

÷

Title: Polarization enhanced two-photon excited fluorescence contrast by shaped laser pulses using a deformable phase plate

Author(s):

Syed Shakir Ashraf Shah Bukhari, Ayan Halder, and Albrecht Lindinger

Document type: Postprint

Terms of Use: Copyright applies. A non-exclusive, non-transferable and limited right to use is granted. This document is intended solely for personal, non-commercial use.

Citation:

"Syed Shakir Ashraf Shah Bukhari, 2023, Applied Optics Vol. 62, Issue 31, pp. 8242-8247; https://doi.org/10.1364/AO.503531" Archiviert unter: http://dx.doi.org/10.17169/refubium-43042 Polarization enhanced two-photon excited

fluorescence contrast by shaped laser pulses using a deformable phase plate

4 SYED SHAKIR ASHRAF SHAH BUKHARI, AYAN HALDER, AND

5 ALBRECHT LINDINGER

Abstract: We utilize spatially and temporally tailored laser pulses for polarization enhanced 6 two-photon excited fluorescence contrasts of dyes. The shaped laser pulses are produced by first passing through a temporal pulse shaper and then through a novel two-dimensional 8 spatial pulse shaper with deformable phase plates. Different spatial beam profiles are presented 9 which demonstrate the potential of the new spatial pulse shaper. Particularly, a polarization 10 enhanced fluorescence contrast between two dyes is reported by utilizing specific phase shaping 11 in perpendicular polarization directions. The tailored laser pulses are further modified by the 12 deformable phase plate and a polarization increased depth-dependent contrast is achieved. This 13 spatial shaping for all polarization directions demonstrates the advantage of deformable phase 14 plate spatial shapers compared to liquid crystals, where only one polarization direction can 15 spatially be modified. The described polarization contrast method allows for three-dimensional 16 scanning of probes and provides new perspectives for biophotonic applications. 17

19 1. Introduction

In the last years laser pulse shaping has evolved to a various tool for several research fields 20 such as optimal control [1-4], nanooptics [5], and control of biomolecules [6]. Thereto, 21 Fourier domain shaping with liquid crystal modulators was used resulting in temporally shaped 22 light fields and furthermore in the modification of the light polarization which extends the 23 pulse shaping possibilities [7–9]. Tailored laser pulses were also employed for multiphoton 24 excited fluorescence [10], whereby intrapulse interference was utilized to selectively excite 25 differing molecules. This approach has lead to three-dimensional imaging in multiphoton 26 microscopy [11, 12]. 27

Spatial modification of the laser beam profile can be conducted by using a two-dimensional 28 modulator and corresponding focussing lenses [13]. It is carried out by liquid crystal modulators 29 [14], or recently by a deformable phase plate modulator [15], whereby the employed phase 30 modulations lead to desired beam shapes in the focal plane. Such spatially tailored beam 31 profiles were employed for deep tissue focussing [16], high resolved spatial imaging [17], 32 and microstructuring [18]. First attempts were undertaken to combine temporal and spatial 33 shaping [19–22], because these entirely tailored pulses enable to steer photo-induced processes 34 concurrently in time and space. 35

Here, simultaneously temporally and spatially polarization shaped laser pulses are employed 36 to achieve two-photon excited fluorescence of dyes. The studies are performed with a particularly 37 developed shaping setup first including a temporal pulse shaper and then a spatial modulator 38 having a deformable phase plate. Initially, phase scans for different polarization conditions 39 are presented, which indicate an increased fluorescence contrast for polarization shaped pulses 40 compared to linear polarized pulses. Then, depth-dependent measurements are conducted by 41 additionally using the deformable phase plate modulator. These experiments are analyzed 42 regarding fluorescence contrasts depending on polarization states. The described pulse shaping 43 technique exhibits novel perspectives for biophotonic applications. 44

