



# ESBL-Producing *Klebsiella pneumoniae* in the Broiler Production Chain and the First Description of ST3128

Katrin Daehre<sup>1\*</sup>, Michaela Projahn<sup>1</sup>, Anika Friese<sup>1</sup>, Torsten Semmler<sup>2</sup>, Sebastian Guenther<sup>3</sup> and Uwe H. Roesler<sup>1</sup>

<sup>1</sup> Institute for Animal Hygiene and Environmental Health, Freie Universität Berlin, Berlin, Germany, <sup>2</sup> NG1-Microbial Genomics, Robert Koch Institute, Berlin, Germany, <sup>3</sup> Pharmaceutical Biology Institute of Pharmacy, Ernst-Moritz-Arndt-Universität Greifswald, Greifswald, Germany

## OPEN ACCESS

### Edited by:

Daniela Ceccarelli,  
Wageningen Bioveterinary Research  
(WBVR), Netherlands

### Reviewed by:

Filipa Grosso,  
Universidade do Porto, Portugal  
Miklos Fuzi,  
Semmelweis University, Hungary

### \*Correspondence:

Katrin Daehre  
katrin.daehre@fu-berlin.de;  
tierhygiene@vetmed.fu-berlin.de

### Specialty section:

This article was submitted to  
Antimicrobials, Resistance and  
Chemotherapy,  
a section of the journal  
Frontiers in Microbiology

**Received:** 14 May 2018

**Accepted:** 10 September 2018

**Published:** 03 October 2018

### Citation:

Daehre K, Projahn M, Friese A,  
Semmler T, Guenther S and  
Roesler UH (2018) ESBL-Producing  
*Klebsiella pneumoniae* in the Broiler  
Production Chain and the First  
Description of ST3128.  
*Front. Microbiol.* 9:2302.  
doi: 10.3389/fmicb.2018.02302

ESBL-producing *Klebsiella pneumoniae* (*K. pneumoniae*) represent an increasing problem both in human and veterinary medicine. As SHV-2 - encoding *K. pneumoniae* were recently detected in the broiler production we were interested in investigating a possible transmission along the broiler production chain and furthermore, in evaluating their possible impact on human health. Therefore, 41 ESBL-producing *K. pneumoniae* originating from a parent flock, from the hatchery's environment during the hatching of that parent flocks' chickens, and from an associated fattening flock were investigated on an Illumina Miseq. Whole genome sequences were analyzed concerning their MLST-type, cgMLST-type, genotypic and phenotypic resistance, plasmid profiles and virulence genes. Irrespective of the origin of isolation all investigated isolates were multi-drug resistant, harbored the same ESBL-gene *bla*<sub>SHV-2</sub>, shared the same sequence type (ST3128) and displayed 100% similarity in core genome multilocus sequence typing (cgMLST). In addition, *in silico* plasmid typing found several Inc/Rep types associated with ESBL-plasmids. Summarizing, identical clones of SHV-2—producing *K. pneumoniae* were detected in different stages of the industrial broiler production in one out of seven investigated broiler chains. This proves the possibility of pseudo-vertical transmission of multi-resistant human pathogens from parent flocks to hatcheries and fattening flocks. Furthermore, the importance of cross-contamination along the production chain was shown. Although the ESBL-producing *K. pneumoniae* clone detected here in the broiler production has not been associated with clinical settings so far, our findings present a potential public health threat.

**Keywords:** extended-spectrum-beta-lactamases, ESBLs, *Klebsiella pneumoniae*, broiler production, broiler chicken

## INTRODUCTION

The emergence of extended-spectrum beta-lactamase-producing Enterobacteriaceae has been of particular interest for years, in both human and veterinary medicine. Especially *Klebsiella pneumoniae* (*K. pneumoniae*), causing community and nosocomial infections of the respiratory and urinary tract as well as bloodstream infections are of critical concern. Resistance against

antimicrobials leads to limited therapeutic options, resulting in increasing difficulties of treatments. In contrast to ESBL-producing *E. coli* which have become very common in veterinary medicine, especially in livestock, companion animals and wildlife, ESBL-producing *K. pneumoniae* were detected rarely in healthy broiler (Hiroi et al., 2012; Yossapol et al., 2017; Mahanti et al., 2018), diseased horses (Vo et al., 2007) dairy cows (Locatelli et al., 2010) and in companion animals (Ewers et al., 2014).

