



Non-Mycorrhizal Fungal Presence Within Roots Increases Across an Urban Gradient in Berlin, Germany

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Symbioses between plants and fungi are important in both promoting plant fitness and maintaining soil structure. The ways in which these relationships change across an urban gradient is subject to debate. Here we measured root colonisation including the presence of arbuscular mycorrhizal fungi, non-mycorrhizal fungi, and root hair presence. We found no evidence of changes in levels of arbuscular mycorrhizal fungal colonisation across an urban gradient, colonisation levels being driven instead by plant community. However, we did observe an increase in non-mycorrhizal fungal colonisation in association with increasing urbanity. Additionally, we observed an urban-related increase in root hair presence. Using principal component analysis we were able to provide strong evidence for these patterns being driven by an “urban syndrome”, rather than soil chemistry. Our findings have important implications for the wider understanding of abiotic stresses on fungal endophyte presence and shed light on the impacts of urbanity upon plant roots.

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INTRODUCTION

We live in the midst of the anthropocene, an age defined by the profound impact of humanity upon the Earth. Urban centres are the pinnacles of human impact upon the planet and contain a concentration of global-change factors not seen elsewhere (heavy metal pollutants, salinity, nitrogen deposition, heat islands and altered rainfall, to name but a few; Ziska et al., 2003; Carreiro and Tripler, 2005). These landscapes are by no means solely human domains and their ecology has been the target for several decades of research, largely due to the wide array of ecosystem services that are provided by their non-human components (Luederitz et al., 2015). A general rule appears to be that urbanisation results in the loss of animal diversity and can either increase or decrease plant diversity depending on the intensity of urbanisation (McKinney, 2008). However, the basis of all plant, and consequently animal, diversity is the soil. Urban soils have variously been described as compressed (Yang and Zhang, 2011), polluted (Hanfi et al., 2020) and lacking aggregates (Whitehead et al., 2021), expressing increased pH (Asabere et al., 2018) and salinity (Dmuchowski et al., 2021). As such, the ability of symbiotic fungi to buffer the negative impacts of urbanity upon plant hosts makes them a vital component of maintaining urban greenspaces. Urban ecosystems have also been described as hotspots for biological invasions (Gaertner et al., 2017). Fungal symbionts play a variety of roles in plant invasions, with some evidence suggesting that invasive plants are more likely to be facultatively symbiotic than native competitors, or at least not dependent on specific symbionts (Vogelsang and Bever, 2009; Hempel et al., 2013) and plants limited to specific symbionts appear to have their

invasive potential constrained (Catford et al., 2009). Additionally, some symbioses are thought to help native plants maintain a competitive advantage over invasive species (Pringle et al., 2009). These interactions appear to be spatially variable, with an apparent split between North America and Europe (Pringle et al., 2009).

One of the most globally significant groups of plant symbionts are the arbuscular mycorrhizal (AM) fungi, which have been demonstrated to not only boost plant fitness (Begum et al., 2019) but also play a key role in the formation and stabilisation of soil aggregates (Piotrowski et al., 2004; Leifheit et al., 2014; Morris et al., 2019). Given the importance of AM fungal symbionts, a substantial amount of effort has been put into understanding the factors that influence AM fungi-plant symbioses. Many of these factors are also known to be impacted by urbanisation. For example; elevated nitrogen (Lovett et al., 2000) and phosphorus (Zhang et al., 2001; Liu et al., 2016) concentrations have been demonstrated to both occur in urban ecosystems and result in a down-regulation of AM fungal symbioses by plants (Bonneau et al., 2013; Nouri et al., 2014). Alterations in soil pH have also been observed to alter AM fungal colonisation, with acidity appearing to limit AM fungal colonisation (Clark, 1997; Coughlan et al., 2000). Despite the influence of acid rain, the overwhelming levels of alkaline compounds provided by concrete dust generally result in a net-alkaline pH for urban soils (Newbound et al., 2010). This elevated pH reduces the availability of heavy metals and thus partially counteracts the high levels of heavy-metal contamination often observed at hotspots of anthropogenic activity (Xian and Shokohifar, 1989). AM fungal colonisation has been reported as being reduced in heavy-metal contaminated soils (Ferrol et al., 2016), although the role of heavy metals in the plant-AM fungi-soil system is likely to be complex, depending on the species involved. Indeed, some AM fungal species confer greater heavy metal tolerance to plants, whilst others can result in plants accumulating heavy metals (Ferrol et al., 2016).