45 2. Experimental

The experimental setup is schematically presented in Fig. 1. A Nd: YVO₄ laser (Verdi V, Coherent, 46 Inc.) pumps a titanium sapphire laser oscillator (MIRA, Coherent Inc) having an average power 47 of 500 mW, a pulse duration of 70 fs, and a repetition rate of 76 MHz. The central wavelength is 48 adjusted to 800 nm with a full width at half maxium of approximately 25 nm. The generated 49 fs-laser pulses pass a 4f pulse shaper having a computer controlled liquid crystal light modulator 50 (SLM 640, Cambridge Research Instruments) with optical axes at $\pm 45^{\circ}$ to the horizontal which 51 allows for shaping the phase and polarization of the laser light. A half-wave plate is placed after 52 the modulator in order to rotate the light field by 45° that the light components modified by 53 the liquid crystals are turned to horizontal and vertical direction, respectively. Such temporally 54 shaped laser pulses are then guided through a deformable phase plate wavefront modulator (Delta 55 7, Phaseform GmbH) being used as a 2D spatial beam shaper in this experimental setup. The 56 deformable phase plate has an electrode array within the active area having a diameter of 10 mm. 57 It is an active device designed to locally change the optical path length, the geometric length 58 times refractive index, traveled by light. This transmissive device is less than 1 mm thick and 59 consists of a sealed liquid filled volume with a flexible polymeric membrane on one side and 60 a rigid, transparent glass substrate on the other. The volume between the membrane and the 61 substrate is filled with a high-refractive-index liquid. The conductive membrane is pulled toward 62 the substrate when a voltage is applied to the electrodes. This actuation displaces the liquid, and 63 changes the effective optical path length of light that refracts through the wavefront modulator. 64 All polarization components can equally be shaped with this modulator. 65



Fig. 1. Schematic experimental setup including a fs-laser (Ti:Sa), a temporal pulse shaper with gratings and a liquid crystal light modulator, and a spatial beam shaper utilizing a 2D deformable phase plate. Furthermore, a cuvette for fluorescence excitation and optical components for detection are depicted. The laser beam is first guided through the temporal pulse shaper, then through the spatial pulse shaper, and finally it is focused into a cuvette. After passing collecting lenses the fluorescence is detected in perpendicular directions, either by cameras or a photomultiplier.

The laser beam profile is enlarged by a telescope before the spatial shaper in order to use the entire active range of the deformable phase plate. A focussing lens (f = 60 mm) is located behind it to focus the laser beam into a cuvette for generating spatially modulated focal profiles.

A dielectric mirror for wavelengths around 800 nm is further placed before the cuvette in order 69 to reflect the laser beam into the cuvette, while the fluorescence light passes the mirror (see 70 Fig. 1). This top view geometry enables simultaneous excitation and detection on the surface 71 normal. The cuvette is filled with the laser dyes coumarin 102 (c102) or coumarin 120 (c120) 72 (received from Radiant Dyes) dispersed in highly viscous glycerol. The generated two-photon 73 excited fluorescence passes an IR glass filter (BG 39) and is directed on cameras (Logitec) or 74 a photomultiplier (Hamamatsu). The IR bandpass filter [23] is used to reduce the laser stray 75 light in order to measure only the fluorescence signals. The fluorescence images are taken in 76 side and top view configuration where the detectors face the side or front surface of the cuvette, 77 respectively. Before starting the measurements, suitable phase functions are inscribed on the 78 temporal modulator to achieve a short pulse resulting in a constant phase which is a prerequisite 79 for the phase-controlled measurements. 80

81 3. Results

⁸² 3.1. Two-photon excited fluorescence by polarization tailored pulses

The present pulse shaper setup allows for generating polarization shaped pulses with independently 83 tailored perpendicular polarization components. This is conducted by separately inscribing 84 the corresponding voltages on the two liquid crystal arrays of the temporal modulator which 85 have perpendicularly arranged optical axes and leads to independent phase shaping of the 86 two perpendicular polarization components. The half-wave plate located behind the temporal 87 modulator turns the light field by 45° in order to orient the light components shaped by the 88 liquid crystals to the horizontal and vertical direction, respectively. This is done to prevent the 89 second polarization dependent grating from changing the angle between the shaped polarization 90 components. The subsequent spatial modulator does not modify the polarization directions. The 91 two-photon excited fluorescence for such pulses is recorded for coumarin 102 and coumarin 120, 92 respectively. 93