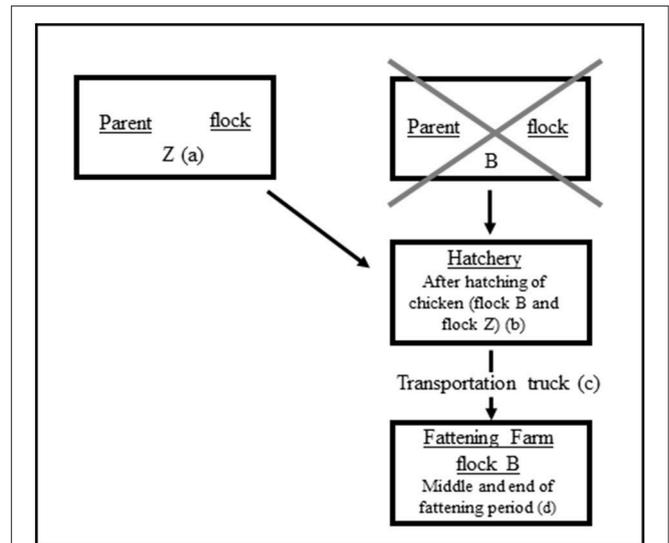
In general, the zoonotic impact of animal-originated pathogens on public health via direct contact or due to the consumption of contaminated meat is assumed (Smet et al., 2010; Marshall and Levy, 2011). This warrants the importance of investigations concerning multi-drug resistant bacteria in food-producing animals at different levels of production, to characterize the impact on humans. In previous studies, we detected ESBL-producing *K. pneumoniae* in a German hatchery (Projahn et al., 2017) as well as in a connected broiler farm (Daehre et al., 2017) in one out of seven investigated broiler chains. To characterize and compare those ESBL-producing *K. pneumoniae* strains detected at different levels of the broiler production we used whole genome sequencing assuring a high resolution of the clonal relationship. We are aiming at revealing possible transmission routes of ESBL-producing *K. pneumoniae* along the production pyramid as well as at assessing a possible impact on human health.

## MATERIALS AND METHODS

### Samples/Flock

In the years 2014–2016, seven German parent flocks, their corresponding hatchlings in the hatchery and later on at the fattening farms as well as the environment of the hatchery and the respective farm were investigated for the occurrence of ESBL-/AmpC-producing Enterobacteriaceae as published by Projahn et al. (2017) and Daehre et al. (2017). There, in one fattening chain (chain B), SHV-2 -producing *K. pneumoniae* strains were detected and those stored bacterial isolates were retrospectively characterized in the present study. In detail, various samples were taken after the hatching of the chicken in the hatchery such as dust, air and swabs from the hatchery's environment. On the fattening farm individual animal samples as well as samples from the housing environment were collected at three different samplings (first day, middle and end of fattening period). In detail, 40 individual animals (cloacal swabs), pooled feces, boot swabs, litter, dust, and air were collected and several surfaces in the barn were swabbed (environmental swabs).

Within this study, additional isolates of a boot swab and a pooled feces sample from another parent flock (flock Z) were investigated as well. This was done due to the fact that the eggs of both parent flocks (B and Z) were bred in the hatchery at the same time. Furthermore, we analyzed isolates originating from environmental swabs from the truck that transported the chicken of parent flock B from the hatchery to the farm. **Figure 1** schematically illustrates the origin of the *bla*<sub>SHV-2</sub> - positive isolates. Detailed information on the investigated isolates are shown in **Table 1** and in **Table S1**.



**FIGURE 1** | Schematic diagram of stages of the broiler production chain tested positive for SHV-2-producing *Klebsiella pneumoniae*. Parent flock B was negative. Positive samples: (a)-pooled feces; (b)-egg shells, environmental swab; (c)-swabs; (d)-cloacal swabs, pooled feces, boot swab, litter, dust, air, environmental swabs.

### Laboratory Methods

All samples were processed as described by Projahn et al. (2017) and Daehre et al. (2017). Finally, the samples were streaked out on MacConkey No. 3 (Oxoid, Wesel, Germany) agar plates with the addition of 1 mg/l cefotaxime (AppliChem, Darmstadt, Germany). The species of all isolates with Enterobacteriaceae-like phenotypes were determined by MALDI-TOF analyses. The detection of the most common class A beta-lactamase-genes including *bla*<sub>SHV</sub> was performed as described by Roschanski et al. (2014) and verified by sequencing using the same primer set as published by Projahn et al. (2017).