Previous research has sought to explore the ways in which urban systems shape AM fungal diversity. Cousins et al. (2003) investigated AM fungal spore diversity across different land uses in Arizona and observed a shift in community composition between urban sites and agricultural sites. Research carried out in the same study system found that AM fungal colonisation of roots was significantly lower in urban desert preserves than in rural desert (Ontiveros-Valencia, 2009). A very thorough study on the same subject by Bainard et al. (2011) also found a similar pattern in Ontario. They lent significant weight to their conclusions by limiting their study to 26 tree species, of which 11 showed a significant reduction in colonisation in urban sites compared to rural sites. Wiseman and Wells (2005) also found a significantly higher level of AM fungal colonisation in *Acer rubrum* in forested sites compared to those in developed areas. However, a study in Michigan has reported no change in AM fungal colonisation rate in saplings across a gradient of urbanity (Tonn and Ibáñez, 2017).

AM fungi are, however, not the only fungi which colonise roots. There are a wide range of other fungal symbioses that occur, including ecto- and ericoid-mycorrhiza and an array of

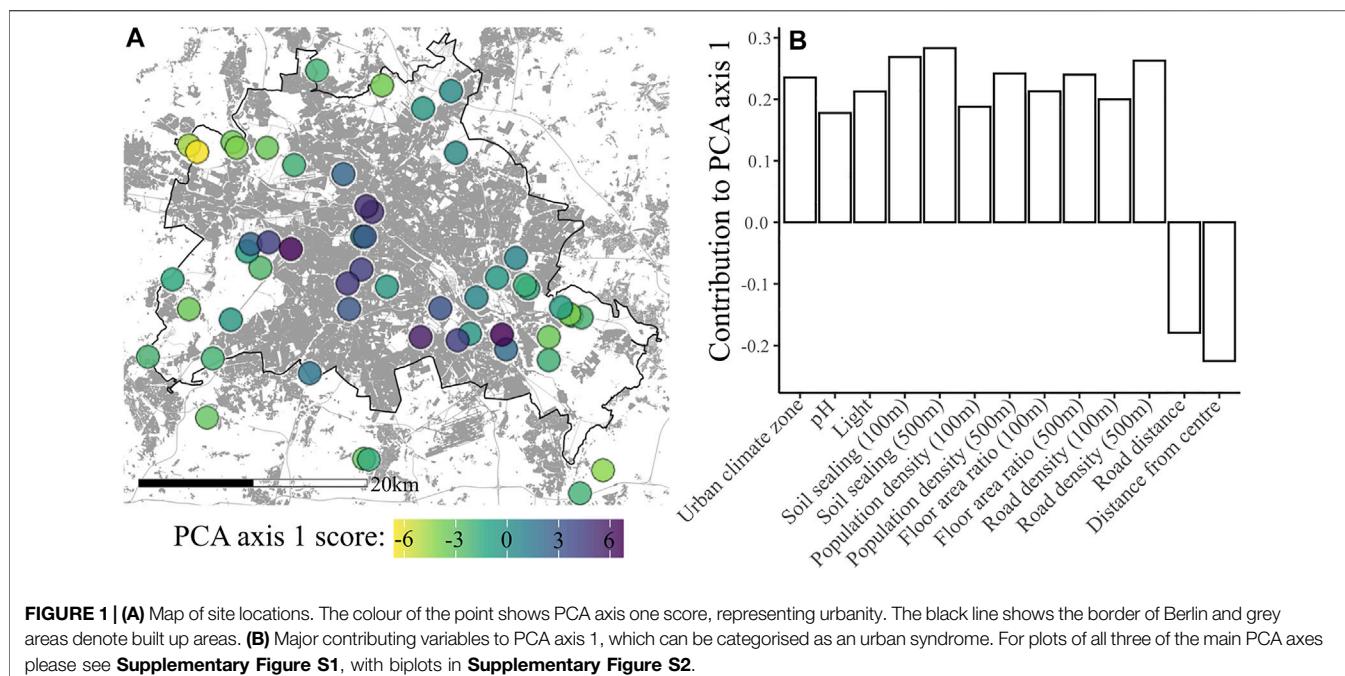
non-mycorrhizal symbioses, the most abundant of which are the dark septate endophytes. The position of these non-mycorrhizal fungi on the “parasitism-mutualism continuum” (Mandyam and Jumpponen, 2015) is varied and often hard to discern, with some evidence for it changing according to the species, or even life stage, of the host plant involved (Berthelot et al., 2019). The benefits to plants of these mutualistic non-mycorrhizal symbioses range from increased heavy metal tolerance to accelerated growth rate (Mandyam and Jumpponen, 2005). The position of a particular symbiosis on this continuum may also be impacted by the environment (Berthelot et al., 2017), but this is debated (Kia et al., 2018). Colonisation rates do, however, appear to vary significantly in response to environmental conditions (Rillig et al., 1998) and to be most common in abiotically stressed environments (Mandyam and Jumpponen, 2005).

