The transcriptional response of *Arcobacter butzleri* to cold shock

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**Keywords**

*A. butzleri; cold adaptation; cold shock-related genes; temporal transcriptional expression*

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*Arcobacter* is an emerging zoonotic pathogen associated with gastrointestinal diseases, such as abdominal cramps and diarrhea, and is widely detected in animals, showing a high prevalence in poultry and seafood. The survival and adaptation of *A. butzleri* to cold temperatures remains poorly studied, although it might be of interest for food safety considerations. To address this, growth patterns of eight *A. butzleri* isolates were determined at 8 °C for 28 days. *A. butzleri* isolates showed strain-dependent behavior: six isolates were unculturable after day 18, one exhibited declining but detectable cell counts until day 28 and one grew to the stationary phase level. Out of 13 *A. butzleri* cold shock-related genes homologous to *Escherichia coli*, 10 were up-regulated in response to a temperature downshift to 8 °C, as demonstrated by reverse transcription-quantitative PCR. Additionally, we compared these data with the cold-shock response in *E. coli*. Overall, we provide a deeper insight into the environmental adaptation capacities of *A. butzleri*, which we find shares similarities with the *E. coli* cold-shock response.

*Arcobacter* spp. is an emerging zoonotic pathogen associated with gastrointestinal diseases such as abdominal cramps and diarrhea [1–3]. Among the 29 species within the genus of *Arcobacter* described so far [4], *A. butzleri* is the most widely detected species in animals, environment and foods, with high prevalences in poultry meat and seafood samples [5–10]. Although the good environmental adaptability of *A. butzleri* is well known, the survival and adaptation of *A. butzleri* to cold remains poorly studied so far, although it might be of interest for food safety considerations. *A. butzleri* has an optimal growth temperature of 30 °C under both aerobic and microaerobic conditions but is likely to be exposed to a wide temperature range during its transmission cycle. The minimum growth temperature of *A. butzleri* was initially reported to be 15 °C [11]. Later, some studies reported lower limits as 10 °C; for example, in Ellinghausen McCullough Johnson Harris medium [12] and in chicken meat juice, as well as in Brain Heart Infusion (BHI) broth [13]. Van Driessche and Houf [14] investigated the survival capability of *A. butzleri* in water with/without organic material at minimum 4 °C and reported that all tested *A. butzleri* strains survived for at least 203 days when organic material is present in the water, indicating the best cold adaptation among all human-related *Arcobacter* species.

With the complete genome of the *A. butzleri* RM4018 sequenced in 2007 [15], the understanding of adaptation to environmental stress could be improved. To date, no data describing the cold-shock response of *Arcobacter* spp. have been reported. Because previous phenotypic studies suggest that *A. butzleri* could adapt to cold temperatures, the presence of cold-related genes is presumed.

The function of the cold-related proteins is best studied in *Escherichia coli*. Jones et al. [16] first reported that, after a cold shock from 37 °C to 10 °C,

**Abbreviations**

BHI, brain heart infusion; CFU, colony-forming unit; RT-qPCR, reverse transcription-quantitative PCR.
the transcription and translation processes for a majority of genes slow down or come to an almost complete stop, whereas a set of cold-shock proteins is preferentially and transiently expressed [17–20]. These proteins were classified in two groups according to their expression pattern [21]. The class I proteins include: CspA (the major cold-shock protein of E. coli, encoded by cspA), DeaD (reassigned as CsdA, a DEAD-box protein, encoded by deaD), NusA (the transcription termination/antitermination protein, encoded by nusA), RbfA (the ribosome binding factor A, encoded by rbfA) and RNase R (encoded by rnr). The induction of the expression level of these proteins varies but is more pronounced compared to class II proteins that include PNPase (the polynucleotide phosphophorylase, encoded by pnp), IF-2 (the translation initiation factors IF2, encoded by infB), Gyra (the DNA gyrase subunit A, encoded by gyrA), RecA (a DNA-dependent ATPase, encoded by recA), DnaA (the replication initiator protein, encoded by dnaA), trigger factor (encoded by tig) and pyruvate dehydrogenase (encoded by aceE and aceF). All of the above-mentioned genes have been reviewed by Gualerzi et al. [18] and Barria et al. [20].

Because little is known about the adaption of A. butzleri to cold, the growth capabilities of eight A. butzleri isolates derived from human, mussels and chicken were investigated at 8 °C and expression at the transcriptional level of the previously described 13 cold-related genes was studied in three A. butzleri strains showing different growth and survival capability after a temperature down shift from 28 °C to 8 °C.

