



# Plant Immune Memory in Systemic Tissue Does Not Involve Changes in Rapid Calcium Signaling

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Upon pathogen recognition, a transient rise in cytoplasmic calcium levels is one of the earliest events in plants and a prerequisite for defense initiation and signal propagation from a local site to systemic plant tissues. However, it is unclear if calcium signaling differs in the context of priming: Do plants exposed to a first pathogen stimulus and have consequently established systemic acquired resistance (SAR) display altered calcium responses to a second pathogen stimulus? Several calcium indicator systems including aequorin, YC3.6 or R-GECO1 have been used to document local calcium responses to the bacterial flg22 peptide but systemic calcium imaging within a single plant remains a technical challenge. Here, we report on an experimental approach to monitor flg22-induced calcium responses in systemic leaves of primed plants. The calcium-dependent protein kinase CPK5 is a key calcium sensor and regulator of the NADPH oxidase RBOHD and plays a role in the systemic calcium-ROS signal propagation. We therefore compared flg22-induced cytoplasmic calcium changes in *Arabidopsis* wild-type, *cpk5* mutant and CPK5-overexpressing plants (exhibiting constitutive priming) by introgressing the calcium indicator R-GECO1-mTurquoise that allows internal normalization through mTurquoise fluorescence. Aequorin-based analyses were included for comparison. Based on the R-GECO1-mTurquoise data, CPK5-OE appears to reinforce an “oscillatory-like” Ca<sup>2+</sup> signature in flg22-treated local tissues. However, no change was observed in the flg22-induced calcium response in the systemic tissues of plants that had been pre-challenged by a priming stimulus – neither in wild-type nor in *cpk5* or CPK5-OE-lines. These data indicate that the mechanistic manifestation of a plant immune memory in distal plant parts required for enhanced pathogen resistance does not include changes in rapid calcium signaling upstream of CPK5 but rather relies on downstream defense responses.

**Keywords:** calcium signature, systemic signaling, priming, R-GECO1, CPK5

## INTRODUCTION

Plants that experience a primary attack by microbial pathogens not only induce rapid local immune responses but are also able to build a more long-term systemic immune memory, the so-called systemic acquired resistance (SAR). Primed plants are sensitized toward a secondary infection and often induce faster and stronger immune reactions that ultimately impede pathogen growth. Defense priming requires an initial recognition of a pathogen (or conserved microbe-derived molecules termed pathogen-associated molecular patterns, PAMPs), the initiation of local immune reactions, a defense signal propagation from local to distal sites of the plant (systemic tissues), and the onset and maintenance of SAR involving phytohormone-dependent changes in gene expression and defense metabolite synthesis (Hake and Romeis, 2019; Hilker and Schmulling, 2019).

A transient change of the cytoplasmic calcium concentration ( $[Ca^{2+}]_{cyt}$ ) is one of the earliest events in plant cells upon pathogen recognition, and many studies applying different calcium indicators reported on distinct  $[Ca^{2+}]_{cyt}$  patterns in response to PAMPs, e.g., bacterial flg22 (Ranf et al., 2011; Maintz et al., 2014; Seybold et al., 2014; Thor and Peiter, 2014; Keinath et al., 2015; Aldon et al., 2018; Tian et al., 2020). Furthermore, calcium and calcium signaling being instrumental during defense signal propagation from a local attacked site to systemic parts of a plant has been observed as a spread of a calcium signals through the plant along the vasculature and beyond (Romeis and Herde, 2014; Kiep et al., 2015; Nguyen et al., 2018; Toyota et al., 2018; Shao et al., 2020; Fichman and Mittler, 2021). However, it is unknown whether  $[Ca^{2+}]_{cyt}$  changes contribute to SAR. Defense priming of SAR is mechanistically correlated to changes in gene expression including accumulation of the master transcription factor *SARD1*, on maintaining phytohormone salicylic acid (SA)-based transcriptional reprogramming, or on epigenetic modifications. But it is unclear whether and how calcium signaling contributes to the mechanism necessary to acquire an immune memory. Do plants that have experienced a first priming pathogen stimulus and have consequently established SAR display a different  $[Ca^{2+}]_{cyt}$  change pattern in response to perception and recognition of a second triggering pathogen stimulus?

Studies on immune-related  $[Ca^{2+}]_{cyt}$  changes have been performed in cell cultures, protoplasts, young seedlings, excised leaf disks or in epidermal peels with a focus on single guard cells. These were often investigated in response to direct pathogen contact or to purified PAMPs such as flg22, pep13, chitin, or liposaccharides. The degree and patterns of calcium signal changes were recorded over time employing genetically encoded calcium indicator (GECI) systems such as aequorin, cameleon YC3.6 or more recently GCaMP/R-GECO1 and their derived variants (Blume et al., 2000; Kwaaitaal et al., 2011; Ranf et al., 2011; Thor and Peiter, 2014; Keinath et al., 2015; DeFalco et al., 2017; Hilleary et al., 2020; Li et al., 2021). Also,  $[Ca^{2+}]_{cyt}$  changes have been employed as a signaling read-out in forward and reverse genetic screens to dissect the roles of various genes in plant defense (Ranf et al., 2011, 2012; Tian et al., 2019; Hilleary et al., 2020; Thor et al., 2020). However, none of these systems

have been employed for imaging of systemic calcium responses in the context of SAR. Such investigations need to be conducted in adult plants competent to mount SAR, where a selected leaf is treated by a local priming stimulus, and after a gap in time of 2 days, a distal leaf is exposed to a secondary stimulus, and the consequence in the induced calcium response is recorded. The challenge of such an approach is the fluorescence emission-based calcium imaging, which in principle would have to cover an entire “primed” leaf, but ideally at single cell resolution.

In plant cells, changes in the intracellular  $[Ca^{2+}]$  are decoded and transduced by calcium sensor proteins and their interaction partners, such as CaMs, CMLs, CBL-CIPKs and CPKs (McCormack et al., 2005; Batistič and Kudla, 2012; Aldon et al., 2018; Kudla et al., 2018; Shi et al., 2018; Bredow and Monaghan, 2019; Mohanta et al., 2019). Among these decoder proteins is the 34 member-containing gene family of calcium-dependent protein kinases (CPK in *Arabidopsis*), in which a calcium sensor and protein kinase effector domain are united within a single molecule (Liese and Romeis, 2013; Schulz et al., 2013; Simeunovic et al., 2016; Yip Delormel and Boudsocq, 2019). Several distinct CPK members have been characterized as positive (CPKs 1,2,4,5,6,11) or negative (CPK28) regulators during the local initiation of PAMP-induced immune responses, subsequent to receptor-mediated pathogen recognition (Boudsocq et al., 2010; Gao et al., 2013; Kadota et al., 2014; Monaghan et al., 2014, 2015; Bredow et al., 2021). Among these, CPK5 is a key signaling hub of immune signaling, which correlates with its unique high affinity for calcium ( $K_d \sim 100$  nM) for enzyme activation. This is near the resting  $[Ca^{2+}]_{cyt}$  of an un-challenged plant cell, thus rendering the enzyme highly responsive to small  $[Ca^{2+}]_{cyt}$  changes (Guerra et al., 2020). Besides its function in local basal immunity, CPK5 was additionally shown to contribute to a calcium- and NADPH-oxidase RBOHD-mediated calcium/ROS-based auto-propagating mechanism that is assumed to be crucial for the defense signal spread from local to distal parts of a plant (Dubiella et al., 2013; Hake and Romeis, 2019). Furthermore, CPK5 is required for priming of a *SARD1*-dependent systemic, long-term immune memory. Enhanced CPK5 signaling in CPK5-YFP overexpressing lines leads to an increased pathogen resistance status, with the plants displaying constitutive priming, manifested by an increase in *SARD1* gene expression, high levels of the defense metabolite *N*-hydroxy pipecolic acid (NHP), and of the phytohormone SA (Guerra et al., 2020). When challenged with a priming infection by an avirulent bacterial pathogen, and a second triggering infection with a virulent pathogen, these plants exhibited enhanced SAR with an almost complete block of pathogen proliferation (i.e., a display of “super-priming”). In contrast, in a *cpk5, cpk6* double mutant line, the lack of CPK5 and its close CPK homolog CPK6 lead to an increase in pathogen susceptibility, and when subjected to a priming and triggering context, plants could not be primed and were unable to mount SAR (Gao et al., 2013; Guerra et al., 2020). CPK5 is a key positive regulator in priming and promoting disease resistance. Here, we aim to address if lines differing in their priming status also display altered stimulus-induced  $[Ca^{2+}]_{cyt}$  changes upon a second (triggering) stimulus.

