E. (1989). Replication of adeno-associated virus in cells irradiated with UV light at 254 nm. *J. Virol.* **63**, 3123-3129.
- Yalkinoglu, A. Ö., Heilbronn, R., Bürkle, A., Schlehofer, J. R., and zur Hausen, H. (1988). DNA amplification of adeno-associated virus as a response to cellular genotoxic stress. *Cancer Res.* **48**, 3123-3129.
- Yan, Z., Zhang, Y., Duan, D., and Engelhardt, J. F. (2000). Trans-splicing vectors expand the utility of adeno-associated virus for gene therapy. *Proc. Natl. Acad. Sci. U S A* **97**, 6716-6721.
- Yang, Q., Chen, F., and Trempe, J. P. (1994). Characterization of cell lines that inducibly express the adeno-associated virus Rep proteins. *J. Virol.* **68**, 4847-4856.
- York, I. A., Roop, C., Andrews, D. W., Riddell, S. R., Graham, F. L., and Johnson, D. C. (1994). A cytosolic herpes simplex virus protein inhibits antigen presentation to CD8+ T lymphocytes. *Cell* **77**, 525-535.
- Young, S. M., Jr., McCarty, D. M., Degtyareva, N., and Samulski, R. J. (2000). Roles of adeno-associated virus Rep protein and human chromosome 19 in site-specific recombination. *J. Virol.* **74**, 3953-3966.
- Zhang, H.-G., Wang, Y. M., Xie, J. F., Liang, X., Hsu, H.-C., Zhang, X., Douglas, J. M., Curiel, D. T., and Mountz, J. D. (2001). Recombinant adenovirus expressing adeno-associated virus cap and rep proteins supports production of high-titer recombinant adeno-associated virus. *Gene Ther.* **8**, 704-712.
- Zhong, L., and Hayward, G. S. (1997). Assembly of complete, functionally active herpes simplex virus DNA replication compartments and recruitment of associated viral and cellular proteins in transient cotransfection assays. *J. Virol.* **71**, 3146-3160.
- Zhou, X., Zolotukhin, I., Im, D.-S., and Muzyczka, N. (1999). Biochemical characterization of adeno-associated virus Rep68 DNA helicase and ATPase activities. *J. Virol.* **73**, 1580-1590.
- Zhu, L., and Weller S. K. (1992a). The UL5 gene of herpes simplex virus type 1: isolation of a *lacZ* insertion mutant and association of the UL5 gene product with other members of the helicase-primase complex. *J. Virol.* **66**, 458-468.
- Zhu, L., and Weller S. K. (1992b). The six conserved helicase motifs of the UL5 gene product, a component of the herpes simplex virus type 1 helicase-primase, are essential for its function. *J. Virol.* **66**, 469-479.
- Zolotukhin, S. (2005). Production of recombinant adeno-associated virus vectors. *Hum. Gene Ther.* **16**, 551-557.

7 ZUSAMMENFASSUNG

Das adenoassoziierte Virus (AAV) ist ein helperabhängiges Virus, das für seine produktive Vermehrung die Koinfektion mit einem Helpervirus benötigt, z.B. dem Herpes-simplex-Virus (HSV). Als HSV-Helperproteine dienen vier HSV-Replikationsproteine, die zusammen mit dem AAV-Replikationsprotein Rep den Minimalkomplex für die Replikation von AAV bilden. Die vier benötigten HSV-Helperproteine sind das *single-strand DNA-binding protein* ICP8 und ein trimerer Protein-Komplex mit Helicase- und Primase-Aktivität. Nach Koinfektion von AAV und HSV wird das einzelsträngige AAV-DNA-Genom in subnukleäre HSV-Replikationskompartimente transloziert, in denen das AAV-Genom repliziert werden kann. Hierbei kolokalisieren HSV-ICP8 und AAV-Rep in Gegenwart des einzelsträngigen (ssDNA) AAV-Genoms.

In der vorliegenden Arbeit konnte gezeigt werden, dass für die ssDNA-abhängige Rekrutierung von AAV-Rep in nukleäre Replikationskompartimente das Zusammenwirken der vier HSV-Helperproteine hinreichend und notwendig ist. Für die Bildung der nukleären Replikationskomplexe mussten weder die HSV-Helicase noch die HSV-Primase enzymatisch aktiv sein. Dies wurde gezeigt durch Mutanten der HSV-Helicase bzw. -Primase, bei denen gezielt einzelne Aminosäure-Austausche in die katalytischen Zentren eingeführt wurden. Alle Mutanten hatten ihre enzymatische Aktivität vollständig verloren unter Erhalt der Interaktionsfähigkeit als Proteinkomponenten des trimeren Helicase-Primase-Komplexes.