Forty-one *K. pneumoniae* isolates with *bla*<sub>SHV-2</sub> - genes from 26 different samples (up to three isolates per sample) were chosen for further characterization. These isolates originated from the other parent flock Z, the hatchery, the truck (transport of chicken from hatchery to farm) as well as from samples from the middle and the end of the fattening period on the farm (cloacal swabs and samples from the housing environment; **Figure 1**, **Table 1**, and **Table S1**).

The Vitek<sup>®</sup>2 system (BioMérieux, Germany; card GN38) was used to determine phenotypic antimicrobial resistance to various  $\beta$ -lactam-antibiotics and other classes of antimicrobials.

To get more information concerning the phylogenetic relationship of the different samples, whole genome sequencing (WGS) was performed. Therefore, DNA was extracted with the MasterPure<sup>™</sup> DNA purification kit (Epicenter, Illumina) and Illumina MiSeq 300-bp paired-end with a coverage between 50x and 120x was used. Following a quality control performed with the NGS tool kit (Patel and Jain, 2012) high quality reads were defined (minimum of 70% of bases having a phred score higher than 20) and *de novo* assembled into contiguous sequences

**TABLE 1** | Information on seven *K. pneumoniae* isolates (ST3128) including sampling timepoint, sample type, isolate ID, plasmid type, genotypic, and phenotypic resistance.

Sampling timepoint	Sample type	Isolate ID	Plasmid type	Genotypic resistance	Phenotypic resistance
I	PF	ITU10028	IncR, IncFIB, IncFII, IncHI1B, Col	<i>bla</i> <sub>SHV-2</sub> , <i>bla</i> <sub>SHV-1</sub> , <i>fosA</i> -like, <i>sul1</i> , <i>dfrA12</i> , <i>tet(D)</i> , <i>aadA1</i> , <i>aadA2</i> , <i>aac(3)</i> -like, <i>oqxA</i> -like, <i>oqxB</i> -like, <i>parC</i> S80I mutation	
II	EggS	ITU10022	IncR, IncFIB, IncFII, IncHI1B, IncX1, IncX3, Col	<i>bla</i> <sub>SHV-2</sub> , <i>bla</i> <sub>SHV-1</sub> , <i>fosA</i> -like, <i>sul1</i> , <i>dfrA12</i> , <i>tet(D)</i> , <i>aadA1</i> , <i>aadA2</i> , <i>aac(3)</i> -like, <i>oqxA</i> -like, <i>oqxB</i> -like, <i>parC</i> S80I mutation	
III	EnvS	ITU10024,	IncR, IncFIB, IncFII, IncHI1B, Col	<i>bla</i> <sub>SHV-2</sub> , <i>bla</i> <sub>SHV-1</sub> , <i>fosA</i> -like, <i>sul1</i> , <i>dfrA12</i> , <i>tet(D)</i> , <i>aadA1</i> , <i>aadA2</i> , <i>aac(3)</i> -like, <i>oqxA</i> -like, <i>oqxB</i> -like, <i>parC</i> S80I mutation	AMX, AMC, AMP, SAM, CFR, CFL, CLT, CFZ, CFM, CPZ, CTX, CFV, CFR, CPD, CAZ, CEX, CFX, DOX, ENR, GEN, MAR, PIP, PUF, TET, TOB, SXT
IV	CS	ITU3949	IncR, IncFIB, IncFII, Col	<i>bla</i> <sub>SHV-2</sub> , <i>bla</i> <sub>SHV-1</sub> , <i>fosA</i> -like, <i>sul1</i> , <i>dfrA12</i> , <i>tet(D)</i> , <i>aadA1</i> , <i>aadA2</i> , <i>aac(3)</i> -like, <i>oqxA</i> -like, <i>oqxB</i> -like, <i>parC</i> S80I mutation	
	Litter	ITU3854	IncR, IncFIB, IncFII, IncHI1B, Col	<i>bla</i> <sub>SHV-2</sub> , <i>bla</i> <sub>SHV-1</sub> , <i>fosA</i> -like, <i>sul1</i> , <i>dfrA12</i> , <i>tet(D)</i> , <i>aadA1</i> , <i>aadA2</i> , <i>aac(3)</i> -like, <i>oqxA</i> -like, <i>oqxB</i> -like, <i>parC</i> S80I mutation	
V	CS 18	ITU4179	IncR, IncFIB, IncFII, IncHI1B, Col	<i>bla</i> <sub>SHV-2</sub> , <i>bla</i> <sub>SHV-1</sub> , <i>fosA</i> -like, <i>sul1</i> , <i>dfrA12</i> , <i>tet(D)</i> , <i>aadA1</i> , <i>aadA2</i> , <i>aac(3)</i> -like, <i>oqxA</i> -like, <i>oqxB</i> -like, <i>QnrS1</i> -like, <i>parC</i> S80I mutation	
	BS	ITU4097	IncR, IncFIB, IncFII, IncHI1B, Col	<i>bla</i> <sub>SHV-2</sub> , <i>bla</i> <sub>SHV-1</sub> , <i>fosA</i> -like, <i>sul1</i> , <i>dfrA12</i> , <i>tet(D)</i> , <i>aadA1</i> , <i>aadA2</i> , <i>aac(3)</i> -like, <i>oqxA</i> -like, <i>oqxB</i> -like, <i>parC</i> S80I mutation	