The GECI R-GECO1 has been successfully used in live cell imaging to resolve PAMP-triggered calcium transients with a high sensitivity but, due to its intensimetric fluorescence readout, suffered from a  $[Ca^{2+}]_{cyt}$  calculation bias if GECI levels differ between lines or during time course of experiments (Keinath et al., 2015; Waadt et al., 2017). To investigate systemic calcium responses and compare different lines in the context of priming in adult plants over a time course of 2 days, we chose an improved next-generation R-GECO1-mTurquoise (RGmT) system, where the reporter additionally incorporates as an internal mTurquoise (mT) fluorescent protein reference for normalization and validation of the GECI amount throughout the experiment (Waadt et al., 2017). The RGmT sensor was introduced into the CPK5-overexpression line *CPK5#7* and into *cpk5*, *rbohD*, and *fls2* mutants by crossing. We found that under conditions of enhanced CPK5 signaling, flg22 induces a more pronounced pattern into distinct peaks of  $[Ca^{2+}]_{cyt}$  changes in local tissue. By contrast, in a systemic leaf, the overall calcium pattern (signal amplitude, form, and timing) upon flg22 stimulation did not significantly differ between plants of different priming status – either through pre-exposure to flg22 priming stimulus, “super-priming” (through overexpressing *CPK5*) or reduced priming (*cpk5* background). These data indicate that the systemic immune signaling causal to enhanced pathogen resistance in SAR does not employ alterations in the early  $[Ca^{2+}]_{cyt}$  change but relies on downstream defense processes.

## MATERIALS AND METHODS

### Plant Material and Growth Conditions

*Arabidopsis thaliana* (Col-0) was used throughout this study. Seeds were either sown on 0.5 x Murashige and Skoog (MS) media containing 500 mg/L MES and vitamins (Duchefa, Netherlands), 1% [w/v] sucrose, 0.8% [w/v] phytoagar (Duchefa), pH 5.7 adjusted with KOH, or grown in soil. Seeds were stratified for 2 days at 4°C, and after transfer to individual pots or jiffy-7 soil (Jiffy Products, Norway), the plants were maintained under short-day conditions (8 h day light, 150  $\mu$ E, 20 – 24°C; 60% RH). Seedlings carrying constructs with fluorophores were screened for fluorescence of the respective fluorescence protein using a fluorescence stereo zoom microscope (Zeiss Axio Zoom.V16, Zeiss, Germany) before the transfer to individual pots. Plants were grown for a total of 6 weeks under short day condition. For aequorin-based calcium assays, seeds were first surface sterilized and stratified for 2 days at 4°C and grown in liquid MS medium for 8–10 days under long day conditions (16 h light, 8 h dark, 21°C). Flg22 and an inactive flagellin variant from *Agrobacterium tumefaciens* were synthesized on an in-house peptide synthesizer and used as previously described (Ranf et al., 2011).

### Generation of Transgenic Plants

For generating transgenic *A. thaliana* lines carrying R-GECO1-mTurquoise (RGmT) Columbia-0 wildtype plants were transformed by floral dip method. Plasmid pGGZ-RW253 (pUBQ10-N-decoy-R-GECO1-GSL-mTurquoise-tHSP18.2M-HygR) was generated *via* GreenGate cloning using previously

published modules (Lampropoulos et al., 2013; Waadt et al., 2017). Seeds of homozygous *RGmT* lines were harvested and crossed with the mutants *cpk5* (SAIL\_657C06), *rbohD* (SALK\_070610), and *fls2* (SALK\_062054) and the CPK5-overexpressing line *CPK5#7* that have been described before (Ranf et al., 2012; Dubiella et al., 2013). The generated lines were selected for BASTA (phosphinothricin, glufosinate ammonium) or hygromycin B resistance and fluorescence to obtain independent transformants. For aequorin luminescence measurements, wildtype plants expressing cytosolic p35S-apoaequorin (pMAQ2) were used (Knight et al., 1991). Seeds of a homozygous line were used to cross with the *cpk5* (SAIL\_657C06) and the CPK5-overexpressing line *CPK5#7*.

## Calcium Measurements

### R-GECO1 Based Calcium Imaging

#### Local Calcium Imaging

For sample preparation, 2 well chambered coverslips (IBIDI, Germany) were coated with a thin layer of medical adhesive (Ulrich Swiss, Switzerland). After 10 min of evaporation of volatile medical adhesive components excess medical adhesive was removed by washing three times with water. The leaf abaxial epidermis was then glued to the medical adhesive layer, and magic tape (Scotch, Germany) was taped to the adaxial epidermis and pulled away. Mesophyll cell layers were then removed carefully by affixing and pulling away with magic tape, leaving a single-layer of lower epidermis behind that was attached to the IBIDI slide. Immediately the coverslip was immersed with 1 ml plant imaging buffer (10 mM MES-Tris pH 5.6, 5 mM KCl, 50  $\mu$ M  $CaCl_2$ ) and the samples were incubated for recovery in a phyto chamber or growth cabinet overnight. The samples were incubated under light at 22–24°C for at least 1 h before imaging. Confocal laser scanning microscopy was performed in bottom imaging mode on a Zeiss LSM 780 or 880 system (Zeiss) using a 40  $\times$  water immersion objective (LD C-APOCHROME, 40 x/1.1 Korr UV-VIS M27; Zeiss) at a zoom factor of 0.8. 16-bit images were acquired every 4 s with a frame size of 512  $\times$  512 pixels and a pinhole of 200. mTurquoise and R-GECO1 were excited with 458 or 561 nm, respectively and an emission-range between 470 and 540 nm for mTurquoise or 590 and 640 nm for R-GECO1 was used for detection. Gain was set to 850 and laser intensity settings were adjusted individually to have comparable baseline intensity values for each experiment. For flg22 treatments, 50-fold and for ATP treatments 10-fold concentrations of the respective agent were prepared in water and added in a 1:50 or 1:10 volume ratio to the imaging chamber to avoid sample movement. The image processing was performed using Fiji (Schindelin et al., 2012). Images acquired in Zen Z2.3 SP1 FP3 black (Zeiss) were imported as single channel files into Fiji and all following steps were conducted for both fluorescence channels (R-GECO1 and mTurquoise): Gaussian blur filter set to 1, conversion into 32-bit, threshold adjustment (stack histogram) and selection of an appropriate look-up table. For further analyses, 26 regions of interest were selected and imported into the ROI manager. ROI\_1 included the whole image. For ROIs 2–26 the image of 512  $\times$  512 pixel was divided into 25 ROIs of 102.4  $\times$  102.4

pixels each (**Supplementary Figure 6A**). The mean gray values of these ROIs were used for further calculations. The RGmT ratio was calculated *via* dividing R-GECO1 mean gray values over mTurquoise mean gray values. The resulting RGmT emission ratio was normalized to the mean of the 10 min (flg22) or 5 min (ATP) base line ratio for each ROI before flg22 or ATP treatment ( $R/R_0$ ). For each ROI the following parameters were analyzed: maximal signal change after treatment; time until maximal signal change; number of local maxima, and time between first two local maxima. ROI\_1 and the selected ROIs with the highest number of local maxima were chosen for comparison of different genotypes. The graphs of single ROIs were generated in R, using R Studio (PBC, United States) incl. the packages tidyverse and reshape2 (**Supplementary Figure 6**; Wickham, 2007; Wickham et al., 2019; R Core Team, 2021). Scripts for the ImageJ macro and analysis in R are available on request.