Materials and methods

Bacterial strains and growth conditions

The A. butzleri strains used in this study were previously described [10,22]. All isolates from human feces (H1, H2 and H3), mussels (M3 and M4) and chicken meat (C1, C2 and C3) were routinely grown in Brucella Broth (BD, Heidelberg, Germany) or on Mueller-Hinton blood agar (Oxoid, Wesel, Germany). The working cultures were maintained under aerobic atmosphere at optimal (28 °C) or cold (8 °C) temperatures.

Growth curve

Briefly, all A. butzleri strains were recovered on Mueller-Hinton blood agars under microaerobic conditions (6% O2, 10% CO2) for 72 h at 28 °C. For each strain, 5 mL of Brucella Broth was inoculated and incubated overnight (18 h) under microaerobic conditions at 28 °C to reach approximately 8 log10 colony-forming units (CFU)-mL⁻¹. These pre-cultures were subsequently diluted 1:10 000 in 5 mL of pre-cooled (8 °C) or pre-warmed (28 °C) Brucella Broth, respectively. The cultures were incubated at the respective temperatures under aerobic conditions and the bacterial load was determined over 28 days. At each indicated time point, serial dilutions of the cultures were plated on Mueller-Hinton blood agar plates and incubated for 48 h at 28 °C under microaerobic conditions before enumeration.

RNA extraction

To screen the transcriptional expression pattern of the presumed cold-related genes in A. butzleri at a very early stage after the sudden temperature downshift, the RNA of A. butzleri isolates from different growth modes at cold temperatures (H2 and C2) and the reference strain (H1/A. butzleri RM4018) were analyzed. Briefly, pre-cultures were prepared as described above but in a volume of 300 mL of fresh Brucella Broth. After overnight incubation, the cultures reached the post-logarithmic phase and were harvested by centrifugation (5 min at 7500 g) at room temperature and resuspended in 200 mL of fresh pre-cooled (8 °C) Brucella Broth. The master culture was split equally into eight working cultures. All working cultures were incubated at 8 °C except the reference sample (0 min), which was further processed immediately. Samples were collected at 5, 30, 60, 120, 180, 240 and 360 min after the temperature downshift. Bacteria from each sample were harvested by centrifugation (5 min at 7500 g) at 8 °C, the pellet resuspended in 1 mL RNA Protect (Qiagen, Hilden, Germany) and stored at −80 °C until further processing. RNAs were isolated by peqGOLD Bacteria RNA Kit (VWR, Radnor, PA, USA) in accordance with the manufacturer’s instructions (Thermo Scientific, Waltham, MA, USA). Afterwards, the RNA was quantified by a NanoDrop 2000 (Thermo Scientific) and subsequently subjected to DNase treatment. Briefly, a total volume of 40 µL containing 750 ng of RNA, 4 U of DNase I, 1 x Reaction Buffer with MgCl2, 40 U of RiboLock (all from Thermo Scientific) and nuclease-free water was incubated at 37 °C for 15 min. The DNase digestion reaction was terminated by addition of 4.5 mM EDTA (Thermo Scientific) and incubation at 65 °C for 10 min. All samples were placed on ice for further steps afterwards.

cDNA transcription

The cDNA of the RNA samples were synthesized using Maxima H Minus First Strand cDNA Synthesis Kit (Thermo Scientific) in accordance with the manufacturer’s instructions. Briefly, the cDNA synthesis mix contained 220 ng of RNA, 0.5 mM dNTP Mix and 5 µM Random
Hexamer Primer in a total volume of 15 µL. The mix was pre-heated to 65 °C for 5 min, before the addition of 4 µL of 5 × RT Buffer and 1 µL of Maxima H Minus Enzyme Mix, and incubated at 25 °C (10 min) followed by 15 min incubation at 50 °C. The reaction was terminated by 5 min incubation at 85 °C. To confirm the efficiency of the DNase digestion, a control without addition of the reverse transcription enzyme of each sample was carried out in parallel. The cDNA was diluted in water at a ratio of 1:10 for reverse transcription-quantitative PCR (RT-qPCR).