I, parent flock Z; II, hatchery, after hatching; III, truck (from hatchery to farm); IV, fattening farm, middle of fattening period; V, fattening farm, end of fattening period; PF, pooled feces; EggS, pooled eggshells; EnvS, environmental swab; CS, cloacal swab, BS, boot swab; ITU, Institut für Tier- und Umwelthygiene; AMX, amoxicillin; AMC, Amoxicillin/clavulanic acid; AMP, ampicillin; SAM, ampicillin/sulbactam; CFR, cefadroxil; CFL, cefalexin; CLT, cefalotin; CFZ, cefazolin; CFM, cefixime; CPZ, cefoperazone; CTX, cefotaxime; CFV, cefovecin; CFR, cefpirome; CPD, cefpodoxime; CAZ, ceftazidime; CEX, ceftiofur; CFX, cefuroxime; DOX, doxycyclin; ENR, enrofloxacin; GEN, gentamicin; MAR, marbofloxacin; PIP, piperacillin; PUF, prulifloxacin; TET, tetracyclin; TOB, tobramycin; SXT, trimethoprim/sulfamethoxazol.

(contigs) using CLC Genomics workbench 9.0 (Qiagen, Venlo, Netherlands). These sequence data have been deposited at DDBJ/ENA/GenBank and the accession numbers can be found in the Table S2.

The *bla*<sub>SHV</sub> genes were sequenced using the same primer set as published by Projahn et al. (2017) and evaluated with BioNumerics 6.6.

WGS data were used for genotypic characterization utilizing the Center for Genomic Epidemiology (Center for Genomic Epidemiology, 2018): multi-locus sequence types [MLST; MLSTFinder 1.8 (Larsen et al., 2012)], plasmids [PlasmidFinder 1.3 (Carattoli et al., 2014)] and resistance genes [ResFinder3.0 (Zankari et al., 2012)] were determined. Additionally, core genome MLST (cgMLST) typing was performed using the cgMLST.org Nomenclature Server<sup>1</sup> and Ridom Seqsphere 4.1 (Ridom GmbH, Muenster, Germany). Within cgMLST, for *K. pneumoniae*, 2358 conserved genome-wide genes are compared, resulting in a very high discriminatory power.

BLAST analyses of the assembled contigs were done and the accordance of *bla*<sub>SHV</sub>-carrying contigs with SHV-encoding plasmids described for *K. pneumoniae* (JX461340.1, CP025463, CP025458, DQ449578, JN247852, and others) were checked using the European nucleotide archive<sup>2</sup>, the European Center for Biotechnology Information<sup>3</sup> and features of Geneious v. 7.1.2 (Kearse et al., 2012) and DNASTAR<sup>®</sup> Lasergene 11 SeqMan Pro<sup>™</sup> (version 11.2.1). The genetic vicinity of the *bla*<sub>SHV</sub> region as well as the occurrence of genes known for the association with virulent *K. pneumoniae* (*magA*, *rmpA*, *entB*, *iutA*, *YbtS*, *Kfu*, *allS*, *mrkD*, *wzi*; Compain et al., 2014) were investigated using the same tools.

## RESULTS AND DISCUSSION

SHV-2 -encoding *K. pneumoniae* strains were detected in all investigated stages of the broiler production chain: 25% of the

<sup>2</sup>European Nucleotide Archive. Available online at: <https://www.ebi.ac.uk/ena> (Accessed April 05, 2018).