### Analysis of Resting $[Ca^{2+}]_{cyt}$ Levels

We monitored R-GECO1 and mT fluorescence without stress application in leaf disks to compare resting  $[Ca^{2+}]_{cyt}$  levels between RGmT crosses. Leaf disks were sampled from 6-week-old stable *A. thaliana* expression lines and transferred to 1 mL distilled water. The sampled plants were transferred back to prior growth conditions for an overnight recovery period to minimize calcium signals induced by wounding. Leaf samples were imaged with a fluorescence stereo zoom microscope (Zeiss Axio Zoom.V16, Zeiss) using a  $1 \times$  objective and a zoom factor of 10. Optical filters for mTurquoise ( $\lambda_{excitation}$  436/20 nm,  $\lambda_{emission}$  480/40 nm) with an exposure time of 1.5 s and for R-GECO1 ( $\lambda_{excitation}$  550/25 nm,  $\lambda_{emission}$  605/70 nm) exposure time of 1.8 s were used for fluorescence detection. For image processing the following steps were conducted for R-GECO1 and mT channels using Fiji: Gaussian blur filter set to 1, conversion to 32 bit, threshold adjustment (stack histogram) and the mean gray values of the whole image area were used for calculating fluorescence intensities. Analyses revealed higher signal intensities for R-GECO1 and mTurquoise channels in the *CPK5#7* crossed line, probably indicating a higher protein abundance (**Supplementary Figure 2A**). However, the denominator channel mTurquoise exhibited an even higher increased fluorescence intensity compared to other RGmT crosses than R-GECO1 channel, and as consequence, the ratiometric readout  $fluorescence_{R-GECO1}/fluorescence_{mTurquoise}$  was decreased, indicating apparent lower resting  $[Ca^{2+}]_{cyt}$  levels. To test if altered fluorescence ratios of R-GECO1/mTurquoise may rely in a high sensor protein amount, we transiently expressed and imaged increasing RGmT protein concentrations in the same *N. benthamiana* leaf. Transient expression in *Nicotiana benthamiana* was conducted as described in Franz et al. (2011). For *N. benthamiana* infiltration reciprocal dilutions of *A. tumefaciens* cultures harboring RGmT plasmid and *A. tumefaciens* cultures harboring a plasmid coding for the N-terminal domain of slow anion channel 1 (SLAC1) (Geiger et al., 2010) were mixed leading to OD<sub>600</sub> concentration gradients of RGmT *A. tumefaciens* cultures from 0.1 to 0.5. The combinatory OD<sub>600</sub> of *A. tumefaciens* cultures harboring RGmT or SLAC1-NT were kept constant at 0.5. Leaf disks were harvest

3 days after transfection of *N. benthamiana*. Preparation and imaging setup was identical to resting  $[Ca^{2+}]_{cyt}$  measurements in *A. thaliana*.

### Systemic Calcium Imaging

Three fully developed “local” leaves of 6-week-old plants were infiltrated with either mock-treatment (10 mM  $MgCl_2$ ) or 200 nM flg22 as priming-stimulus using a needleless syringe. After 48 h, a 4 mm diameter leaf disk was harvested from a systemic leaf and fixed adaxial on a 8 well chambered coverslip (IBIDI) using medical adhesive (Ulrich Swiss) and immersed with distilled water. Selection of local and systemic leaves as described in Dubiella et al. (2013). The sample was transferred back to prior growth condition and incubated overnight to minimize calcium signals induced by wounding. The next day calcium imaging of the sample was conducted on the upright fluorescence stereomicroscope Leica M165 FC and M205 FA (Leica, Germany) using a  $2x/0.04$  objective and a zoom factor of 0.83. During in planta imaging, single RGB images (frame size of all images:  $1920 \times 1440$  pixels) of the R-GECO1 ( $\lambda_{Ex} = 546/10$  nm,  $\lambda_{Em} = 605/70$  nm, exposure time 1.5 s) and mTurquoise channels ( $\lambda_{Em} = 436/20$  nm  $\lambda_{Em} = 480/40$  nm, exposure time 1.5 s) were acquired. In the following experiment to enable, a high imaging frequency only R-GECO1 was detected every 4 s over 42 min. This step was required due to the technical limitations of the microscopes fluorescence filter system. After 2 min 10-fold concentrations of flg22 (200 nM final conc.) prepared in water was added in a 1:10 volume ratio to the imaging chamber as triggering-stimulus to avoid sample movement. Finally, another single image with both channels was recorded. Images acquired during the calcium assay in LAS X (Leica application software) were imported into Fiji and all following steps were conducted for the R-GECO1 channel: Conversion to 8-bit; brightness and contrast adjustment, conversion into 32-bit, threshold adjustment (stack histogram) and selection of 16 colors look-up table. R-GECO1 fluorescence during the time course was normalized to R-GECO1 fluorescence at 2 min (flg22-application; fluorescence time point  $t_0 = F_0$ ) *via* dividing each of the following created 600 images over the R-GECO1 signal at time point  $t_0$  ( $F/F_0$ ) using the image calculator tool in Fiji. The resulting mean gray value was used for further calculations. The RGmT ratio in the beginning and at the end of the assay was calculated *via* dividing R-GECO1 mean gray values by mTurquoise mean gray values using the acquired raw data.

### Aequorin-Based Calcium Assays

Individual 8–10-day-old seedlings were transferred into 96-well plates, reconstituted with 10  $\mu$ M coelenterazine and cytosolic calcium measurements were performed as described (Trempe et al., 2016). The calculation for intracellular calcium concentrations is based on the previously described equation:  $pCa = 0.332588 (-\log k) + 5.5593$  (where  $k = L/L_{max}$  i.e., luminescence counts per s/total remaining luminescence counts) (Rentel and Knight, 2004). To facilitate visualization of small differences, the values for relative calcium concentrations ( $L/L_{max}$ ) are shown. An aequorin-expressing line in the *rbohD*

background (Ranf et al., 2011) served as a control for the lack of a second flg22-induced calcium peak.

## Western Blotting and Immunodetection

Leaf material was homogenized in liquid nitrogen in a laboratory mill and incubated with a 2:1 weight ratio of 2x SDS sample buffer (60 mM Tris, pH 6.8; 100 mM DTT; 10% (v/v) glycerol; 2% (w/v) SDS; 0.004% (w/v) bromophenol blue) for 5 min at 95°C for subsequent SDS-Page western blotting and immunodetection. For RGmT detection the  $\alpha$ -RFP (Rockland, United States) primary antibody was used. For visualization a HRP-conjugated secondary  $\alpha$ -rabbit-HRP antibody (Sigma-Aldrich, United States) was applied. Blots were developed using enhanced chemiluminescence (Thermo Scientific, United States) detection.

## Gene Expression Analyses (RT-qPCR)

RNA, isolated with Trizol reagent, was treated with DNaseI (to remove any remaining DNA contamination) and cDNA was synthesized using the RevertAid kit (Thermo Scientific). For realtime quantitative PCR (qPCR), amplicons from cDNA samples were analyzed on an Mx3005P qPCR system (Agilent, United States) after amplification with Maxima SYBR Green qPCR Master Mixes (Thermo Scientific). Relative gene expression values of the genes-of-interest were calculated with the comparative CT method (Schmittgen and Livak, 2008) using the reference gene PP2A [AT1G13320; (Czechowski et al., 2005)]. Primers are listed in **Supplementary Table 1**.

## RESULTS

### Generation and Characterization of R-GECO-mT Calcium Reporter Lines in *cpk5* and CPK5-OE Lines

To be able to investigate stimulus-induced  $[Ca^{2+}]_{cyt}$  changes in response flg22 perception, we generated  $Ca^{2+}$  reporter lines expressing R-GECO1-mTurquoise (RGmT) in *cpk5* and CPK5-OE (CPK5#7) backgrounds by crossing. Since all GECl lines have been reported to exhibit some level of reduced growth (Waadt et al., 2017), we first characterized the resulting RGmT crosses for any obvious growth phenotype. In 6-week-old plants grown under short day conditions, no obvious defects were observed, except for the slightly smaller plants in the CPK5#7 overexpressing background and in *rbohD*. Both genotypes have already been previously reported to be smaller compared to wild-type (Torres et al., 2002; Dubiella et al., 2013). Importantly, the smaller stature of CPK5#7 that was previously correlated with a constitutively higher basal resistance compared to wild-type (Dubiella et al., 2013), was not further aggravated through introgression of the RGmT reporter (**Figure 1A**). Elevated basal expression of the defense marker genes, *PR1* and *NHL10*, characteristic for enhanced CPK5 signaling and a constitutively higher immune status in CPK5-OE, was also confirmed for the CPK5#7xRGmT cross (**Supplementary Figures 1A,B**). In addition, this leads to an enhanced flg22 responsiveness in the