Reverse transcription-quantitative PCR

The gene expression profile was analyzed by RT-qPCR. Primers used in this assay were designed using PRIMER3 (http://frodo.wi.mit.edu) and are listed in Table 1. The RT-qPCR was carried out using SsoFastTM EvaGreen Supermix® (Bio-Rad, Munich, Germany) and the CFX Connect Real-Time System (Bio-Rad) in accordance with the manufacturer’s instructions. Briefly, 7.5 µL of SsoFast EvaGreen Supermix, 36 nmol of each forward and reverse primer, and 1 µL of cDNA was mixed in a total volume of 15 µL. The RT-qPCR was started by an initial preheating step for 30 s at 95 °C, followed by 40 cycles of 5 s of denaturation at 95 °C and 5 s of annealing (corresponding annealing temperatures for each gene is shown in Table 1). Here, rpoA was used as housekeeping gene for normalization of expression. Fold changes were calculated according to the \( \Delta\Delta CT \) method [23] and genes with an expression ratio \( \geq 2.0 \) were considered to be up-regulated and those with an expression ratio \( \leq 0.5 \) were considered to be down-regulated.

**Table 1.** List of primers and annealing temperatures used in the RT-qPCR assays.

<table>
<thead>
<tr>
<th>Primer ID</th>
<th>Sequence (5’- to 3’)</th>
<th>Target gene</th>
<th>Amplicon</th>
<th>Annealing</th>
<th>Strain</th>
</tr>
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<tr>
<td>Abu_1472 f</td>
<td>AGC CTT AAT CAC TTG GAG CT</td>
<td>aceE</td>
<td>164</td>
<td>60 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>Abu_1472 r</td>
<td>AAC TCC CCA TGG AGC TCC TC</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Abu_1473 f</td>
<td>GGA GTT GTG GAT ACA CCA</td>
<td>aceF</td>
<td>200</td>
<td>60 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>Abu_1473 r</td>
<td>GGC GTG AAG TAT GTT CCA CC</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Abu_0042 f</td>
<td>TGG ACA AGC GCA TAC AGG TA</td>
<td>deaD</td>
<td>195</td>
<td>60 °C</td>
<td>C2, H1</td>
</tr>
<tr>
<td>Abu_0042 r</td>
<td>GCT GTT CCA CGG TAA ACA GT</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>deaD 1-F</td>
<td>ACAGCAGGTTTGCACTTCC</td>
<td>deaD</td>
<td>167</td>
<td>55 °C</td>
<td>H2</td>
</tr>
<tr>
<td>deaD 1-R</td>
<td>GCTTGAACCCCTGAAACGT</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Abu_0001 f</td>
<td>CGT GTG GGG ACC ATC AAA CC</td>
<td>dnaA</td>
<td>108</td>
<td>60 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>Abu_0001 r</td>
<td>CCA AGT CCC GTT CCA CCA TA</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Abu_2043 f</td>
<td>TGA TAC TCC AGG TCA AGC AG</td>
<td>infB</td>
<td>91</td>
<td>60 °C</td>
<td>C2, H1</td>
</tr>
<tr>
<td>Abu_2043 r</td>
<td>GTC ATC AGC AGC AAC AAC GA</td>
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<tr>
<td>infB 1-F</td>
<td>AGGATTTAATGTAGCACAA</td>
<td>infB</td>
<td>129</td>
<td>52 °C</td>
<td>H2</td>
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<tr>
<td>infB 1-R</td>
<td>CTCAAGATCACCGTAAAGTG</td>
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<td></td>
<td></td>
<td></td>
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<tr>
<td>Abu_1183 f</td>
<td>TGC TGG ACC TAA AGA TGG ACA</td>
<td>pnp</td>
<td>108</td>
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<td>C2, H1, H2</td>
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<tr>
<td>Abu_1183 r</td>
<td>GCT TCA CCA ACC CTG CCT</td>
<td></td>
<td></td>
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<tr>
<td>Abu_2241 f</td>
<td>TAA ATG CAC AAA CGC CAC CA</td>
<td>recA</td>
<td>131</td>
<td>60 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>Abu_2241 r</td>
<td>AGC TTT AGG TGT TGG AGG ACT</td>
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<tr>
<td>Abu_1702 f</td>
<td>ACC GTG AAA TGC AAC ACC AT</td>
<td>tig</td>
<td>112</td>
<td>60 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>Abu_1702 r</td>
<td>TGC TCA ATC ATC TGC ACC AT</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Abu_0246 f</td>
<td>ACC TTT TAT TCC GCC AAT TTC GA</td>
<td>nusA</td>
<td>176</td>
<td>55 °C</td>
<td>C2, H2</td>
</tr>
<tr>
<td>Abu_0246 r</td>
<td>TGG GAA GAA ATG CTT CCA CA</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nusA 1-F</td>
<td>TGG AGA TGT TGT AAA GCC TGG TG</td>
<td>nusA</td>
<td>240</td>
<td>55 °C</td>
<td>H1</td>
</tr>
<tr>
<td>nusA 1-R</td>
<td>ACT CCA ACT ACT GCA CCA AT</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>rfbA 1-F</td>
<td>CGG AAT AGT GTA TTT TAA TGG T</td>
<td>rfbA</td>
<td>127</td>
<td>55 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>rfbA 1-R</td>
<td>ACA TTT ATC CCA ACT TGT ACT TGG</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>cspA 1-F</td>
<td>TGG CAA ATC AAA ATA TGG GAA CA</td>
<td>cspA</td>
<td>197</td>
<td>55 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>cspA 1-R</td>
<td>AGG TGG AGC TTT TTT ATC ATT</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>mr 1-F</td>
<td>AGG GCA TTT TGT TTA TTT TAA ATG ATT CT</td>
<td>mr</td>
<td>205</td>
<td>55 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>mr 1-R</td>
<td>GGC TCC GGT ATT TGG TTA TCT</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>gyrA 1-F</td>
<td>TGG AGC TGG ATT ACC TGG TA</td>
<td>gyrA</td>
<td>192</td>
<td>55 °C</td>
<td>C2, H1</td>
</tr>
<tr>
<td>gyrA 1-R</td>
<td>TCT GCC ATT CTT ACA ACT GCA</td>
<td></td>
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<tr>
<td>gyrA 2-F</td>
<td>TTTGGCGCTGCGTTACTCAGT</td>
<td>gyrA</td>
<td>193</td>
<td>55 °C</td>
<td>H2</td>
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<tr>
<td>gyrA 2-R</td>
<td>TGTCGGTATTCTTACACGTTCA</td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>AB_rpoA ex F</td>
<td>TAGCCAAAATGCGAGG</td>
<td>rpoA</td>
<td>50</td>
<td>60 °C, 55 °C, or 52 °C</td>
<td>C2, H1, H2</td>
</tr>
<tr>
<td>AB_rpoA ex R</td>
<td>CGCAAACCAACTGATGAAC</td>
<td></td>
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</tbody>
</table>