<sup>3</sup>National Center for Biotechnology Information. Available online at: <https://www.ncbi.nlm.nih.gov/> (Accessed April 05, 2018).

<sup>1</sup>cgMLST.org Nomenclature Server. Available online at: <http://www.cgmlst.org/ncs> (Accessed April 05, 2018).

samples from parent flock Z (1/4) (but not in samples from parent flock B), 22.2% from the hatchery's environment (2/9) and 28.8% of individual animals (23/80) resp. 68.2% of samples from the housing environment (15/22) of flock B (middle and end of fattening period) were tested positive. All 41 *K. pneumoniae* isolates were multi-drug resistant, harbored the genes for the ESBL beta-lactamase SHV-2 and were assigned to the newly described MLST type ST3128. cgMLST typing revealed 100% similarity. Therefore, a clonal relationship between the isolates detected at the different stages of the broiler production chain can be stated.

Antibiotic resistant *K. pneumoniae* isolates, especially ESBL- and/or carbapenemase-producers with resistance toward third/fourth generation cephalosporins and carbapenems are of great concern in both human and veterinary medicine. To our knowledge, this is the first finding of ESBL-producing *K. pneumoniae* detected in various stages within the same broiler production chain. To get more information on these strains, 41 isolates detected in samples from a parent flock, the hatchery, the transportation vehicle and a fattening flock of one out of seven investigated broiler chains were further characterized.

All *K. pneumoniae* isolates harbor the *bla<sub>SHV-2</sub>* - gene, belong to the previously unknown *K. pneumoniae* MLST type ST3128 and show phenotypical resistance against various antimicrobials including third and fourth generation cephalosporins, but not against carbapenems (Table 1 and Table S1). With regard to the fluoroquinolone resistance, all 41 isolates have a mutation (S80I) in the *parC* gene known for fluoroquinolone-resistant *K. pneumoniae* (Correia et al., 2017) but mutations in the QRDR-region of the *gyrA* gene were not detected. All investigated *K. pneumoniae* isolates possess *entB* and *mrkD*, but none of the other genes, associated with virulence of *K. pneumoniae* (*magA*, *rmpA*, *iutA*, *YbtS*, *Kfu*, *allS*, *wzi*). Enterobactin (*entB*) is a prototypical catecholate siderophore as part of iron acquisition systems. But *entB* only is known for virulence when occurring in a combination with iron acquisition systems (*iutA*, *YbtS*, *Kfu*). The same applies to *mrkD*. *mrkD* is believed to function as the type 3 fimbrial adhesion and mediates binding to extracellular matrix (Jagnow and Clegg, 2003) but in virulent *K. pneumoniae* strains only occurs in combination with other virulence factors, which were tested negative in our isolates. Therefore, the 41 ESBL-producing *K. pneumoniae* strains, detected in healthy broiler chicken, do not harbor virulence genes that were described in any clinical association. Inc typing using plasmidFinder found the plasmid types IncR, IncFIB, IncFII, and IncHI1B known for the presence of ESBL-encoding genes in all strains (Table 1 and Table S1). The DNA sequences of the *bla<sub>SHV-2</sub>* carrying contigs of ~7,400 bp length were homologous to plasmid p1658/97 from *Escherichia coli* (accession number: AF550679) and plasmid pSEM from *Salmonella enterica* (AJ245670), containing a *recF* gene upstream and a *deoR* gene downstream from the *bla<sub>SHV-2</sub>*. These genes are also present in a *bla<sub>SHV</sub>*-carrying plasmid of *K. pneumoniae* published by Yu et al. (2006). The *recF* gene may contribute to the mobilization of the *bla* gene to a plasmid via the *recF* recombination pathway (Kolodner et al., 1985).

Additionally, a *bla*-SHV-2 carrying fragment (~3,500 bp) of the contigs was detected in plasmids of *K. pneumoniae*

(JX461340.1, CP025463, CP025458, DQ449578, JN247852) (more than 99% identity). The adjacent DNA sequence was identical to other plasmids (CP027613, LT985275, and others). These findings make the location of the *bla<sub>SHV-2</sub>* on a plasmid very likely.

To elucidate the epidemiological relationship and, therefore, the transmission dynamics along the production chain, cgMLST was performed. cgMLST, comparing 2358 genes for *K. pneumoniae*, revealed 100% similarity. This demonstrates that identical clones of *K. pneumoniae* (ST3128), encoding for SHV-2 were detected in the different stages of the broiler production chain pointing toward ongoing transmission processes.

