

## 8. SUMMARY

As previously described, *Icsbp* deletion leads to changed differentiation program of myeloid progenitors reflected in disproportional high production of granulocytes and low production of macrophages. In order to study the underlying mechanism of this switch, the gene expression profiles of isolated bipotent granulocyte-macrophage progenitors (GMP) from *Icsbp*<sup>+/+</sup> and *Icsbp*<sup>-/-</sup> mice were compared by Affymetrix technology. This analysis revealed deregulation of several other hematopoietic transcription factors, which led us to hypothesize that defective myelopoiesis of *Icsbp*<sup>-/-</sup> mice is caused by deregulation of multiple transcription factors, rather than *Icsbp* deletion alone. The goal of this work was to investigate the contribution of one of these additional deregulations to the observed *Icsbp*<sup>-/-</sup> mouse phenotype, namely the down-regulation of the transcription factor *Klf4*.

*Klf4* was chosen for this analysis as one of most strongly down-regulated factors (10 fold) in *Icsbp*<sup>-/-</sup> progenitors. At the time this study began, the expression of *Klf4* in the hematopoietic system has not yet been reported. However, its essential role in the differentiation of epithelial tissues and important roles of its analogues *Klf1* and *Klf2* in erythroid i.e. lymphoid cells, prompted us to analyze the role of *Klf4* in the development of myeloid cells.

The experiments described in this work identify *Klf4* as a factor which has a strong influence on myeloid progenitors, directing them to differentiate along the macrophage lineage, as shown by significantly higher percentage of macrophage colonies developing from *Klf4-ER*<sup>T2</sup> over-expressing myeloid progenitors. *Klf4-ER*<sup>T2</sup> led to the macrophage maturation enhancement even when the progenitor cells were cultured with granulocyte-colony stimulating factor (G-CSF) or when it was expressed in the *Icsbp*<sup>-/-</sup> cells (confering genetic misbalance which results in a propensity to form granulocytes), suggesting the ability of *Klf4* over-expression to override granulocyte-promoting effects of both extracellular cytokine signals and intracellular factors and reprogram the cell fate to the macrophage lineage. Analysis of the early gene expression changes caused by *Klf4-ER*<sup>T2</sup> over-expression in myeloid progenitors revealed up-regulation of several putative macrophage genes and down-regulation of genes involved in granulocyte proliferation, suggesting that *Klf4* exerts its macrophage promoting effect by coordinating molecular changes which precede the commitment to this lineage.

Another feature of the *Klf4-ER*<sup>T2</sup> over-expression is its strong cytostatic effect on developing myeloid cells, identified by the significant reduction in the total colony formation. The proliferation inhibition coupled with the macrophage maturation promoting activity of *Klf4-ER*<sup>T2</sup> suggest that *Klf4* acts as a proliferation/differentiation switch in the myeloid development.

*Klf4* effect on macrophage maturation is independent from *Icsbp*, as shown by uncompromised *Klf4* function in *Icsbp*<sup>-/-</sup> cells and uncompromised *Icsbp* function in *Klf4*<sup>-/-</sup> cells.

Unlike *Icsbp*<sup>-/-</sup> mice, the mice with a conditional *Klf4* deletion showed no overt deregulation in the hematopoietic development, suggesting that the role of *Klf4* in the steady-state hematopoiesis is redundant. Alternatively, *Klf4* could play a role in stress-induced conditions like infections (since it

up-regulates macrophage-activation markers) or in the protection from oncogenic processes (since it leads to p21<sup>Waf1</sup> up-regulation, cell-cycle arrest and maturation).

Klf4 effects are based on its transcriptional activity and interaction with other nuclear factors. Deletion of the protein-interaction responsible domain rendered Klf4 inactive, while deletion of the DNA-binding zinc-finger region resulted in a maturation block and an extensive proliferation of immature myeloid cells. This result suggests the oncogenic potential of Klf4 in the myeloid system.

One of the Klf4 transcriptional targets (confirmed by our study) is p21<sup>Waf1</sup>. The p21<sup>Waf1</sup> contribution to the proliferation inhibition and macrophage maturation enhancement caused by Klf4 over-expression was analyzed by testing the effect of p21<sup>Waf1</sup> over-expression in Klf4<sup>+/+</sup> and Klf4<sup>-/-</sup> myeloid progenitors. In addition to the full-length p21<sup>Waf1</sup> over-expression, the compartmentalized p21<sup>Waf1</sup> expression was simulated by the ability of the p21-ER<sup>T2</sup> fusion construct to translocate in the nucleus in the presence of the ER<sup>T2</sup>-ligand (4-OHT) or to stay sequestered in the cytosol in the ligand absence. Enhanced macrophage formation was observed in all cases of forced p21<sup>Waf1</sup> expression (constitutive, cytosolic or nuclear), in wild type and Klf4<sup>-/-</sup> progenitor cells, suggesting that forced expression of p21<sup>Waf1</sup> itself enhances macrophage maturation. Gene expression analysis of p21-ER<sup>T2</sup> expressing progenitors in early differentiation stages showed the same molecular changes as Klf4 over-expression, suggesting that p21<sup>Waf1</sup> mediates Klf4 macrophage maturation effect. However, the intensity of p21<sup>Waf1</sup> effect on the maturation tended to be milder than that of Klf4, indicating that Klf4 activates other, p21<sup>Waf1</sup> independent factors which contribute to the macrophage maturation.

The cytostatic effect of Klf4-ER<sup>T2</sup> over-expression was largely reproduced by p21<sup>Waf1</sup> over-expression, suggesting that this aspect of Klf4 activity could also be mediated through p21<sup>Waf1</sup>. Since the reduction in the colony formation was observed only when p21<sup>Waf1</sup> was allowed to enter the nucleus, we conclude that the roles of p21<sup>Waf1</sup> in the differentiation of myeloid cells and cell cycle control can be decoupled and depend on the subcellular localization of p21<sup>Waf1</sup>.

Apart from analyzing the factors that govern the development of macrophages and neutrophilic granulocytes, another aspect of myelopoiesis, development of eosinophilic granulocytes, was analyzed in this work. The gene profiling of Icsbp<sup>+/+</sup> and Icsbp<sup>-/-</sup> GMPs described above identified strong down-regulation of several eosinophil-specific genes in the absence of Icsbp, suggesting perturbed eosinophilopoiesis in Icsbp<sup>-/-</sup> mice. Indeed, experiments described here confirmed that Icsbp<sup>-/-</sup> mice have reduced number of eosinophil progenitors and reduced capability to elicit eosinophilia in response to parasite infection or sterile inflammation. This defective eosinophil proliferation is based on reduced expression of the main eosinophil growth factor receptor, Il-5R $\alpha$  and reduced expression of an important intrinsic factor in defining the eosinophilic lineage, Gata1.

Summarized, this work identifies Klf4 as novel macrophage maturation promoting factor, whose effects in myelopoiesis are partially mediated by its downstream target p21<sup>Waf1</sup>. Additionally, this work revealed previously undetected myeloipoietic defect in Icsbp<sup>-/-</sup> mice, namely perturbed development of eosinophils, identifying thereby Icsbp as an important factor in eosinophilopoiesis.