*Depending on the corresponding protocol used.*
regulated [24]. The expression of each gene was analyzed using three independent cDNA samples with two technical replicates in each run.

**Statistical analysis**

Temporal expression patterns of the selected *A. butzleri* genes were analyzed and compared by K-means clustering using GENESIS, version 1.8.1 (TU Graz, Graz, Austria) with three clusters, a maximum of 10 000 iterations and five runs.

The difference in temporal expression pattern among three *A. butzleri* isolates was analyzed by two-way analysis of variance using PRISM, version 6.00 (GraphPad Software Inc., La Jolla, CA, USA).

**Results and Discussion**

**Growth curves of *A. butzleri* isolates**

Because *A. butzleri* is highly prevalent in chicken meat and seafood products, *A. butzleri* strains isolated from these matrices, as well as from a human stool specimen, were included in our cold adaptation study. The growth of eight *A. butzleri* isolates from human (H), mussels (M) and chicken (C) was determined at 28°C and 8°C for 28 days or until undetectable on Mueller-Hinton blood agar plates. No difference in the growth capabilities of all eight strains could be determined during incubation at 28°C under aerobic conditions (data not shown). For six of the eight investigated isolates (75%), stable CFU counts of approx. 4 log_{10} CFU/mL were determined until 8 days after incubation at 8°C, followed by declining CFU counts until 22 days, whereas, afterwards, no colony counts could be determined (Fig. 1). This behavior is represented by strain C2 in Fig. 2. For the reference strain H1 (*A. butzleri* RM4018), a declining tendency of cell counts by 2 log_{10} level was also determined, although colonies were still detectable on Mueller-Hinton blood agar plates at day 28 (Fig. 2). Overall, we determined only survival for strain H1 at 8°C; however, in two of eight replicates, we observed slightly growing capabilities of this strain. By contrast, the strain H2 was able to grow at 8°C, reaching the stationary phase with 9 log_{10} CFU·mL^{-1} around 13 days in all replicates (Fig. 2). No correlation between the strain origin and the growth behavior at 8°C was observed, which is comparable with results of the previous study by Van Driessche and Houf [14].