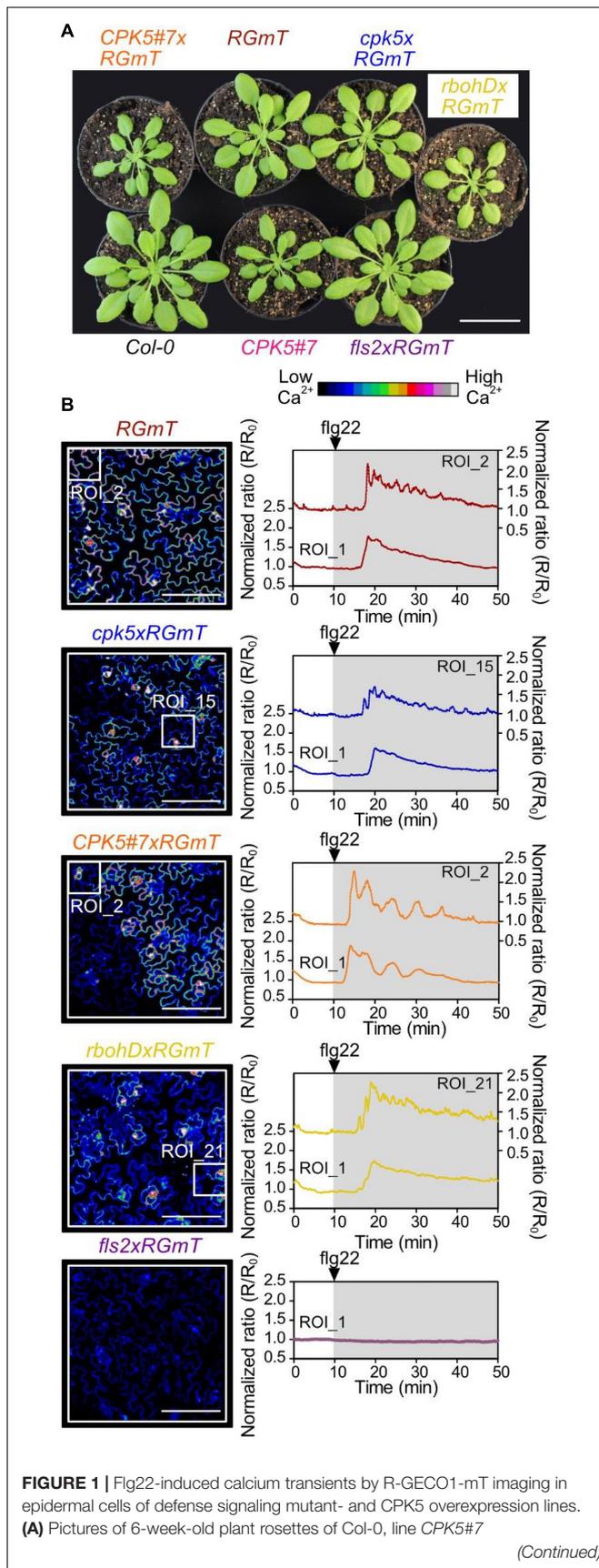
CPK5#7 background as is evident from the expression pattern of the flg22-responsive gene, *FRK1* (**Supplementary Figure 1C**).

To verify comparable expression of the RGmT reporter, both RT-qPCR and immunoblotting were performed where RGmT was shown to be well expressed in all lines. However, the CPK5#7xRGmT line expressed about threefold as much RGmT compared to the parental line or RGmT crosses with *cpk5*, *rbohD*, or *fls2* (**Supplementary Figures 2A,B**). We next compared resting  $[Ca^{2+}]_{cyt}$  based on R-GECO imaging normalized to mT. Interestingly, an apparently lower  $F_{R-GECO1}/F_{mT}$  ratio indicative of a lower  $[Ca^{2+}]_{cyt}$  was observed in CPK5#7xRGmT (**Supplementary Figure 2C**). We thus addressed whether differential levels of the RGmT reporter can distort the  $[Ca^{2+}]_{cyt}$  calculation. Using the *N. benthamiana* transient expression system, increasing protein levels were independently assessed for R-GECO and mT emissions. The  $F_{R-GECO1}/F_{mT}$  ratio showed that higher reporter levels indeed generated a lower estimation of the apparent  $[Ca^{2+}]_{cyt}$  (**Supplementary Figures 2D,E**) (see the section “Materials and Methods” for further details). This must be taken into consideration for data interpretation of resting  $[Ca^{2+}]_{cyt}$  when comparing lines of different genetic backgrounds. Despite this caveat, the RGmT reporter is still one of the most suitable systems for imaging stimulus-induced changes in  $Ca^{2+}$  transients within systemic tissues of adult plants with high sensitivity.

### Enhanced CPK5 Signaling Triggers Distinct Flg22-Induced Calcium Signatures

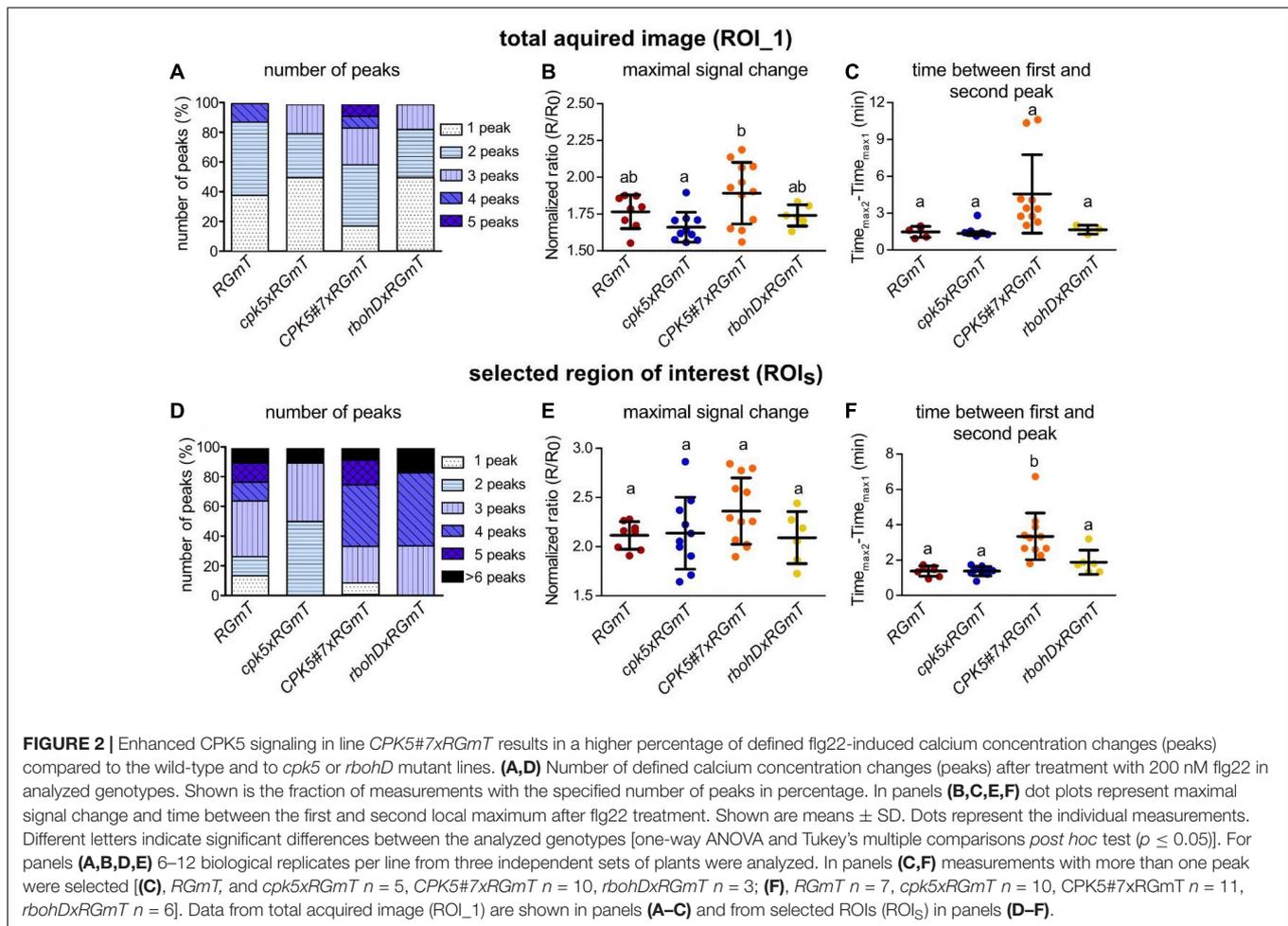
Since both, basal and flg22-stimulated defense transcription responses were elevated in the CPK5#7 plants (**Supplementary Figure 1C**), we asked if  $Ca^{2+}$  signaling responsiveness is also primed. We first focused on the  $Ca^{2+}$  response in local tissues exposed to flg22 peptide as a trigger and compared the different lines for stimulus-induced changes in  $Ca^{2+}$  transients in epidermal cells of 6-week-old plants. Mathematical models have previously predicted that  $Ca^{2+}$  responses reflect the number of measured cells (Dodd et al., 2006), which was proven using R-GECO1-based  $[Ca^{2+}]_{cyt}$  imaging (Keinath et al., 2015) where the “oscillatory behavior” of the flg22 response was better resolved by focusing on a specific region (i.e., encompassing fewer cells) than an area covering more cells. Hence, we chose similar experimental conditions, where we imaged a region-of-interest (ROI<sub>1</sub>) covering  $47 \pm 17$  epidermal cells for a global view and a second smaller ROI (designated as ROI<sub>S</sub>) of  $6 \pm 2$  cells for improved resolution of the  $Ca^{2+}$  dynamics (**Figure 1B**, left panels). ROI<sub>S</sub> was selected to represent a sub-region with more defined flg22-induced  $[Ca^{2+}]_{cyt}$  transients (for details of ROI selection see Materials and Methods and **Supplementary Figure 6**).