Fig. 2. Two-photon excited fluorescence signals by scanning the excitation phase center wavelength are presented for coumarin 102 (red) and coumarin 120 (blue) generated for perpendicularly polarized subpulses. Third order phase scans of the first subpulse were performed and the fluorescence was recorded behind a horizontally oriented polarizer. A red-shift of the curve for coumarin 102 compared to coumarin 120 is obtained.

The two perpendicularly oriented polarization components can further be phase-shaped to obtain selective two-photon excitations. Antisymmetric third order phase functions are used to receive spectrally narrow two-photon excited fluorescence maxima due to constructive ⁹⁷ interference, similar as described in [10]. Phase functions $\phi(\omega) = \frac{b_3}{6}(\omega - \omega_0)^3$ with a third order ⁹⁸ phase factor b_3 and differently tuned phase center frequencies ω_0 are independently inscribed on ⁹⁹ the perpendicular polarization components of the temporal pulse shaper. The phase center value ¹⁰⁰ is thereby stepwise moved from lower to higher wavelengths by inscribing the corresponding ¹⁰¹ voltage values on the modulator. Constructive interference close to ω_0 results in spectrally narrow ¹⁰² two-photon maxima and enables two-photon excited fluorescence contrast enhancement.

Fig. 2 shows the two-photon excited fluorescence of coumarin 102 and coumarin 120, 103 respectively, for a wavelength scan of $\lambda_0 = 2\pi c/\omega_0$ with a phase factor of $b_3 = 5 \cdot 10^5$ fs³ of 104 the first horizontally polarized subpulse. This is done in the top-view geometry, where the 105 photomultiplier faces the cuvette wall of the laser entrance (see Fig. 1). A horizontally oriented 106 polarizer is thereby placed in the fluorescence path. A red-shift of coumarin 102 signals compared 107 to coumarin 120 can be observed, where a value of 1.8 ± 0.8 nm was found for the red-shift by 108 calculating the difference of the arithmetic means of the two curves. This can be rationalized 109 by a higher two-photon absorption cross-section of coumarin 102 relative to coumarin 120 by 110 proceeding to larger wavelengths. Contrast curves can be determined from the scan data. 111

112 3.2. Contrast enhancement with polarization shaped pulses

The two-photon excited fluorescence characteristics by polarization-shaped laser pulses is 113 investigated for different polarization conditions. Thereto, a polarizer is placed at different 114 positions and the results are compared. The fluorescence yield is influenced by antisymmetric 115 third order phase functions inscribed on the temporal pulse shaper which lead to constructive 116 two-photon excited fluorescence close to the antisymmetry wavelength of the phase function. 117 The applied laser pulses consist of two perpendicularly polarized subpulses with a time delay 118 of 400 fs. The first horizontally polarized subpulse exhibits a third order phase with a phase 119 factor of $b_3 = 5 \cdot 10^5$ fs³ and the antisymmetry point will be spectrally scanned. The second 120 vertically polarized subpulse has a third order phase with a phase factor of $b_3 = 5 \cdot 10^4$ fs³, an 121 antisymmetry wavelength of $\lambda_0 = 790$ nm, and is kept constant during the measurement. 122