To the best of our knowledge, 8°C was the lowest temperature to ever been reported for the observation of continued growing up to 5 log_{10} level of *A. butzleri* under aerobic conditions in media. Consistent with a previous study, a low temperature resulted in delayed growth in any medium [13]. In a study by Kjeldgaard et al. [13], *A. butzleri* strain ATCC 49616 (H1 in the present study) was able to grow at 10°C in BHI broth and chicken meat juice and could survive in both media when stored at 5°C. A recent study by Silha et al. [25] observed non-significant multiplication with gradually decreasing viability until 14 days for *A. butzleri* CCUG 30484 (H1 in the present study) at 5°C in BHI culture.

In previous reports, *A. butzleri* isolates were inoculated in different food matrices to investigate their growth and survival capability at different refrigeration temperatures. In ultrahigh-temperature-treated, pasteurized and raw milk, *A. butzleri* could remain viable at both 4°C and 10°C until 6 days [26]. *A. butzleri* RM4018 count remained stable during 5 days of storage at 6°C in artisanal ricotta cheese,
although it decreased by 2.5 log_{10} level during 22 days storage in industrial ricotta cheese. However, the A. butzleri count increased by up to 8 log_{10} CFU·g^{-1} in industrial cheese sample at 7 days and remained stable until the end of the shelf life (22 days) when stored at 12 °C [27]. These results confirmed the hypothesis that the critical growing temperature of the strain A. butzleri RM4018 was between 6 and 12 °C at least in cheese media. Other studies also demonstrate the growing ability of A. butzleri at even lower temperatures, although only over a short time period. Van Driessche and Houf [14] reported slightly increasing cell counts of some A. butzleri strains within 7 days at 4 °C and 7 °C in pure water before the counts declined. Likewise, Badilla-Ramirez et al. [28] tested the survival rate of A. butzleri on chicken legs. Their results showed that A. butzleri had a short growing capacity until 3 days at 4 °C and 10 °C followed by a declining tendency afterwards.

Taken together, we suggest that, even though cell counts of the majority of A. butzleri isolates start to decline after 8 days of storage at 8 °C, some strains were able to maintain a stable cell count or could even grow at this temperature.

The expression profiles of selected A. butzleri genes after cold shock at 8 °C

Even though several studies have described the prolonged survival or growth at lower temperatures for A. butzleri, knowledge of the cellular processes involved in adaptation to cold temperature for this species is still scarce. In general, one of the most important cold-shock response mechanisms of bacterial species is known to sustain the translational processes, which are strongly impeded at low temperatures [29]. For several proteins involved in the cold stress response of E. coli, homologous genes could be determined in the genome of A. butzleri, including the genes encoding for RNA modifying proteins (cspA, rnr, deaD and pnp), translational regulators (infB, rbfA), chromosome modulators (gyrA, recA), transcriptional regulators (musA, dnaA), a protein chaperon (tig) and metabolic enzymes (aceE, aceF). The expression profiles of these putative cold-related genes were investigated by RT-qPCR after a temperature down shift to 8 °C under aerobic conditions for the A. butzleri isolates H2 and C2 (representing different growth tendencies at cold temperatures) and the reference strain H1. Temporal expression patterns of the genes were analyzed and compared by K-means clustering. As shown in Fig. 3, the expression level of 10 of the 13 investigated genes (77%) was up-regulated in response to the cold shock, and the genes were grouped into the high up-regulated cluster I (> 4.5 log_{2} fold change) or the moderate up-regulated cluster II (1.5–3 log_{2} fold change). For all three strains, the expression patterns of cspA, rnr and dnaA were grouped in the high up-regulated cluster I (Fig. 3A), whereas deaD, gyrA, infB, pnp and rbfA were grouped in the moderate up-regulated cluster II (Fig. 3B). Cluster III comprises the genes aceE, aceF and tig, for which the expression levels were non-regulated within the first 6 h after cold shock in all three isolates (Fig. 3C). By contrast to the above mentioned similarities in the gene expression pattern, differences could be observed for the expression pattern of recA and musA in the three investigated A. butzleri strains (Fig. 3D).