Um zu testen, ob während des weiteren Verlaufs der AAV-DNA-Replikation die enzymatische Aktivität von HSV-Primase bzw. -Helicase benötigt wird, wurden AAV-DNA-Replikationsanalysen nach Transfektion der vier HSV-Helfergenkonstrukte durchgeführt. Auch hierbei wurden sowohl Helicase als auch Primase vor allem als strukturelle Komponenten der Replikationskomplexe benötigt. Dies entsprach den Erwartungen, da das AAV-ssDNA-Genom an den Enden über partiell doppelsträngige palindromische Strukturen verfügt, die durch komplementäre Rückfaltung die Funktion des Primers für die DNA-Polymerase übernehmen. Bei Präsenz der HSV-Polymerase nach HSV-Infektion zeigten die AAV-Replikationsuntersuchungen, dass bei enzymatisch aktiver HSV-Primase AAV effizienter repliziert wird, möglicherweise durch Rekrutierung des HSV-Polymerase-komplexes zur Replikation des AAV-Genoms.

Die HSV-Helicase besitzt wie AAV-Rep ATPase- und Helicase-Aktivität. Deshalb überraschte der weitere Befund, dass die enzymatisch aktive HSV-Helicase bei Transfektion des minimalen HSV-Replikationskomplexes eine nachweisbare, wenn auch geringe Steigerung der AAV-DNA-Replikationsrate zeigte. Als Erklärung kommt eine Stimulation der homologen Rekombination zur Auflösung hochmolekularer AAV-DNA-Replikationsintermediate in Frage. Aufgrund der entgegengesetzten Polarität der HSV-Helicase könnte diese auch die Rep-Helicase funktionell komplementieren. Die Quantifizierung der AAV-Replikationsintermediate zeigte zudem, dass zusätzlich bislang noch unbekannte, weitere HSV- oder zelluläre Faktoren existieren müssen, die die AAV-Replikation weiter stimulieren.

Aufbauende Untersuchungen werden nötig sein, um die strukturellen und funktionellen Interaktionen des Multiprotein-Komplexes aus AAV-Rep, HSV-*single-strand DNA-binding protein*, dem trimeren HSV-Helicase-Primase-Komplex und der AAV-ssDNA zu entschlüsseln. Neben dieser grundlegenden Frage sind die Untersuchungen auch für die Weiterentwicklung der AAV-Vektortechnologie von Bedeutung. Für die weitere Optimierung effizienter Verpackungssysteme für die AAV-Vektorproduktion im biotechnologisch großen Maßstab sind HSV-basierte Systeme äußerst vielversprechend. Deren Weiterentwicklung hängt wesentlich von einem guten Verständnis der Interaktionen zwischen AAV, HSV und zellulären Faktoren während der einzelnen Schritte der AAV-Replikation ab.

8 ANHANG

8.1 MONOCLONAL ANTIBODIES AGAINST UL5

The HSV helicase-primase complex (UL5-UL8-UL52 complex) was purified from *Spodoptera frugiperda* cells infected with a recombinant baculovirus that expresses all three proteins (AcUL5-UL8-UL52, essentially as described (Dodson *et al.*, 1989)). The purified protein was used to immunize female BALB/c mice, and hybridoma lines secreting monoclonal antibodies (mAbs) were generated as described previously (McLean *et al.*, 1994). Mabs reactive with the complex were initially identified by an ELISA assay, and subsequently assessed for their ability to interact with the individual components by Western Blotting using extracts of *Spodoptera frugiperda* cells infected with baculovirus expressing UL5, UL8 or UL52 (Stow *et al.*, 1992).

Dodson, M. S., Crute, J. J., Bruckner, R. C., and Lehman, I. R. (1989). Overexpression and assembly of the herpes simplex virus type 1 helicase-primase in insected cells. *J. Biol. Chem.* **264**, 20835-20838.

McLean, G. W., Abbotts, A. P., Marsden, H. S., and Stow, N. D. (1994). The herpes simplex virus type 1 origin-binding protein interacts specifically with the viral UL8 protein. *J. Gen. Virol.* **75**, 2699-2706.

Stow, N. D. (1992). Herpes simplex virus type 1 origin-dependent DNA replication in insect cells using recombinant baculoviruses. *J. Gen. Virol.* **73**, 313-321.

8.2 VERÖFFENTLICHUNGEN

A) Artikel in Zeitschriften:

Hattermann, K., Maerz, A., Slanina, H., Schmitt, C., and Mankertz, A. (2004). Assessing the risk potential of porcine circoviruses for xenotransplantation: consensus primer-PCR-based search for a human circovirus. *Xenotransplantation* **11**, 547-550.