The circulation of ESBL-/AmpC-producing Enterobacteriaceae along the broiler production process was described in previous studies (Dierikx et al., 2013; Nilsson et al., 2014). Projahn et al. (2017) conducted transmission investigations with a special focus on the hatchery and *Escherichia coli* (*E. coli*). There, the introduction of ESBL-producing *E. coli* strains directly from the parent flock into the hatchery, despite the eggs' disinfection, was shown. Additionally, a pseudo-vertical transmission, in detail, the introduction of ESBL-producing Enterobacteriaceae into the hatchery and the chickens' colonization by the uptake of resistant bacteria from the environment of the hatchery was discussed.

Our results reinforce these hypotheses. Parent flock Z (the other parent flock) was tested positive for SHV-2-producing *K. pneumoniae*. In the hatchery, clones of this strain were detected after the chickens hatching in the hatchery's environment (environmental swab and eggshells). We showed that the chicken of flock B, that hatched at the same time in the hatchery were colonized with those clones. Furthermore, the SHV-2-producing *K. pneumoniae* were detected during the whole fattening period of flock B both, in individual animals as well as in samples from the environment. This clearly demonstrates a transmission from parent flocks, via the hatchery, into the fattening flocks. This is in accordance to Projahn et al. (2018). They showed that ESBL-producing *E. coli* were already present in the hatchery and colonized the recently hatched chickens. Therefore, they also confirmed the hatchery as a contamination source for the fattening period.

In our study, clones of the SHV-2-producing *K. pneumoniae* were detected in parent flock Z, in the hatchery after the chickens hatching and during the fattening period of flock B. As the eggs of parent flock Z were bred at the same time as the eggs of flock B, the chicken of flock Z and B hatched at the same time in the same surrounding. This means the original source of the ESBL-producing *K. pneumoniae* strains was not the respective parent flock B but an unrelated parent flock. This clearly emphasizes the importance of cross-contamination via the environment, especially at hatchery-level.

As described by Dierikx et al. (2013) the broiler production system seems rather simple: only a few breeding companies produce broilers for many farms. The breeding eggs of several parent flocks are processed in a few hatcheries and the hatched broiler chicken are delivered to various fattening farms. Thus, the introduction of resistant bacteria from one parent flock into the hatchery can cause the spread of these strains in several

fattening farms. Consequently, the absence of antibiotic resistant bacteria in parent flocks is essential to minimize the occurrence of these bacteria in the production pyramid and one high prevalent parent flock has the potential to contaminate all hatchlings of one course of hatching.

Besides, *K. pneumoniae* clones were also detected in the truck, transporting the chicken from the hatchery to the farms (environmental swabs from the truck's ground and walls). Projahn et al. (2018) also detected ESBL-producing Enterobacteriaceae in a transportation truck and indicated a transmission of resistant bacteria via the transportation process. As the trucks transport animals for several farms, cross-contamination even during the transport could be of importance as well.

As described above, the *K. pneumoniae* isolates were detected during the investigation on ESBL-/AmpC-producing Enterobacteriaceae in parent flocks, the hatchery and at fattening farms (Daehre et al., 2017; Projahn et al., 2017). Thus, next to the *K. pneumoniae*, several ESBL-/AmpC-producing *E. coli* were detected. These *E. coli* isolates encoded for SHV-12, CMY-2 or CTX-M-1, but not for SHV-2, in contrast to the *K. pneumoniae* isolates. Therefore, with our investigated isolates we cannot show any association between ESBL-producing *E. coli* and ESBL-producing *K. pneumoniae* in the broiler production chain.

The detected antibiotic resistant *K. pneumoniae* strains of ST3128 have not been reported in clinical settings, yet, and, therefore, did not have an impact on human health so far. However, plasmids with resistance genes are transferable between strains and species. Therefore, resistance-carrying plasmids detected in food-producing animals always pose a possible risk for human health.

In summary, our results demonstrate the presence of SHV-2-producing *K. pneumoniae* clones in several stages of the broiler production pyramid. A pseudo-vertical transmission of ESBL-producing *K. pneumoniae*, resulting in a positive fattening flock caused by the uptake of bacteria that were introduced into the hatchery by another parent flock was shown for the first time.