In Fig. 3 the center wavelength of the third order phase is scanned for the first subpulse. The 123 contrasts $c = (I_{c102} - I_{c120})/(I_{c102} + I_{c120})$ of third order phase center wavelength scans with 124 different polarization adjustments are presented, where I_{c102} and I_{c120} indicate the fluorescence 125 intensities of coumarin 102 and 120, respectively. The measurements were performed with 126 horizontally or vertically oriented polarizer in the fluorescence path and with a polarizer placed 127 in the laser excitation path, oriented at 45° to the horizontal, which delivers a linearly polarized 128 laser pulse with two delayed sub pulses. The recorded data allow for a comparison between 129 polarization-shaped and linear polarized laser pulses. The maximal contrast difference of the 130 data with vertical polarizer ($\Delta c = 0.12$) is smaller than for the linearly polarized case ($\Delta c = 0.2$), 131 whereas the contrast difference for the horizontal polarizer is larger ($\Delta c = 0.3$). This indicates 132 the dependency on polarization and proves that the contrast difference can be enhanced by 133 polarization shaped pulses. 134

A further experiment was performed in order to explore the polarization-dependent contrast. 135 Two perpendicularly polarized subpulses were generated, where the first exhibits a phase factor of 136 $b_3 = 5 \cdot 10^5$ fs³ and an antisymmetry wavelength of $\lambda_0 = 812$ nm, and the second with the same 137 properties as explained above. The left hand inset from Fig. 3 shows a schematic 3D-image of 138 the polarization shaped pulse. Two-photon excited fluorescence measurements were performed 139 for both dyes, whereby the polarizer in the fluorescence path was turned in order to receive 140 polarization-dependent data. The right hand inset of Fig. 3 displays the two-photon excited 141 fluorescence contrast between coumarin 102 and coumarin 120 by tuning the polarizer angle 142 relative to the horizontal. A maximum at 0° and a minimum close to 90° is obtained. This 143 indicates that coumarin 102 is predominantly excited by the horizontally polarized subpulse and 144 coumarin 120 by the vertically polarized subpulse. It is in agreement with the phase scan results 145



Fig. 3. Contrast curves between coumarin 102 and 120 for third order phase center wavelength scans with different polarization conditions. The measurements were conducted for polarization-tailored pulses with two subpulses, whereby a horizontally (red) or vertically (blue) oriented polarizer is located in the fluorescence path. By placing a polarizer in the laser path, contrast data of a linearly polarized laser pulse (black) were generated. A polarization dependence of the maximal contrast difference is obtained and an enhanced contrast difference for polarization-shaped pulses is received. The left hand inset shows a schematic 3D-image of the polarization shaped pulse. The right hand inset displays the two-photon excited fluorescence contrast by rotating the polarizer in the fluorescence path. A maximum at the horizontal and a minimum close to the vertical direction is observed which indicates that coumarin 102 is selectively excited by the horizontally polarized subpulse and coumarin 120 by the vertically polarized subpulse.

from Fig. 3 and shows that one dye is selectively excited in one polarization direction and the
 other dye in the other polarization direction. This can be regarded as a polarization enhanced
 two-photon excited fluorescence contrast.

3.3. Spatially shaped fluorescence structures generated by a deformable phase plate modulator

Spatially tailored laser profiles can be generated by the deformable phase plate modulator 151 located behind the temporal pulse shaper. Computer-controlled voltage values are applied 152 on the individual modulator elements to modify their thickness, and Zernike polynomials are 153 used for phase control which leads to spatial modulation in the focal plane of the convex lens 154 behind the modulator. Zernike polynomials are used in optics e.g. to describe wave front 155 aberrations. They have a polynomial radial and an azimuthal angular part and are separated 156 into even $Z_n^m(\rho, \phi) = R_n^m(\rho) \cos(m\phi)$ and odd $Z_n^{-m}(\rho, \phi) = R_n^m(\rho) \sin(m\phi)$ functions, with the 157 radius ρ and the azimuthal angle ϕ . In the following the value multiplied with the Zernike 158 polynomial will be called prefactor. 159

Fig. 4 shows the astigmatic influence of different Zernike polynomials. Solely for this experiment, a screen is placed at the position of the cuvette and photos of the beam profiles were taken. The spot can be e.g. elliptically shaped by Z_2^2 polynomials (Fig. 4 (b) and (c)) or by Z_2^{-2} polynomials (Fig. 4 (d) and (e)). It can be observed that the profiles are elliptically shaped with