Slanina, H., Weger, S., Stow, N. D., Kuhrs, A., and Heilbronn, R. (2006). Role of the Herpes Simplex Virus Helicase-Primase Complex during Adeno-Associated Virus DNA Replication. *J. Virol.* **80**, 5241-5250.

B) Vorträge:

Slanina, H., Weger, S., Kuhrs, A., Stow, N. D., Weller, S. K., and Heilbronn, R. (2003). Functional Analysis of the HSV Helicase-Primase-Complex for the Replication of Adeno-Associated Virus. *Jahrestagung der Gesellschaft für Virologie*, Berlin.

Slanina, H., Weger, S., Stow, N. D., Kuhrs, A., and Heilbronn, R. (2006). Role of the Herpes Simplex Virus Helicase-Primase Complex during Adeno-Associated Virus DNA Replication. *IXth Parvovirus Workshop*, Les Diablerets, Schweiz.

C) Poster:

Slanina, H., Weger, S., Kuhrs, A., Stow, N. D., and Heilbronn, R. (2005). Functional Analysis of the HSV Helicase-Primase-Complex for AAV DNA replication. *Jahrestagung der Gesellschaft für Virologie*, Hannover.

Slanina, H., Weger, S., Stow, N. D., Kuhrs, A., and Heilbronn, R. (2006). Role of the Herpes Simplex Virus Helicase-Primase Complex during Adeno-Associated Virus DNA Replication. *Jahrestagung der Gesellschaft für Virologie*, München.

8.3 SELBSTSTÄNDIGKEITSERKLÄRUNG

„Ich, Heiko Slanina, erkläre, dass ich die vorgelegte Dissertationsschrift mit dem Thema „Die Rolle des Helicase-Primase-Komplexes von Herpes-simplex-Virus Typ 1 bei der DNA-Replikation des adenoassoziierten Virus“ selbst verfasst und keine anderen als die angegebenen Quellen und Hilfsmittel benutzt, ohne die Hilfe Dritter verfasst und auch in Teilen keine Kopien anderer Arbeiten dargestellt habe.“

Berlin, 26.09.2006

Heiko Slanina

8.4 DANKSAGUNGEN

An dieser Stelle möchte ich mich bei allen Personen herzlich bedanken, die auf das Zustandekommen dieser Arbeit durch ihre fachliche und moralische Unterstützung positiven Einfluss genommen haben.

Mein besonderer Dank gilt Frau Prof. Dr. Regine Heilbronn für die freundliche Aufnahme in ihre Arbeitsgruppe, die Überlassung des interessanten Themas, die fachliche Betreuung und die Möglichkeit, die Arbeit auf vielen Kongressen vorstellen und publizieren zu können.

Besonders herzlich möchte ich mich bei Herrn Dr. Stefan Weger für die Einführung in das wissenschaftliche Arbeiten bedanken. Er stand mir stets bei allen Fragen und Diskussionen geduldig und fachkundig zur Seite.

Bei Frau Prof. Dr. Sandra K. Weller bedanke ich mich für die Überlassung der Zinkfinger-Mutante und für produktive fachliche Diskussionen. Herrn Prof. Dr. Nigel D. Stow gilt mein Dank für die Helicase- und Primaseantikörper sowie für die Mitarbeit an der Publikation. Ein Dankeschön geht auch an Herrn Prof. Dr. Jim P. Trempe für die großzügige Überlassung des Anti-rabbit Rep-Antiserums.

Herrn PD Dr. Joachim Mankertz und Jan Richter danke ich für die schnelle und unkomplizierte Hilfe bei der technischen Unterstützung der konfokalen Mikroskopie.

Ein großes Dankeschön geht an alle Kolleginnen und Kollegen für die gute und harmonische Arbeitsatmosphäre. Sie standen mir nicht nur fachlich zur Seite, sondern auch bei den kleinen Problemen des Alltags. Hier möchte ich besonders Dr. Annette Kuhrs, die für mich die von mir verwendeten Viren gezüchtet und titriert hat, und Frau Eva Hammer, die fachlich und moralisch immer eine große Stütze war, erwähnen.

Last but not least bedanke ich mich bei meiner Familie, Freunden und Bekannten, die mich stets bei meinem Studium und der Fertigstellung dieser Arbeit unterstützt und mir auch großes Verständnis entgegengebracht haben.

8.5 LEBENSLAUF

Mein Lebenslauf wird aus Datenschutzgründen in der elektronischen Version meiner Arbeit nicht mit veröffentlicht.