Here, we carried out an assessment of AM fungal and non-AM fungal colonisation from grassland sites across an urbanity gradient in Berlin, Germany. Through limiting the study to one habitat type and using Principal Component Analysis to extract three major axes of variation from a substantial database of environmental data, we were able to isolate the impacts of an urban ‘syndrome’, here referred to as urbanity, upon root colonisation and morphology. By using a gradient of sites rather than a paired-sample design we were able to use linear statistical models and have a greater degree of control over covarying environmental variables.

METHODS

Berlin is a temperate central European city with around 3.5 million inhabitants. It is surrounded by the significantly more rural state of Brandenburg. Berlin’s soil texture ranges from sand to sandy loam with a pH range of 4.1–7.5. Fifty-four grassland sites distributed across Berlin and into Brandenburg were sampled between 30th June and 13 August 2020 for this study. These 16 m² sites were part of the CityScapeLabs research platform, a network of grassland sites situated along an urban-rural gradient (von der Lippe et al., 2020). Root samples were taken from three locales within each site. Sampling locales were selected which reflected the diversity of plants present in each site. A random selection of roots was taken from each locale. Samples were stored in paper bags before being dried at 60°C overnight. Samples were subsequently stored at 4°C until processing. Only roots that were springy and had an intact cortex and tissue structure were selected for further analysis. Samples were bleached for 30 min in KOH, washed, acidified with 1% HCl for 12 min, and stained using 0.05% Trypan Blue in lactoglycerol for 40 min (Boedijn, 1956). Bleaching, acidification and staining were all performed in a 90°C water bath. Roots were subsequently mounted on slides in lactoglycerol.

AM fungal colonisation was quantified under ×200 magnification in accordance with McGonigle et al. (1990). At ~100 intersects per slide any AM fungal structure was recorded in a binary fashion (presence/absence), with additional binary counts of arbuscules, AM fungal vesicles and AM fungal coils. Non-AM fungal structures and root hairs were also recorded in



the same way; we counted as non-AM fungal structures any fungal presence that was not attributable to AM fungi. Count data were used to calculate the percentage of presence for each variable.

