

7. Appendix

Abstract

The completion of the sequencing of the human genome and the development of global high throughput approaches have changed and enriched biological experimentation. The most important task is not the elucidation of the function of all encoded proteins. One of the limitations that prevent the application of biochemical methods to high-throughput experimentation is the inability to express and purify large numbers of proteins in high-throughput format.

To develop a high-throughput method to purify proteins from *E. coli*, a 96-well format compatible protein purification process was developed for His₆-tagged proteins under denaturing conditions. Under non-denaturing conditions the choice of the purification tag can have a significant impact on the purification success of individual proteins. To develop a general method for the parallel purification of many proteins, four different purification tags were evaluated using a test set of 32 sequence-verified human cDNAs of varying sizes and activities. The basic purification process was adapted to the four different chemistries and all 128 constructs were purified and characterized with respect to yield and purity. This analysis revealed that the GST-tag and the MBP-tag were equally effective and purified 28/32 proteins. The purification methods were shown to be compatible with the functional integrity of the proteins.

In order to evaluate the developed methods on a larger test set, 771 and 428 different proteins were purified with the His₆-tag under denaturing conditions and the GST-tag under non-denaturing conditions respectively. In this experiment 67% of all His₆-tagged proteins were purified under non-denaturing and 49% of the GST-tagged proteins were purified under non-denaturing conditions.

The biochemical and biophysical parameters of the purified proteins were analyzed to identify protein properties that are predictive of protein purification success from bacteria. We found that the length of the single longest hydrophobic stretch in the primary structure of His₆-tagged proteins determines the likelihood of successful protein purification.

Zusammenfassung

Die vollständige Sequenzierung des menschlichen Genoms und die Entwicklung von globalen experimentellen Ansätzen haben die biologische Forschung verändert und bereichert. Die funktionelle Charakterisierung aller Proteine ist zur Zeit eine der dringenden Aufgaben. Leider sind biochemische Methoden für funktionelle Studien im Verfahren mit hohem Durchsatz zur Zeit nicht nutzbar, da die hierzu erforderlichen Proteine nicht im Hochdurchsatzverfahren aufgereinigt werden können.

Um eine allgemein anwendbare Methode für die parallele Aufreinigung von vielen verschiedenen Proteinen unter nicht-denaturierenden Bedingungen zu entwickeln, wurde zunächst ein Reaktionsplatten-kompatibler Prozess für Proteinaufreinigungen von Bakterien erarbeitet und anschließend wurden vier verschiedene Affinitätsankerproteine im Kontext von 32 Test-Proteinen evaluiert. Alle 128 Fusionsproteine wurden im Reaktionsplattenformat aufgereinigt und in Bezug auf Reinheit und Ausbeute charakterisiert. Diese Analyse ergab, dass der Glutathione-S-Transferase-Anker ('GST-tag') und der Maltose-Bindungs-Protein-Anker ('MBP-tag') gleich effektiv waren und die Aufreinigung von 28/32 beziehungsweise 28/31 Proteinen ermöglichten. Es wurde gezeigt, dass beide Aufreinigungsmethoden funktionelle Proteine produzierten.

Um die entwickelten Methoden mittels eines größeren Proteinsatzes zu evaluieren, wurden 771 beziehungsweise 428 verschiedene Proteine als His₆-Fusionsproteine beziehungsweise als GST-Fusionsproteine aufgereinigt. In diesem Experiment konnten 67% aller His₆-Fusionsproteine unter denaturierenden Bedingungen und 49% aller GST-Fusionsproteine unter nicht-denaturierenden Bedingungen aufgereinigt werden.

Letztlich wurden biochemische und biophysikalische Daten aller Proteine untersucht um Eigenschaften zu identifizieren, die es ermöglichen den Aufreinigungserfolg einzelner Proteine vorherzusagen. Unter anderem wurde festgestellt, dass die Länge, des längsten durchgehenden Abschnittes der Primärstruktur mit einer durchschnittlichen Hydrophobizität >0.85 (nach Goldman, Engelman, Steiz) mit dem Aufreinigungserfolg von His₆-Fusionsproteinen korreliert.