Overall, flg22-elicitation induces RGmT responses that are predominantly of a monophasic shape for ROI<sub>1</sub> (i.e., the average of multiple cells) over 40 min, and a more pronounced spiky response for ROI<sub>S</sub> (**Figure 1B**, right panels, **Supplementary Movie 1**). This  $Ca^{2+}$  response is specific since no flg22-induced rise in  $[Ca^{2+}]_{cyt}$  was detected in the *fls2* receptor



**FIGURE 1** | overexpressing *CPK5-YFP*, and plants carrying the ratiometric  $\text{Ca}^{2+}$ -sensor R-GECO1-mTurquoise (RGmT) generated by crossing with *CPK5#7*, *cpk5*, *rbohD*, and *fls2* as indicated. **(B)** Changes in the cytosolic calcium concentration  $[\text{Ca}^{2+}]_{\text{cyt}}$  were visualized in response to 200 nM flg22 in epidermal peels of 6 week old plants (left panels) recorded using RGmT for a time period of 40 min after flg22 treatment (right panels). Normalized fluorescence ratios ( $R/R_0$ ) over time were calculated from the total acquired image (Region of interest 1 (ROI\_1) – left axis) and from a selected ROI (ROIs; marked by small squares – right axis) that exhibited the highest number of peaks. Graphs were normalized to mean RGmT ratio of the 10 min before flg22 treatment. The 40 min time interval of recording after the 200 nM flg22 treatment is indicated by the underlying gray area (right panels). Microscopic images represent R-GECO1 fluorescence intensities at the time point of the maximal calcium signal change. Shown are representative experiments with  $n \geq 6$  per analyzed line from three independent sets of plants. Scale bars in panel **(B)** represent 100  $\mu\text{m}$  and 4 cm in panel **(A)**.

mutant (**Figure 1B**, bottom traces). Important to note is that there is substantial variation in the  $\text{Ca}^{2+}$  response traces between experiments (**Supplementary Figure 3**), possibly due to asynchronous responses across the tissues, or variations in the perception of the flg22 peptide in the imaged regions. For instance, the lag time between flg22 application to the first  $\text{Ca}^{2+}$  peak is  $9.2 \pm 5.8$  min ( $n = 36$ ). Here, the large variation in onset of  $\text{Ca}^{2+}$  signaling may be attributable to technical restrictions related to poor flg22 diffusion/accessibility in the tight space between the leaf surface and the coverslip in the bottom imaging mode. Interestingly, however, traces of the overexpressing *CPK5#7xRGmT* line exhibited a distinct  $\text{Ca}^{2+}$  signature characterized by defined peaks and longer intervals between the peaks (**Figures 1B, 2** and **Supplementary Movie 2**). To substantiate this perceived difference in  $[\text{Ca}^{2+}]_{\text{cyt}}$  changes in *CPK5#7xRGmT*, we consolidated the data of all our RGmT measurements into three parameters defining the  $\text{Ca}^{2+}$  signature: (1) the number of peaks within the 40 min recording time, (2) the maximum signal change (amplitude), and (3) the time between the first and second peak. A distribution plot showed a shift toward  $\text{Ca}^{2+}$  traces with an increased “number of peaks” in the *CPK5#7xRGmT* line, which is consistent for both ROI\_1 and ROIs (**Figures 2A,D**). Additionally, the “time between first and second peak” for ROIs (**Figure 2F**) was significantly enhanced in the *CPK5#7xRGmT* line. Moreover, a statistically significant difference between *CPK5#7xRGmT* compared with *cpk5xRGmT* in the “maximal signal change” for ROI\_1 was observed (**Figure 2B**) with a higher amplitude in the overexpressing and lower amplitude in the corresponding mutant line, respectively. Notably, this apparent boost of the signal amplitude in *CPK5#7xRGmT* or reduction in *cpk5xRGmT* was not seen when ATP was used as a trigger (**Supplementary Figure 4A**), so that this change in  $\text{Ca}^{2+}$  signature is elicitor specific. In addition, we could rule out that a somewhat smaller plant size as seen in the *CPK5#7* and *rbohD* crosses may lead to different number of cells being imaged within the ROI compared to the other lines, which has been reported to possibly affect the features of a  $\text{Ca}^{2+}$  response and confound data interpretation (Dodd et al., 2006). However, no correlation between the flg22 response and the number of imaged cells per ROI\_1 for the different genotypes was observed (**Supplementary Figure 4B**).



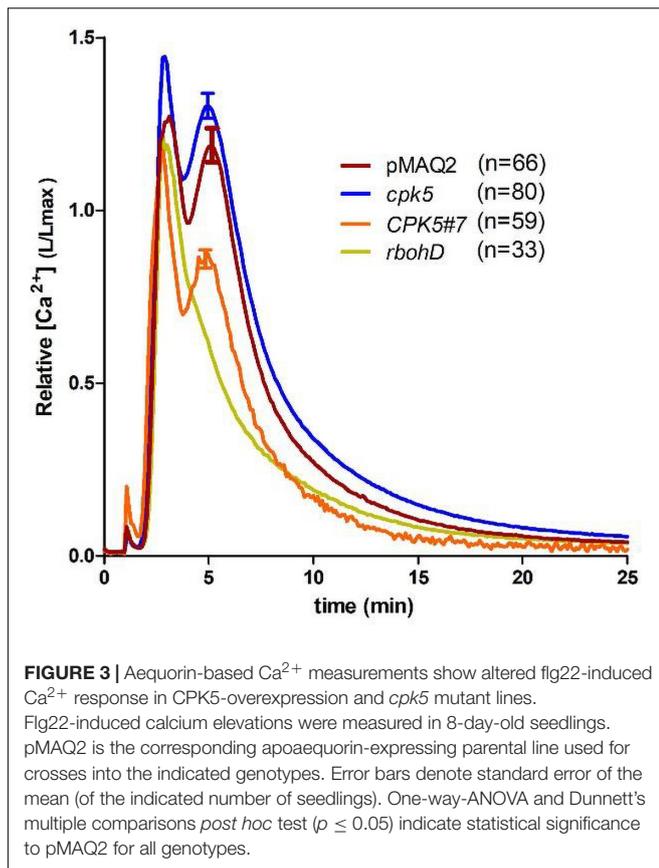
These data indicate that enhanced immune signaling through overexpression of CPK5 triggers a distinct  $\text{Ca}^{2+}$  signature in epidermal cells of 6-week-old plants. This can largely be described as increasing number of well-defined peaks with higher amplitude and an increased time-gap between first and second peak.

We next compared the flg22-induced  $\text{Ca}^{2+}$  response using the well-established aequorin-based assay, which has been shown to typically produce a highly reproducible and quantitative response in young seedlings. Therefore, the aequorin-expressing line, pMAQ2, was crossed into *cpk5* and *CPK5#7*. We observed the typical flg22-induced  $\text{Ca}^{2+}$  change reported for the aequorin system, which comprises of a rapid  $\text{Ca}^{2+}$  increase with twin peaks at 2–3 min and 5–6 min, respectively, and a gradual return to resting  $[\text{Ca}^{2+}]_{\text{cyt}}$  within 20–25 min (**Figure 3**). The overall calcium signature is thus very similar to the monophasic response (ROI<sub>1</sub>) recorded with RGmT (**Figure 1B**). When the previously described aequorin-expressing *rbohD* line (Ranf et al., 2011) was used as a control, the second flg22-induced  $[\text{Ca}^{2+}]_{\text{cyt}}$  peak was not detected. This indicates that the second peak is ROS-dependent. The profile of the calcium response (kinetics and twin peak profile) is generally not affected by *cpk5* mutation or *CPK5* overexpression. However, a significant reduction in the

$\text{Ca}^{2+}$  amplitude in the *CPK5#7* line and an increase in the *cpk5* mutant was observed (**Figure 3**). Taken together, both aequorin and RGmT imaging techniques report an altered  $\text{Ca}^{2+}$  signature upon a local flg22 stimulus in the *CPK5* overexpressing line. These data imply that enhanced CPK5 signaling, correlating with an increase in basal immunity status, display a different  $\text{Ca}^{2+}$  signature in response to a local PAMP trigger (such as flg22).