## REFERENCES

- Carattoli, A., Zankari, E., García-Fernández, A., Voldby Larsen, M., Lund, O., Villa, L., et al. (2014). *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. *Antimicrob. Agents Chemother.* 58, 3895–3903. doi: 10.1128/AAC.02412-14
- Center for Genomic Epidemiology, (2018). Available online at: <http://www.genomicepidemiology.org/> (Accessed April 05, 2018).
- Compain, F., Babosan, A., Brisse, S., Genel, N., Audo, J., Ailloud, F., et al. (2014). Multiplex PCR for detection of seven virulence factors and K1/K2 capsular serotypes of *Klebsiella pneumoniae*. *J. Clin. Microbiol.* 52, 4377–4380. doi: 10.1128/JCM.02316-14
- Correia, S., Poeta, P., Hébraud, M., Capelo, J. L., and Igrejas, G. (2017). Mechanisms of quinolone action and resistance: where do we stand? *J. Med. Microbiol.* 66, 551–559. doi: 10.1099/jmm.0.000475
- Daehre, K., Projahn, M., Semmler, T., Roesler, U., and Friese, A. (2017). Extended-spectrum beta-lactamase-/AmpC beta-lactamase-producing enterobacteriaceae in broiler farms: transmission dynamics at farm level. *Microb. Drug Resist.* 24, 511–518. doi: 10.1089/mdr.2017.0150

This also indicates the importance of cross-contamination. As *K. pneumoniae* of ST3128 have not been known previously, clones of our strains have never been reported in clinical associations. Therefore, to date, these strains did not have an impact on human health. Nevertheless, a reduction of antibiotic-resistant bacteria in food-producing animals should be achieved, in order to not worsen the situation in human and veterinary medicine.

## AUTHOR CONTRIBUTIONS

UR and AF designed the study. KD and MP performed the samplings and the laboratory work. TS and SG performed bioinformatic analysis. KD analyzed the data and wrote the manuscript. All authors have read and approved the final draft of the article.

## FUNDING

This project was funded by the Federal Ministry of Education and Research (grant 01KI1313C) and is part of the RESET research consortium (<http://www.reset-verbund.de/index.htm>).

## ACKNOWLEDGMENTS

We thank the employees from the hatchery, the farm and all coordinating staff for their kind cooperation and permanent support. Many thanks to Nicole Roschanski for scientific advice and Maja Thieck for excellent technical assistance in the laboratory. We thank the team of the curators of the Institute Pasteur MLST system (Paris, France) for importing novel profiles and isolates at <http://bigsdweb.pasteur.fr>.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmicb.2018.02302/full#supplementary-material>

- Dierikx, C. M., van der Goot, J. A., Smith, H. E., Kant, A., and Mevius, D. J. (2013). Presence of ESBL/AmpC-producing *Escherichia coli* in the broiler production pyramid: a descriptive study. *PLoS ONE* 8:e79005. doi: 10.1371/journal.pone.0079005
- Ewers, C., Stamm, I., Pfeifer, Y., Wieler, L. H., Kopp, P. A., Schönning, K., et al. (2014). Clonal spread of highly successful ST15-CTX-M-15 *Klebsiella pneumoniae* in companion animals and horses. *J. Antimicrob. Chemother.* 69, 2676–2680. doi: 10.1093/jac/dku.217
- Hiroi, M., Yamazaki, F., Harada, T., Takahashi, N., Iida, N., Noda, Y., et al. (2012). Prevalence of extended-spectrum  $\beta$ -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in food-producing animals. *J. Vet. Med. Sci.* 74, 189–195. doi: 10.1292/jvms.11-0372
- Jagnow, J., and Clegg, S. (2003). *Klebsiella pneumoniae* MrkD-mediated biofilm formation on extracellular matrix- and collagen-coated surfaces. *Microbiology* 149, 2397–2405. doi: 10.1099/mic.0.26434-0
- Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., et al. (2012). Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28, 1647–1649. doi: 10.1093/bioinformatics/bts199