Principal Component Analysis (PCA), performed using the ade4 package (Thiououlouse et al., 2018) in R (R Core Team, 2021), was used to extract three main axes of variation from a pre-existing database of environmental data for the sampled sites (von der Lippe et al., 2020). These three axes of data were found to relate to three ecologically relevant syndromes. The PCA axis scores, the age of sites (a binary variable representing pre-vs. post-1945 establishment as a grassland), and the day of the year that sampling took place (to control for seasonal effects) were used to create 10 multiple linear regression models to explain each colonisation/root trait. Akaike model selection, performed using the AICmodavg package (Mazerolle, 2020) in R, was then used to select the most appropriate model, taking into consideration the percentage of cumulative model weight. For a list of models included please see **Supplementary Table S1**.

A pre-existing dataset of plant communities was used to explore the impact of plant community composition on the traits measured in this study. This dataset consisted of the percentage-cover of 233 grassland plant species within each study site. Plant communities were plotted using non-metric multidimensional scaling (NMDS) with the root responses and the three PCA environmental axes plotted as contour lines, this was achieved using the vegan package (Oksanen et al., 2020) in R. These data were used to qualitatively assess any possible relationships between plant community, environmental variation and root traits. Responses which demonstrated relationships with plant community were selected for further investigation. A series of multiple linear regression models were created which included the axis scores of the correlating NMDS

axis, which functioned as a proxy for plant community. Akaike model selection was then used to select the most appropriate multiple linear regression model.

RESULTS

The PCA of environmental variables created three main axes of variation that could be categorised as 1) urbanity (**Figure 1**), 2) soil chemistry and water content, and 3) site connectivity and pH. For a full list of variable loadings see **Supplementary Table S1**. Akaike model selection found that the most appropriate model for each of the response variables varied. For the percentage of AM fungal presence, arbuscules, vesicles (log transformed) and coils (log transformed) a multiple linear regression model containing PCA axes one and two was found to be appropriate. However, for the percentage of non-AM fungal presence and root hairs a single linear regression using only PCA axis 1 as a predictor was a more appropriate model. For the full output of Akaike model selection please see **Supplementary Table S3**.

We found that only two variables exhibited strong evidence for being driven by the environmental parameters included in their selected linear regression models. These variables were the presence of non-AM fungi and root hairs. Both of these responses showed a positive correlation with PCA axis one scores, meaning values increased at more urban sites (**Figure 2**). For the results of regression analyses please see **Supplementary Table S4**.

Through plotting plant communities using NMDS (**Supplementary Figure S4**), it was clear that plant community composition had a relationship with the levels of AM fungal, arbuscule and root hair presence. Plant communities did not