Fig. 4. Beam profiles for different Zernike polynomials. The images display the light intensities on a screen without spatial shaping (a) and with astigmatic spatial phase shaping by a Z_2^2 -Zernike polynomial with the prefactors 5 (b) and -5 (c), and by a Z_2^{-2} -Zernike polynomial with the prefactors 5 (d) and -5 (e).

horizontal (b), vertical (c), $+45^{\circ}$ (d), and -45° (e) main axes which is in agreement with the expected astigmatic outcomes.

Particularly, the two-photon excited fluorescence structure can be modified by forming the beam profile. This is demonstrated by moving the focal depth position with Z_2^0 -Zernike polynomials. Fluorescence images are presented in Fig. 5 where two adjacent cuvettes are observed from the side with coumarin 102 filled in the left (rear) and coumarin 120 in the smaller right (front) cuvette. The green fluorescing focus is located in the rear cuvette by using a prefactor of -5 and the blue fluorescing focus in the front cuvette with a prefactor of 5. These measurements were conducted with short pulses having a constant spectral phase.

173 3.4. Depth-dependent two-photon excited fluorescence of polarization shaped pulses

A good contrast between different substances on small distances is important for fluorescence 174 imaging. Here, this goal will be approached by simultaneous temporally and spatially tailored 175 pulses in order to perform selective depth-dependent two-photon excitations of different dyes. 176 Hence, a depth-dependent contrast measurement with polarization-shaped pulses is conducted. 177 The employed pulses exhibit a first horizontally polarized subpulse with a third order phase 178 factor of $b_3 = 5 \cdot 10^5$ fs³ and an antisymmetry wavelength of $\lambda_0 = 812$ nm, and a 400 fs delayed 179 second vertically polarized subpulse with a third order phase factor of $b_3 = 5 \cdot 10^4$ fs³ and an 180 antisymmetry wavelength of $\lambda_0 = 790$ nm. The polarized pulse focus is moved by about 2 mm 181 (see Fig. 5) from one cuvette to the other by tuning the prefactor of the Z_2^0 -Zernike polynomial, 182 and the fluorescence is measured on the front side. The front cuvette is filled with coumarin 120 183 and the rear cuvette with coumarin 102. This order is favorable because the lower frequency 184 fluorescence from the rear cuvette will not be absorbed by the front cuvette, whereas absorption 185 would occur the other way around. 186

¹⁸⁷ The experiment is again conducted with a horizontally or vertically oriented polarizer in the

¹⁸⁸ fluorescence path and with a polarizer in the laser path, oriented at 45° to the horizontal, the



Fig. 5. Modification of the focal depth position by the Z_2^0 -Zernike polynomial. Two adjacent cuvettes are visible from the side with coumarin 102 filled in the left (rear) and coumarin 120 in the smaller right (front) cuvette. The focus is located in the rear cuvette by using a prefactor of -5 (a) and it is moved to the front cuvette by a prefactor of 5 (b).

latter leading to a linearly polarized laser pulse. The two-photon excited fluorescence data for the 189 three different polarization conditions are shown in Fig. 6. The dip in the curves close to zero can 190 be explained by reduced fluorescence due to the cuvette walls. At these focus positions both dyes 191 may contribute to the signal since the focal range is partially present in both cuvettes. The lower 192 intensity of the vertical compared the horizontal component can be explained by the reduced 193 reflection of the second grating in the temporal modulator for vertical polarization. Particularly 194 for the horizontally oriented polarizer, an enhanced signal ratio is obtained for the rear to the 195 front cuvette compared to the linearly polarized pulse and a reduced ratio is observed for the 196 vertically oriented polarizer. The corresponding contrast for the horizontal case of 0.06 is hence 197 increased compared to the contrast of 0.03 for the linearly polarized case, and the contrast for 198 the vertical case is decreased to -0.02. This demonstrates that the spatially dependent contrast 199 can be enhanced by polarization shaped laser pulses. Since these polarization tailored laser 200 pulses can well be spatially moved by the deformable phase plate modulator, three-dimensional 201 scans of probes can be performed. The spatial modification without changing the polarization 202 state is beneficial compared to spatial shaping by liquid crystal modulators, where only a single 203 polarization direction can be modulated. 204