## Establishment of an Experimental Set-Up for Systemic Calcium Response Analyses

We next investigated whether systemic tissues of plants pre-treated with a first local flg22 trigger, and thus having been “primed” toward an immune memory, display an altered  $\text{Ca}^{2+}$  response when exposed to a secondary flg22 trigger. As illustrated in the scheme in **Figure 4A**, three leaves of 6-week-old plants were infiltrated with 200 nM flg22 (or 10 mM  $\text{MgCl}_2$  as a mock control). After 2 days incubation in a growth chamber, a disk from a systemic leaf was sampled and mounted on a microscope chamber slide, incubated again over-night in a growth cabinet for recovery, and incubated for 1 h in light before the second flg22 treatment and RGmT-based calcium imaging. This modification



of the experimental setup was necessary because the preparation of epidermal peels (as used in investigation of local responses, **Figure 1**) can interfere with the priming stimulus in the systemic tissues. The  $\text{Ca}^{2+}$  response was monitored for 2 min before and 40 min after the flg22 stimulus. We could verify that the mT fluorescence monitored before and after measuring remained uniform for all lines indicating constant RGM1 protein level throughout the analysis period. This allowed us to apply an intensimetric quantification for R-GECO1, which simplified data recording at high imaging frequencies. **Figure 4B** shows the first 13 min after flg22 treatment of an exemplary imaging series, depicting the flg22-induced  $[\text{Ca}^{2+}]_{\text{cyt}}$  increase beginning from the edge of the leaf disk and moving toward the middle. This inward movement  $[\text{Ca}^{2+}]_{\text{cyt}}$  signals may reflect a gradual access of flg22 from the wounded leaf disk edges or perhaps cell-to-cell  $\text{Ca}^{2+}$  propagation. Additionally, we also observed occasional local spots of  $\text{Ca}^{2+}$  increase in some leaf disks but these were not associated to a certain genotype or pre-treatment (see **Supplementary Movie 3**). Nevertheless, the described system is suitable for monitoring  $\text{Ca}^{2+}$  responses in systemic tissues of primed plants.

### Flg22-Induced Calcium Response Patterns Are Independent of Priming

For comparing flg22-induced  $[\text{Ca}^{2+}]_{\text{cyt}}$  changes in systemic leaf disks from primed vs. mock-treated plants, the pooled data

of all measurements are summarized in **Figure 5**, with the individual  $\text{Ca}^{2+}$  traces of the 40 min recording period displayed in **Supplementary Figure 5**. The whole area of the leaf disk was quantified for a global overview of all cells within these disks. We have decided for this global overview analyses, in contrast to the analyses of selected smaller regions (as in **Figure 1**) for two reasons: (1) we cannot predict which cell has been “primed” for systemic immunity, and (2) it is known from our RGM1 measurements that the flg22-induced response may not be synchronous (**Figure 1** and **Supplementary Figure 3**).

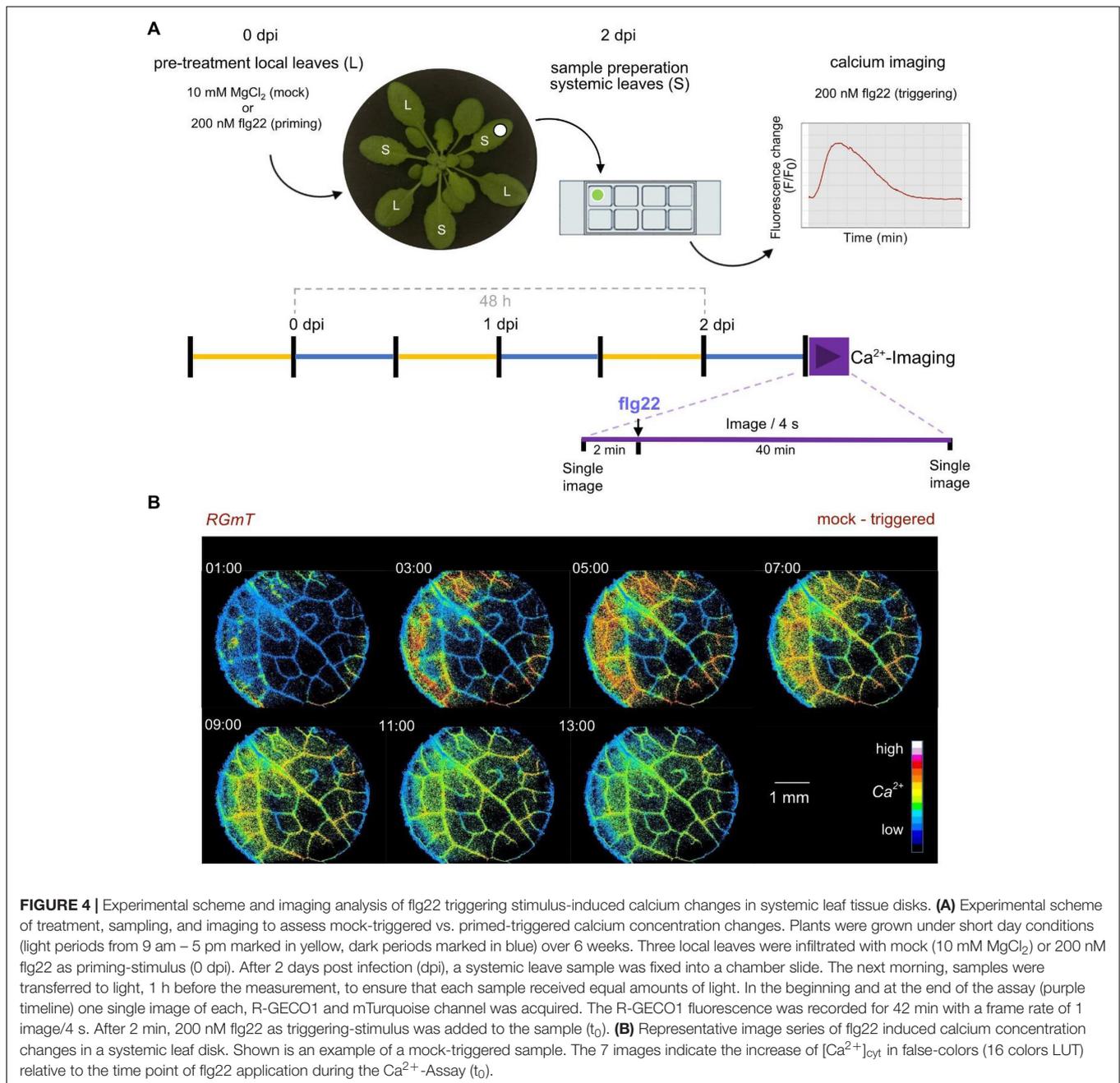
In general, no significant differences were discernible that could be associated with the priming status of the plants, neither in wild type (**Figure 5A**), nor in the *cpk5* mutant (**Figure 5B**) or CPK5-OE lines (**Figure 5C**). More specifically, the quantitative data did not reveal any differences in the  $\text{Ca}^{2+}$  response in terms of the overall  $\text{Ca}^{2+}$  signature (**Figures 5A–C**), maximal fluorescence signal change (**Figure 5D**) or peak response time (i.e., time after flg22 application until maximal signal change) (**Figure 5E**). When assessing the basal resting  $[\text{Ca}^{2+}]_{\text{cyt}}$ , we observed an apparent lower value in CPK5#7 similar to what we have seen before in a local flg22 response in the absence of a pre-treatment. We therefore re-evaluated the resting  $[\text{Ca}^{2+}]_{\text{cyt}}$  based on R-GECO1 imaging normalized to mT (fluorescence ratio) within our priming experiments and found that the resting  $[\text{Ca}^{2+}]_{\text{cyt}}$  was with statistical significance lower in the CPK5-overexpressing line *CPK5#7xRGM1* compared to the wild-type or *cpk5* carrying RGM1 (**Figure 5F**), irrespectively of a priming or a mock pre-treatment.

Taking our data from wild-type subjected to a priming and triggering treatment together with those obtained with the CPK5-OE line, which is already constitutively “primed” and capable of stimulus-dependent “super-priming,” these data indicate that the immune status of a plant has no (detectable) influence on a stimulus-induced  $\text{Ca}^{2+}$  response downstream of flg22 perception in systemic tissues under the imposed conditions.