Abbreviations

2DE	2 Dimensional Electrophoresis
2D-LC	2 Dimensional Liquid Chromatography
2-HA	2 Hybrid Assay
ALL	Acute Lymphoblastic Leukemia
AML	Acute Myelogenous Leukemia
APC	Anaphase Promoting Complex
Appr ^{>p}	Adenosine Diphosphate Ribose 1''-2'' Cyclic Phosphate
B	Bead Bound Fraction
BAC	Bacterial Artificial Chromosome
b-gal	beta-galactosidase
CAP	Cellulose Associated Protein
CBP	Calmodulin Binding Peptide
CDK	Cyclin Dependent Kinase
cDNA	Complementary DNA
CDS	Coding Sequence
DDR	DNA Damage Response
DNA	Deoxyribonucleic Acid
EDTA	Ethylenediaminetetraacetic Acid
ELISA	Enzyme Linked Immunosorbant Assay
Elu	Elate
EST	Expressed Sequence Tag
FLEX	Full Length Expression
Gal-1-P	Galactose 1 Phosphate
Gal4-AD	Gal4 Activation Domain
GB1	B1 Domain of Protein G
GFP	Green Fluorescent Protein
GO	Gene Ontology
GST	Glutathione-S-Transferase
HA	Hemagglutinin

HGP	Human Genome Project
Hrs	Hours
HT	High Throughput
IHF	Integration Host Factor
In	Input (Fraction)
INK4	Inhibitor of Cyclin Dependent Kinase 4
Int	Integrase
IPTG	Isopropylthio- β -D-galactoside
kDa	Kilodalton
LB	Luria-Bertani
M	Moles
MALDI	Matrix-Assisted Laser Desorption
MBP	Maltose Binding Protein
Md	Maryland
min	Minutes
ml	Microliter
mM	Millimolar
MML	Myelomonocytic Leukemia
mRNA	Messenger RNA
MS	Mass Spectrometry
n/a	Not Applicable
Ni-NTA	Nickel
NMR	Nuclear Magnetic Resonance
NP-40	Nonidet P-40
O/N	Over Night
OD	Optical Density
ORF	Open Reading Frame
PAGE	Polyacrylamide Gel Electrophoresis
PCR	Polymerase Chain Reaction
PTM	Post Translational Modification
QC	Quality Control

RNA	Ribonucleic Acid
rRNA	Ribosomal RNA
S	Supernatant
SAGA	Spt-Ada-Gcn5-acetyltransferase transcriptional activator–histone acetyltransferase complex
SDS-PAGE	Sodium-Dodecyl-Sulfate Polyacrylamide Gel Electrophoresis
sec	Seconds
SNP	Single Nucleotide Polymorphism
T	Total (starting material)
TB	Terrific Broth
TM	Transmembrane protein(s)
Trx	Thioredoxin
UTR	Untranslated Region
W	Wash (Fraction)
Xis	Excisionase

Publications

Braun P, Hu Y, Weng H, Schick M, Korn B, LaBaer J; Different Parameters Dictate Protein Expression Success of His₆- and GST-tagged Proteins in Bacteria, manuscript in preparation

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Curriculum Vitae

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1997 – present	Graduate Studies	Institute of Proteomics – Harvard Medical School Advisor: Ed Harlow
1996	Diploma in Biochemistry	Free University of Berlin
1991	Abitur (High School degree)	Staedtisches Gymnasium, Luisenschule Essen

Experience/Techniques:

Laboratory: Cell culture of mammalian and insect cell lines; protein expression in bacteria, human cells and in the baculovirus system; protein purification using ion exchange chromatography and protein affinity tags; immunoprecipitation of endogenous and overexpressed proteins, western blot analysis, enzyme assays, immunofluorescence, recombinational cloning, enzymatic cloning techniques, PCR, hybridization, TECAN Gemini, TECAN GenMate, high throughput experimentation

B. Report of Teaching

Training and supervision of undergraduate students	2003 - present
Cold Spring Harbor Course Proteomics Teaching Assistant	2002
Training and supervision of technical and laboratory assistants	1999- 2002

Part III: Bibliography

A. Publications: see above

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Merck Pilot Research Program Grant	2003
Boehringer Ingelheim Fonds (Fellowship)	1997 - 2000
Society for Biological Chemistry – GBCh (Training Award)	1995

C. Selected Presentations

ABRF (invited speaker)	2003
Proteome Society (selected speaker)	2002
IBC Conference - Bioinformatics (invited speaker)	2000
Cold Spring Harbor - Cancer Genetics and Tumor Suppressor Genes (selected speaker)	2000

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