205 4. Conclusion

Spatially and temporally polarization shaped laser pulses were applied for two-photon excited 206 fluorescence of dyes. The pulses were shaped including different polarization directions which 207 means that they exhibit a temporal profile, where the polarization direction is changed during the 208 pulse. These investigations were conducted by using a combination of a temporal pulse shaper 209 and a novel deformable phase plate spatial modulator. Characteristic third order phase scans 210 were recorded for different polarization directions. They exhibit an enhanced two-photon excited 211 fluorescence contrast difference for polarization shaped pulses compared to linear polarized 212 pulses. Moreover, different spatially modified beam profiles were presented. Particularly, 213 depth-dependent two-photon excited fluorescence experiments were performed by utilizing the 214 deformable phase plate shaper. The measurements feature an increased spatial fluorescence 215 contrast for specific polarization-shaped pulses compared to linearly polarized pulses. The 216 polarization-tailored laser pulses, which lead to increased contrast differences, can hence spatially 217



Fig. 6. Depth-dependent fluorescence for differing polarization conditions. The measurement is recorded with a horizontally (red) or vertically (blue) oriented polarizer in the fluorescence path and a polarizer in the laser path, oriented at 45° to the horizontal (black), which results in a linearly polarized pulse. The two-photon excited fluorescence is measured for the three polarization conditions. In horizontal direction an enhanced signal ratio is obtained for the rear to the front cuvette compared to the linearly polarized pulse and to the vertical direction which shows that the spatially dependent contrast can be increased for polarization-tailored pulses.

²¹⁸ be modified by the deformable phase plate modulator. The described spatial modification for

all polarization directions is favorable compared to spatial shaping by liquid crystal modulators,

whereby only one polarization component can spatially be modulated. Since the deformable

phase plate modulator is relatively small it can well be integrated into microscopes in order to

²²² improve the image by adaptive optics. The polarization contrast technique developed here can be

- employed for three-dimensional scans of biological probes and has a high potential for imaging
- 224 applications.

Acknowledgements. The authors thank Prof. Dr. W. Kuch and the team of Phaseform GmbH for supporting this experimental study.

227 Disclosures. The authors declare no conflicts of interest.

Data Availability Statement. Data underlying the results presented in this paper are not publicly available at this time but may be obtained from the authors upon reasonable request.

230 References

- A. Assion, T. Baumert, M. Bergt, T. Brixner, B. Kiefer, V. Seyfried, M. Strehle and G. Gerber, "Control of chemical reactions by feedback-optimized phase-shaped femtosecond laser pulses," Science. 282, 919-922 (1998).
- T. Brixner and G. Gerber, "Quantum Control of Gas-Phase and Liquid-Phase Femtochemistry," ChemPhysChem 4, 418-438 (2003).
- G. Vogt, G. Krampert, P. Niklaus, P. Nuernberger and G. Gerber, "Optimal Control of Photoisomerization," Phys. Rev. Lett. 94, 068305 (2005).
- A. Lindinger, C. Lupulescu, M. Plewicki, F. Vetter, A. Merli, S. M. Weber and L. Wöste, "Isotope Selective Ionization by Optimal Control Using Shaped Femtosecond Laser Pulses," Phys. Rev. Lett. 93, 033001 (2004).
- M. Aeschlimann, M. Bauer, D. Bayer, T. Brixner, F. J. Garcia de Abajo, W. Pfeiffer, M. Rohmer, C. Spindler and F. Steeb, "Adaptive subwavelength control of nano-optical fields," Nature 446, 301-304 (2007).
- W. Wohlleben, T. Buckup, J. L. Herek and M. Motzkus, "Coherent Control for Spectroscopy and Manipulation of Biological Dynamics," ChemPhysChem 6, 850-857 (2005).
- L. Polachek, D. Oron, and Y. Silberberg, "Full control of the spectral polarization of ultrashort pulses," Opt. Lett. 31, 631-633 (2006).
- 8. T. Brixner and G. Gerber, "Femtosecond polarization pulse shaping," Opt. Lett. 26, 557-559 (2001).