To the best of our knowledge, no data describing the cold-shock response of Arcobacter spp. have been published so far; therefore, we compared our data with E. coli, for which the cold-shock response has been intensively studied [19]. Recently, a comprehensive study by Zhang et al. [30] demonstrated a two-member mRNA surveillance system composed of the exonuclease RNase R and the RNA chaperon CspA, enabling the recovery of translation during cold acclimation in E. coli. Enhanced mRNA levels for cspA and rnr were also determined for all three A. butzleri isolates, suggesting utilization of a similar control mechanism. However, similar to E. coli, the mRNA level reached a maximum at 4 h after cold shock and then decreased in strains H1 and H2, whereas it did not decline in strain C2 until the end of the assay.

Furthermore, we determined a similar tendency of gene expression of the other 11 corresponding genes investigated in the present study compared to E. coli [30]. Enhanced expression level for deaD and pnp, both encoding for components of the RNA degradosome [31], have also been determined for all three A. butzleri isolates. However, for E. coli, a higher expression level for deaD could be determined compared to the mRNA level of deaD in A. butzleri.

Similar to our results for A. butzleri, no changes in mRNA level of the genes aceE, aceF (both involved in the pyruvate metabolism) and tig (a protein chaperon, active at the later stage of adaptation to cold temperatures [32]) were reported for E. coli within the investigated 6 h after cold shock [30].

The expression level of dnaA, the chromosomal replication initiator protein, was increased in all three
A. butzleri strains, as well as in E. coli. However, for A. butzleri, the expression pattern was grouped into the highly regulated cluster, whereas its expression pattern in E. coli is only moderately enhanced. However, the precise role of enhanced dnaA mRNA/protein level has not yet been clarified in detail.

In E. coli, the genes nusA-infB-rbfA are located in an operon and mRNA levels for all three genes increased after cold shock [33]. In A. butzleri, only infB and rbfA (Abu_2043 and Abu_2044) are located in the direct neighborhood, whereas nusA (Abu_0246) is located on a different region on the chromosome. Moderate enhanced expression levels of rbfA (encoding a protein rendering the ribosomes to translate non-cold-shock mRNAs under cold conditions [31]) and infB (encoding the translation initiation factor IF2) were determined for all three A. butzleri isolates, which is also in concordance to data reported for E. coli [30]. By contrast, differences in the nusA expression pattern between the three A. butzleri strains were determined. The protein NusA is a transcription factor functionally involved in the regulation of pausing of the RNA polymerase during RNA chain elongation, as well as in the termination of transcription [34,35]. Comparable to E. coli, the expression level of nusA was moderately increased in the strains H1 and H2, whereas it was not increased in the strain C2.

Furthermore, the expression of recA is only moderately induced in the two A. butzleri strains H1 and H2, as well as in E. coli [30], although it is highly induced in the A. butzleri strain C2. RecA is the major DNA recombinase involved in homologues recombination and DNA repair [36]. Another protein involved in the chromosomal structure is the DNA gyrase alpha subunit (GyrA), which regulates chromosomal DNA supercoiling upon cold shock and also provides a level of DNA supercoiling and favors its efficient transcription [32]. The expression of gyrA is regulated by CspA [37] and, consistently, we determined moderately enhanced expression level for gyrA in all three A. butzleri isolates.

Taken together, we have determined several similarities in the cold-shock response of A. butzleri with E. coli. However, additional studies are required to
clarify whether and how differences in the expression levels of *nusA* and *recA* between the three *A. butzleri* might be responsible for the different growth behaviors determined at low temperatures under aerobic conditions.

**Conclusions**

Overall, our data indicate that most of the analysed *A. butzleri* isolates from human, mussel and chicken were not able to grow at 8 °C under aerobic conditions. However, one *A. butzleri* isolate was able to grow to the stationary phase until the end of the assay.

Furthermore, the presence of 13 homologues genes encoding cold shock-related proteins in *E. coli* was determined in the genome of *A. butzleri*, including *cspA*, *rrn*, *deA*, *pnp*, *infB*, *rbfA*, *gyrA*, *recA*, *nusA*, *dnaA*, *tig*, *aceE* and *aceF*. The temporal transcriptional expression profiles of these genes in the early cold stress response of *A. butzleri* are reported for the first time and showed high similarities compared to *E. coli*. These data provide deeper insight into the environmental adaptation capacities of *A. butzleri*.

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**Conflict of interests**

The authors declare that they have no conflicts of interest.

**Author contributions**

GG and TA supervised the study. XZ and GG designed the experiments. XZ and YS performed the experiments. XZ and GG analyzed data. XZ wrote the manuscript. XZ, GG, YS and TA made manuscript revisions.

**Data accessibility**

The raw data of this study are available from the corresponding author on reasonable request.

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