- Kolodner, R., Fishel, R. A., and Howard, M. (1985). Genetic recombination of bacterial plasmid DNA: effect of RecF pathway mutations on plasmid recombination in *Escherichia coli*. *J. Bacteriol.* 163, 1060–1066.
- Larsen, M. V., Cosentino, S., Rasmussen, S., Friis, C., Hasman, H., Marvig, R. L., et al. (2012). Multilocus sequence typing of total-genome-sequenced bacteria. *J. Clin. Microbiol.* 50, 1355–1361. doi: 10.1128/JCM.06094-11
- Locatelli, C., Scaccabarozzi, L., Pisoni, G., and Moroni, P. (2010). CTX-M1 ESBL-producing *Klebsiella pneumoniae* subsp. *pneumoniae* isolated from cases of bovine mastitis. *J. Clin. Microbiol.* 48, 3822–3823. doi: 10.1128/JCM.00941-10
- Mahanti, A., Ghosh, P., Samanta, I., Joardar, S. N., Bandyopadhyay, S., Bhattacharyya, D., et al. (2018). Prevalence of CTX-M-Producing *Klebsiella* spp. in broiler, kuroiler, and indigenous poultry in West Bengal State, India. *Microb. Drug Resist.* 24, 299–306. doi: 10.1089/mdr.2016.0096
- Marshall, B. M., and Levy, S. B. (2011). Food animals and antimicrobials: impacts on human health. *Clin. Microbiol. Rev.* 24, 718–733. doi: 10.1128/CMR.00002-11
- Nilsson, O., Börjesson, S., Landén, A., and Bengtsson, B. (2014). Vertical transmission of *Escherichia coli* carrying plasmid-mediated AmpC (pAmpC) through the broiler production pyramid. *J. Antimicrob. Chemother.* 69, 1497–1500. doi: 10.1093/jac/dku030
- Patel, R. K., and Jain, M. (2012). NGS QC Toolkit: a toolkit for quality control of next generation sequencing data. *PLoS ONE* 7:e30619. doi: 10.1371/journal.pone.0030619.g001
- Projahn, M., Daehre, K., Roesler, U., and Friese, A. (2017). Extended-spectrum-beta-lactamase- and plasmid-encoded cephamycinase-producing enterobacteria in the broiler hatchery as a potential mode of pseudo-vertical transmission. *Appl. Environ. Microbiol.* 83:e02364-16. doi: 10.1128/AEM.02364-16
- Projahn, M., Daehre, K., Semmler, T., Guenther, S., Roesler, U., and Friese, A. (2018). Environmental adaptation and vertical dissemination of ESBL-/pAmpC-producing *Escherichia coli* in an integrated broiler production chain in the absence of an antibiotic treatment. *Microb. Biotechnol.* doi: 10.1111/1751-7915.13040. [Epub ahead of print].
- Roschanski, N., Fischer, J., Guerra, B., and Roesler, U. (2014). Development of a multiplex real-time PCR for the rapid detection of the predominant beta-lactamase genes CTX-M, SHV, TEM and CIT-type AmpCs in *Enterobacteriaceae*. *PLoS ONE* 9:e100956. doi: 10.1371/journal.pone.0100956
- Smet, A., Martel, A., Persoons, D., Dewulf, J., Heyndrickx, M., Herman, L., et al. (2010). Broad-spectrum  $\beta$ -lactamases among *Enterobacteriaceae* of animal origin: molecular aspects, mobility and impact on public health. *FEMS Microbiol. Rev.* 34, 295–316. doi: 10.1111/j.1574-6976.2009.00198.x
- Vo, A. T., van Duijkeren, E., Fluit, A. C., and Gastra, W. (2007). Characteristics of extended-spectrum cephalosporin-resistant *Escherichia coli* and *Klebsiella pneumoniae* isolates from horses. *Veter. Microbiol.* 124, 248–255. doi: 10.1016/j.vetmic.2007.04.027
- Yosapal, M., Sugiyama, M., and Asai, T. (2017). The occurrence of CTX-M-25-producing *Enterobacteriaceae* in day-old broiler chicks in Japan. *J. Veter. Med. Sci.* 79, 1644–1647. doi: 10.1292/jvms.17-0294
- Yu, W. L., Chen, S. C., Hung, S. W., Chuang, Y. C., Chung, J. G., Chen, I. C., et al. (2006). Genetic association of blaSHV-5 with transposable elements IS26 and IS5 in *Klebsiella pneumoniae* from Taiwan. *Clin. Microbiol. Infect.* 12, 806–809. doi: 10.1111/j.1469-0691.2006.01488.x
- Zankari, E., Hasman, H., Cosentino, S., Vestergaard, M., Rasmussen, S., Lund, O., et al. (2012). Identification of acquired antimicrobial resistance genes. *J. Antimicrob. Chemother.* 67, 2640–2644. doi: 10.1093/jac/dks261

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Daehre, Projahn, Friese, Semmler, Guenther and Roesler. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.