- F. Weise and A. Lindinger, "Full parametric pulse shaping in phase, amplitude, and polarization using an effective four-array modulator," Appl. Phys. B 101, 79-91 (2010).
- 248 10. V. V. Lozovoy, I. Pastirk, K. A. Walowicz and M. Dantus, "Multiphoton intrapulse interference. II. Control of twoand three-photon laser induced fluorescence with shaped pulses," Chem. Phys. **118**, 3187-3196 (2003).
- 11. S. Perry, R. Burke and E Brown, "Two-Photon and Second Harmonic Microscopy in Clinical and Translational Cancer Research," Ann. Biomed. Eng. 40, 277-291 (2012).
- W. Denk, J. H. Strickler and W. W. Webb, "Two-photon laser scanning fluorescence microscopy," Science 248, 73-76 (1990).
- 13. C. Maurer, A. Jesacher, S. Bernet and M. Ritsch-Marte, "What spatial light modulators can do for optical microscopy,"
 Laser Photonics Rev. 5, 81-101 (2011).
- 14. N. Sanner, N. Huot, E. Audouard, C. Larat and J.-P. Huignard, "Programmable focal spot shaping of amplified femtosecond laser pulses," Opt. Lett. 30, 1479-1481 (2005).
- K. Banerjee, P. Rajaeipour, C. Ataman, and H. Zappe, "Optofluidic adaptive optics," Appl. Opt. 57, 6338-6344 (2018).
- 16. A. Tanabe, T. Hibi, S. Ipponjima, K. Matsumoto, M. Yokoyama, M. Kurihara, N. Hashimoto and T. Nemoto,
 "Correcting spherical aberrations in a biospecimen using a transmissive liquid crystal device in two-photon excitation
 laser scanning microscopy," J. Biomed. Opt. 20, 101204 (2015).
- 17. S. Hell and J. Wichmann, "Breaking the diffraction resolution limit by stimulated emission: stimulated-emission depletion fluorescence microscopy," Opt. Lett. 19, 780-782 (1994).
- 18. N. Sanner, N. Huot, E. Audouard, C. Larat and J.-P. Huignard, "Direct ultrafast laser micro-structuring of materials
 using programmable beam shaping," Opt. Lasers Eng. 45, 737-741 (2007).
- T. Feurer, J. C. Vaughan, R. M. Koehl and K. A. Nelson, "Multidimensional control of femtosecond pulses by use of
 a programmable liquid-crystal matrix," Opt. Lett. 27, 652-654 (2002).
- 269 20. D. J. McCabe, A. Tajalli, D. R. Austin, P. Bondareff, I. A. Walmsley, S. Gigan and B. Chatel, "Spatio-temporal focusing of an ultrafast pulse through a multiply scattering medium," Nature Communications 2, 447 (2011).
- 21. O. Katz, E. Small, Y. Bromberg and Y. Silberberg, "Focusing and compression of ultrashort pulses through scattering
 media," Nature Photonics 5, 372-377 (2011).
- 273 22. T. Feurer, J. C. Vaughan and K. A. Nelson, "Spatiotemporal coherent control of lattice vibrational waves," Science
 274 299, 374-377 (2003).
- 275 23. Schott AG, "Product sheet of the optical bandpass filter BG39," https://www.schott.com/shop/
- advanced-optics/en/Matt-Filter-Plates/BG39/c/glass-BG39.