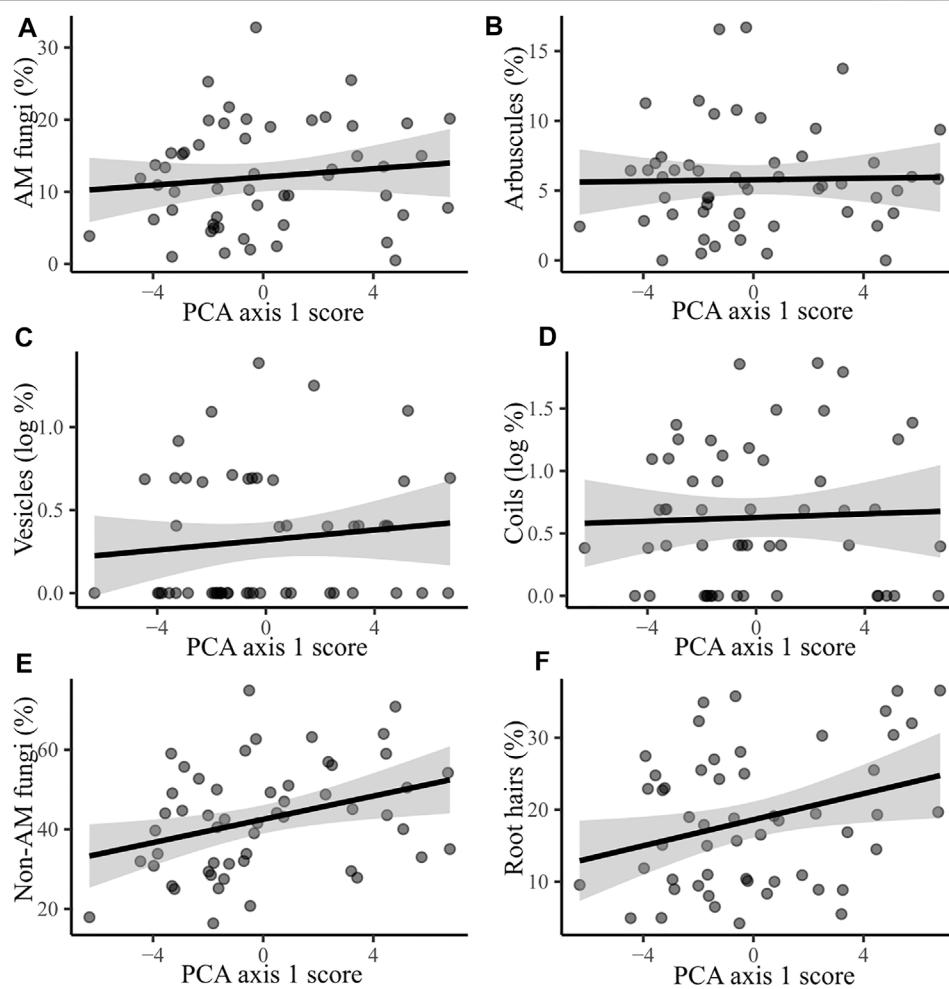


FIGURE 2 | The correlation between PCA axis one score (representing urbanity) and the percentage of root locations containing **(A)** AM fungi; $R^2 = 0.016$, $F = 0.821$, $p = 0.369$. **(B)** Arbuscules; $R^2 = 0.001$, $F = 0.026$, $p = 0.873$. **(C)** Vesicles (log transformed); $R^2 = 0.015$, $F = 0.787$, $p = 0.379$. **(D)** Coils (log transformed); $R^2 = 0.002$, $F = 0.088$, $p = 0.768$. **(E)** Non-AM fungi; $R^2 = 0.117$, $F = 6.763$, $p = 0.012$. **(F)** Root hairs; $R^2 = 0.094$, $F = 5.283$, $p = 0.026$. Shaded areas represent 95% confidence intervals. For scatter plots of responses to all three PCA axes, see **Supplementary Figure S3**.

appear to clearly shift with any of the environmental PCA axes although a trend with PCA axis 3, representing site connectivity, could be seen. Multiple linear regressions demonstrated that for AM fungi and arbuscules, presence was driven by plant community, whilst for root hairs the separate impacts of PCA axis one and plant community could not easily be disentangled, although there did not appear to be a significant interaction (Supplementary Table S6).

4 DISCUSSION

We found strong evidence for AM fungi-plant interactions, including the presence of arbuscules, being resilient to the effects of urbanity, being shaped instead by plant community. Contrastingly, our results provide strong evidence for non-AM fungal interactions with plant roots increasing with urbanity. Specifically, our study suggests that this change is due to an

intrinsic element of an urban syndrome, rather than just soil chemistry. Functional roles of these non-mycorrhizal fungi within the roots may have been varied. The non-AM fungal structures observed could have belonged to parasitic fungi, non-mycorrhizal mutualists, or AM fungi that were not exhibiting AM characteristics (Orchard et al., 2017). It is unlikely that they resulted from ectomycorrhiza, due to the absence of any distinguishable ectomycorrhizal structures being observed, or ericoid mycorrhiza, given the rarity of ericoid plants within Berlin's grasslands. If the increase in non-AM fungi was due to a reduction in AM structures being exhibited by AM fungi, we would also expect to see a proportional reduction in observed AM structures, which we did not.