## DISCUSSION

“Priming” in plant immunity describes the state of a plant, in which a preceding primary infection by microbial pathogens induced an immune memory, so that the plant is prepared when challenged by subsequent infection. In SAR, the ability to display faster and stronger defense reactions is correlated with molecular changes of the plant characterized by distinct patterns of systemic defense signaling molecules (NHP), phytohormone levels (SA, JA) and expression of key transcription factors (*SARD1*) (Truman et al., 2007; Chen et al., 2018; Hartmann et al., 2018; Sun et al., 2018, 2020; Kim et al., 2020; Lim et al., 2020; Schnake et al., 2020; Vlot et al., 2021). Furthermore, these characteristic responses are often interconnected with each other in auto-activation and -synthesis loops. Our objective in this work is to investigate whether the intracellular  $\text{Ca}^{2+}$  change in response to a direct secondary, triggering stimulus is likewise altered as a consequence of its integration in such a systemic activation loop.

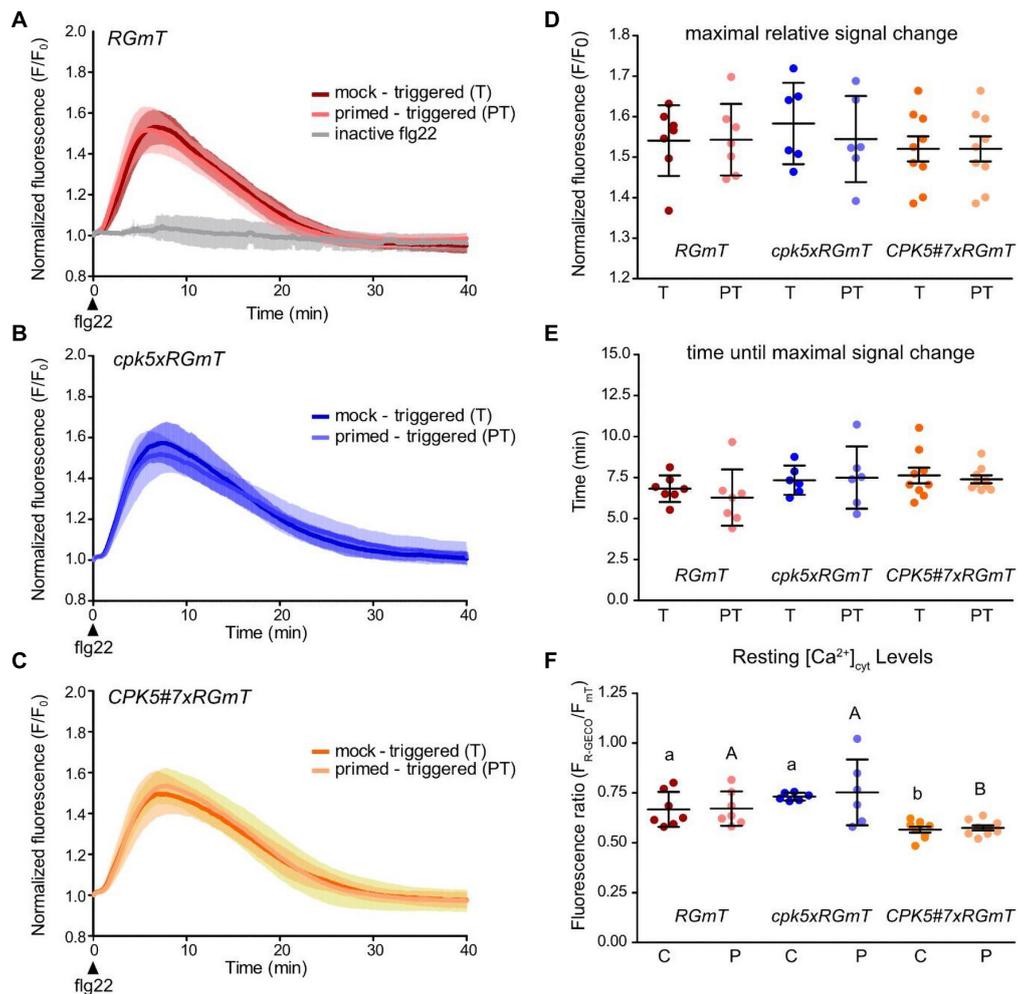
In the context of priming, we quantified the  $\text{Ca}^{2+}$  response in leaf disks of adult plants upon a triggering flg22 stimulus



**FIGURE 4** | Experimental scheme and imaging analysis of flg22 triggering stimulus-induced calcium changes in systemic leaf tissue disks. **(A)** Experimental scheme of treatment, sampling, and imaging to assess mock-triggered vs. primed-triggered calcium concentration changes. Plants were grown under short day conditions (light periods from 9 am – 5 pm marked in yellow, dark periods marked in blue) over 6 weeks. Three local leaves were infiltrated with mock (10 mM MgCl<sub>2</sub>) or 200 nM flg22 as priming-stimulus (0 dpi). After 2 days post infection (dpi), a systemic leaf sample was fixed into a chamber slide. The next morning, samples were transferred to light, 1 h before the measurement, to ensure that each sample received equal amounts of light. In the beginning and at the end of the assay (purple timeline) one single image of each, R-GECO1 and mTurquoise channel was acquired. The R-GECO1 fluorescence was recorded for 42 min with a frame rate of 1 image/4 s. After 2 min, 200 nM flg22 as triggering-stimulus was added to the sample ( $t_0$ ). **(B)** Representative image series of flg22 induced calcium concentration changes in a systemic leaf disk. Shown is an example of a mock-triggered sample. The 7 images indicate the increase of  $[Ca^{2+}]_{cyt}$  in false-colors (16 colors LUT) relative to the time point of flg22 application during the Ca<sup>2+</sup>-Assay ( $t_0$ ).

from plants that had or had not experienced a previous flg22 priming treatment. The leaf disk approach bears the opportunity to average signals over a multitude ( $>n 10^3$ ) of cells. While the aequorin-based system (Ranf et al., 2011) can similarly provide global Ca<sup>2+</sup> response of multiple cells simultaneously, it is not suitable for our study because the seedlings commonly used are too young to mount a systemic immune memory. Furthermore, concurrent observation of multiple cells is of particular importance in SAR because it is unpredictable which and how many cells have undergone systemic priming and built an immune memory upon the pre-treatment 2 days earlier.

Only these cells would give rise to priming-dependent  $[Ca^{2+}]$  changes. To exclude that our chosen imaging approach may average out the individual cell responses if only a minority of cells react differently after priming, we included the CPK5-OE line in our studies. This line is characterized through molecular markers to be constitutively primed (**Supplementary Figure 1**) and capable of “super-priming” responses (Guerra et al., 2020). Additionally, averaging of single cell Ca<sup>2+</sup> signals led successfully to the characterization of priming in response to cold stress (Knight et al., 1996; Knight and Knight, 2000).



**FIGURE 5** | Defense priming has no significant effect on stimulus-induced calcium concentration changes in systemic tissue. **(A–C)** Flg22-induced calcium changes were recorded in systemic leaf disks 2 days after pre-treatment *via* infiltration of local leaves with 10 mM  $MgCl_2$  (mock) or 200 nM flg22 (primed). As triggering stimulus in the systemic leaf, 200 nM flg22 was applied, leading to the samples mock – triggered (T) or primed – triggered (PT), respectively. For detailed experimental setup, see **Figure 4**. As a control, inactive flg22 from *A. tumefaciens* was used as triggering stimulus with local mock pre-treatment. Data are averages of R-GECO1 fluorescence over time normalized to R-GECO1 fluorescence intensity at time point of flg22 application ( $t_0$ ). Error bars denote mean  $\pm$  SD for  $n$  replicates (*RGmT*  $n_T = 7$ ,  $n_{PT} = 7$ ,  $n_{inactiveflg22} = 3$ , **(B)**, *cpk5xRGmT*  $n_T = 6$ ,  $n_{PT} = 6$  and **(C)**, *CPK5#7xRGmT*  $n_T = 9$ ,  $n_{PT} = 9$ ). Parameters of flg22 induced calcium changes of measurements shown in panels **(A–C)** are analyzed in panels **(D–F)**. In panel **(D)** dot plots represent the amplitude of the maximal signal change, while in panel **(E)** the correlating time points of maximum signal after flg22 treatment are shown. Dots represent the individual measurements. Shown are means  $\pm$  SD of 6 – 9 biological replicates per line for each treatment (T vs. PT). **(F)** Apparent resting  $F_{R-GECO1}/F_{mT}$  in systemic leaf disks of local mock (10 mM  $MgCl_2$ , control = C) or 200 nM flg22 (primed = P) treated plants. Shown are means  $\pm$  SD ( $n \geq 6$ ). Dots represent the individual measurements. Two-way ANOVA and Bonferroni post test ( $p \leq 0.05$ ) reveals no significant differences between the pre-treatments but significant differences between genotypes indicated by different letters.