The direction of the relationship between observed non-AM fungal presence and urbanity is rather surprising, because almost all generally-accepted urban-related variables are thought to result in reduced levels of fungal activity. Indeed, many environmental parameters often thought indicative of urban

systems were found to be contributors to PCA axes 2 and 3, rather than being collinear with variables such as population density and soil sealing in PCA axis 1. Heavy metals and increased pH (although pH also made a small contribution to PCA axis 1) are two such parameters, both of which are thought to negatively impact fungi in urban systems (Newbound et al., 2010). There are only a small number of urban-related parameters which may be expected to increase fungal activity, of these, the heat island effect is the only one that is represented in our dataset. The presence of an ‘urban climate zone’ parameter within PCA axis one makes temperature a strong candidate for being the cause of increased fungal activity. Additional factors not taken into account in this study such as the presence of animal faeces and remains have previously been demonstrated to increase fungal presence (Sagara, 1995; Watling, 1997; Newbound et al., 2010). Within our dataset graveyards accounted for some of the sites with highest PCA axis one scores, however, the sampling locations within these were not directly associated with graves and in one case was in a section of the graveyard that had been cleared for the creation of the Berlin wall. Site usage, such as dog walking, could also potentially be an explanation for altered fungal presence. However, the unimportance of soil chemistry in our study suggests that soil fertilisation via faecal deposition or decomposition in graveyards is an unlikely causal factor.

Through our use of PCA we were able to separate urbanity from soil chemistry. Our study provides evidence that we cannot solely rely on soil chemistry research to explain plant-fungal interactions in urban systems. Previous research from the same study system has suggested that urban soils have lower levels of aggregate stability (Whitehead et al., 2021), our current study suggests that a lack of AM fungi, previously described as key causal agents in aggregate formation (Lehmann et al., 2020), is likely not the leading cause of the reduction in aggregate stability in this system.

The increase in root hairs with increasing urbanity is a novel result which lacks any obvious explanation. It is surprising that such a trait, so clearly linked to the acquisition of nutrients, correlates with the PCA axis representing urbanity rather than that which represents soil chemistry. The trade-offs involved in root morphology are driven by a complex economy of carbon partitioning, growth rate and mycorrhizal colonisation (Bergmann et al., 2020). Within our study system however, there is no apparent trade-off between root hair presence and AM fungal colonisation, indeed, the only observable trend in fungal activity is collinear with root hair presence. One possible explanation for a change in root hair presence is a change in plant community, evidence for such an explanation can be seen in the NMDS plots in **Supplementary Figure S4**. However, whilst plant community does indeed appear to have a significant role in determining root hair presence, the lack of any interaction between plant community and PCA axis one suggests that this is likely not the decisive link between root hair presence and urbanity, although it cannot be ruled out.

Our findings suggest that an intrinsic element of urbanity has ecologically significant impacts on fungal presence in plant roots. This increase in non-AM fungal colonisation may be as a result of higher parasite load or an up-regulation of non-mycorrhizal mutualisms. We also found a general lack of significant response in AM fungi colonisation to urbanity, supporting the previous observations of Tonn and Ibáñez (2017). Our results for AM fungal colonisation share the lack of response to urbanity seen in ectomycorrhizal symbioses (Hui et al., 2017). The increase in root hair presence with urbanity is also interesting as there did not appear to be any trade-off with fungal colonisation.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: Figshare: <https://doi.org/10.6084/m9.figshare.19248926.v1> and <https://doi.org/10.6084/m9.figshare.19276481.v1>.

AUTHOR CONTRIBUTIONS

JW, SH, and MR designed the research. JW conducted fieldwork and lab analyses, and wrote the first draft of the paper. All authors added to and edited the text.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fenvs.2022.888310/full#supplementary-material>

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