By contrast, when comparing a local induced  $Ca^{2+}$  response in different genetic knock-out lines,  $Ca^{2+}$  changes can be traced in a few or even a single cell resolution, assuming that all cells are equally affected in a defined genetic background. Indeed, when selecting ROIs that cover only few cells, we observed distinct traces that displayed an oscillating  $Ca^{2+}$  pattern for approximately 30 min reminiscent to what has been reported by Keinath et al. (2015). While these distinct peaks became evident in selected ROIs covering single to few cells, an oscillatory  $Ca^{2+}$  pattern was less pronounced in the whole ROI image. These data validate the suitability of the R-GECO1 calcium sensor

for investigating calcium changes in 6-week-old plants, required for priming experiments, and for the recording in epidermal peels combined with a bottom imaging setup. Under conditions of enhanced CPK5 signaling, more distinct  $Ca^{2+}$  peaks with slightly higher amplitudes could be recorded compared to the wild type and even more so compared to *cpk5* and *rbohD* mutants (**Figures 1, 2** and **Supplementary Figure 3**). We have validated for the whole imaging ROI that the  $Ca^{2+}$  pattern was not affected by the number of imaged ( $47 \pm 17$ ) cells (**Supplementary Figure 4**). Therefore, the more distinct  $Ca^{2+}$  transients observed in the CPK5-OE lines may have a biological cause directly related

to CPK5 activity itself. CPK5 is known to activate NADPH-oxidase RBOHD, and a  $\text{Ca}^{2+}$ - and ROS-mediated defense signal propagation has been discussed (Dubiella et al., 2013). Our data suggest that upon flg22 stimulation, enhanced CPK5 activity leads to a reinforcement and local synchronization of  $\text{Ca}^{2+}$  signaling in these lines. Such interpretation is corroborated by the video depicting distinct (local) waves of  $\text{Ca}^{2+}$  signals induced by flg22 in the CPK5-OE line compared to *RGmT* (**Supplementary Movies 1, 2**). It is tempting to speculate that this reinforced  $\text{Ca}^{2+}$  signature mediated by CPK5 is not only part of the signal propagation to neighboring cells but is likewise responsible in each single cell to translate the  $\text{Ca}^{2+}$  signature into downstream defense reactions. In this context, it is worth to note that the whole image ROI shows fewer distinct peaks in the *rbohD* and *cpk5* mutants (**Figure 2A**). Likewise, the selected ROIs single cell traces also show fewer peaks in *cpk5* (**Figure 2D**). In support of the proposed amplificatory role of CPK5 in signal propagation (Dubiella et al., 2013; Guerra et al., 2020), statistically significant increase of the peak intervals is seen in the CPK5-OE (**Figure 2F**). Taken together, these data indicate that CPK5 may synchronize  $\text{Ca}^{2+}$  signaling within and between cells, and the absence of CPK5 leads to a reduction in both. RBOHD, which is biochemically phosphorylated and activated by CPK5, is required for the  $\text{Ca}^{2+}$  signal propagation to neighboring cells. This interpretation is consistent with our previous data showing that RBOHD and CPK5 constitute an auto-activating mechanism for defense signal spread to distal tissues (Dubiella et al., 2013). Here, we provide evidence that RBOHD contributes to the CPK5-reinforced synchronization for intercellular  $\text{Ca}^{2+}$  signal propagation.

How can the “oscillatory”  $\text{Ca}^{2+}$  signature in the CPK5-OE line be explained mechanistically? One possible explanation is a more coordinated in- and efflux of cytosolic  $\text{Ca}^{2+}$ , i.e., activation of the  $\text{Ca}^{2+}$  efflux and inhibition of  $\text{Ca}^{2+}$  influx. Thus, CPK5 may contribute to the  $[\text{Ca}^{2+}]_{\text{cyt}}$  homeostasis by promoting  $\text{Ca}^{2+}$  efflux out of the cytosol. Corresponding  $\text{Ca}^{2+}$  efflux transporters may be found among  $\text{Ca}^{2+}/\text{H}^{+}$  exchangers (CAX) driven by electrochemical gradients of  $\text{H}^{+}$  and autoinhibited P-type II  $\text{Ca}^{2+}$ -ATPases (ACA) (Sanders et al., 2002; Kudla et al., 2010; Bose et al., 2011; Demidchik et al., 2018). ACAs have been described in immune signaling (Boursiac et al., 2010; Frei dit Frey et al., 2012). Interestingly, the analysis of *aca4 aca11* mutants lacking ACA type of  $\text{Ca}^{2+}$  pumps revealed an increase in basal and flg22-induced rise in  $[\text{Ca}^{2+}]_{\text{cyt}}$  (Hilleary et al., 2020). P-type II ACA-  $\text{Ca}^{2+}$ -pumps are regulated by protein phosphorylation and some members are characterized by an N-terminal autoinhibition domain (Giacometti et al., 2012; Costa et al., 2017). Similarly, PAMP-responsive  $\text{Ca}^{2+}$ -permeable channels are known to be regulated by phosphorylation (Tian et al., 2019; Thor et al., 2020). Whether such  $\text{Ca}^{2+}$  pumps and  $\text{Ca}^{2+}$  channels can be directly phosphorylated by CPK5 and contribute to the encoding of oscillations remains to be shown. It is noteworthy, in this context, that CPK5 displays a rather low  $K_d$  for  $\text{Ca}^{2+}$  of  $\sim 100$  nM for kinase activity. For other CDPKs auto-phosphorylation at its N-terminal domain can shift substrate accessibility (Ito et al., 2010, 2017, 2018). Such a

mechanism could provide an additional layer in CPK5 regulation. Both mechanisms may render CPK5 as a suitable  $\text{Ca}^{2+}$  sensor-kinase effector-protein to reinforce “oscillatory”  $\text{Ca}^{2+}$  changes, which possibly include a  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release mechanism (Choi et al., 2014).

A natural follow-up question is: “How are the “more pronounced and defined number of  $\text{Ca}^{2+}$  peaks” observed in the CPK5-OE line decoded into enhanced basal defense responses within a cell?” Based on what is known from plant immune signaling in pattern-triggered, effector-triggered or systemic immunity in SAR, this will involve the myriad of different  $\text{Ca}^{2+}$ -sensor, -relay and -effector proteins, such as CaM/CML, kinases or CAMTA transcription factors (Kudla et al., 2010). In the absence of additional evidence, one may further speculate that the decoding of  $\text{Ca}^{2+}$  transients into downstream defense responses may depend on the “counting” of encoded cytosolic (and/or nuclear)  $\text{Ca}^{2+}$  changes (up and downs in  $[\text{Ca}^{2+}]$ ) rather than a monophasic  $\text{Ca}^{2+}$  change.

In summary, our data provide evidence for an altered CPK5-dependent  $\text{Ca}^{2+}$  signature upon flg22 treatment in local tissues. This is consistent with CPK5 function as a predominant  $\text{Ca}^{2+}$  sensor and effector in pattern-triggered immunity mediating  $\text{Ca}^{2+}$  signal synchronization and defense response activation. However, the flg22-induced  $[\text{Ca}^{2+}]_{\text{cyt}}$  increase in systemic cells are not different in the various priming context. Therefore, at least according to our experimental system, rapid  $\text{Ca}^{2+}$  changes in systemic tissues do not reflect the plant memory of “having been primed.”

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## AUTHOR CONTRIBUTIONS

BE, SL, JL, AL, and TR conceived and designed the strategy and experiments. BE, SL, and AL generated the *RGmT* lines. BE conducted all priming experiments and analyzed the systemic calcium imaging. SL and AL analyzed the local calcium imaging. FT conducted the aequorin-based calcium assays. XJ conducted the molecular characterization of *RGmT* lines. TG generated the aequorin lines. RW designed and generated the *RGmT* construct. BE, SL, XJ, AL, and JL prepared the figures. JL, AL, and TR wrote the manuscript. All authors discussed the results and commented on the manuscript.

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## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2021.798230/full#supplementary-material>

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