

## 5. Summary

In recent years, adult stem cells have become the subject of intense study. They have been analyzed in the context of normal physiology and have been associated with the development and propagation of some benign and malignant tumors. The presence of adult stem cells in human thyroid nodule tissues was first reported in our previous study. However, the isolation of thyroid stem cells has so far been hampered by the absence of a surface antigen that allows the separation by FACS. Therefore, the present study aimed to (1) achieve progresses in thyroid stem cell enrichment and purification, (2) analyze under which conditions quiescent stem cells derived from human nodules can be propagated to outgrow and (3) clarify if these cells have retained the capacity to differentiate into thyroid cells.

Taking advantage of the fact that stem cells express ABCG2 membrane transporters, thyroid stem cells from human nodules were separated by FACS analysis as a side population (SP), from a non-side population (non-SP) of cells that consisted of endodermal progenitor and differentiated cells, characterized by their typical gene profiles. Moreover, thyroid SP exhibited a characteristic morphology of adult stem cells, as compared with non-SP cells, after cytopspin and Giemsa staining.

However, proliferation and differentiation of SP cells were limited by their poor viability and their quiescent state. To overcome this problem new isolation and stimulation techniques were employed. A small number of thyroid stem cells grew out from primary cell cultures in response to intense growth stimulation to form non-adherent, three-dimensional spheroid clones, which we termed 'thyro-spheres'. These spheres consisted of highly proliferating stem and progenitor cells with their characteristic expression profiles. Upon differentiation induction with serum and TSH, these sphere-forming cells grew as monolayer and differentiated into thyrocytes that expressed PAX8, thyroglobulin, sodium iodide symporter, thyroid-stimulating hormone receptor, and thyroperoxidase mRNA. Importantly, when embedded in collagen, these cells showed specific <sup>125</sup>I iodide uptake activity, a hallmark of differentiated thyroid cells.

This study demonstrates that human thyroid nodule tissues contain a population of stem cells with SP phenotype and the capacity of clonal expansion in response to intense growth stimulation. These stem cells express typical gene profiles and are characterized by the ability of

self-renewal and differentiation into thyroid cells. Compared to thyrocytes, stem cells display a much higher proliferation rate upon acute growth stimulation which suggests a putative role of stem cells in the chronic growth factor-stimulated nodular transformation of the thyroid.

Further work is necessary to analyze how stem cell growth may potentially contribute to neoplastic thyroid growth that arises in nodular goiters and has to focus on the molecular and cellular aberrations that may occur on the long way from adult stem cells to differentiated thyroid cells *in vivo* and may be at the very beginning of thyroid tumorigenesis.

## **Zusammenfassung in deutscher Sprache**

In den letzten Jahren sind adulte Stammzellen Gegenstand intensiver Forschung sowohl hinsichtlich ihrer physiologischen Bedeutung als auch in Hinblick auf ihre mögliche Bedeutung für die Entstehung und das Wachstum benigner und maligner Tumoren geworden. In den vorausgehenden Arbeiten unserer Arbeitsgruppe konnte zum ersten Mal der Nachweis von adulten Stammzellen in menschlichen Schilddrüsengeweben geführt werden. Bisher ist jedoch die Isolation von Schilddrüsenstammzellen aus dem Schilddrüsengewebe am Fehlen spezifischer Oberflächlichen-Antigenen gescheitert. Solche Antigenmarker sind für die Selektion und Isolierung intakter Zellen mittels FACS erforderlich. In der vorliegenden Arbeit sollten daher 1. die Anreicherungs- und Isolierungs-Techniken für Schilddrüsenstammzellen etabliert werden, 2. die Bedingungen analysiert werden, unter denen die aus menschlichen Schilddrüsenknoten stammende, ruhenden (d.h. nicht proliferierenden) Stammzellen zum Wachstum gebracht werden können und 3. die Frage geklärt werden, ob diese Zellen das Potenzial zur Differenzierung in Schilddrüsenzellen beibehalten haben.

Die in dieser Arbeit angewandte Isolationstechnik von Schilddrüsenstammzellen aus menschlichen Schilddrüsenknoten mittels FACS basierte auf dem Nachweis der Expression von ABCG2-Membrantransportern. Aufgrund dieser typischen Genexpressionsprofile konnten eine Seitenpopulation (sogen. side population) von Stammzellen (SP-Zellen) von den übrigen Zellen der Schilddrüse (non-SP-Zellfraktion), bestehend aus endodermalen Vorläuferzellen und differenzierten Thyreozyten, differenziert werden. Morphologisch wiesen die SP-Zellen lichtmikroskopisch eine charakteristische Stammzellmorphologie.

Die Vermehrung und Differenzierung von isolierten Stammzellen wurde jedoch durch ihren Ruhezustand (sogen. quiescent state) und ihre verminderte Lebensfähigkeit *in vitro* begrenzt. Dieses Problem konnte durch neue Isolation- und Stimulationsmethoden überwunden werden. Nach intensiver Wachstumsstimulation in Co-Kultur mit normalen Schilddrüsenzellen und sogenannten Nischenzellen wuchs eine kleine Zahl von Schilddrüsenstammzellen aus primären Zellkulturen heran und bildete dreidimensionale kugelförmige Klone. Diese nannten wir „thyro-spheres“. Diese „thyro-spheres“ bestanden aus sich stark vermehrenden Stamm- und Vorläuferzellen mit typischen Genexpressionsprofilen. Nach Induktion der Differenzierung durch Serum und TSH, wuchsen diese „thyro-spheres“-Zellen als Monolayer aus und ließen sich in Thyreozyten mit

typischen mRNA-Expressionsmustern (PAX8-, Thyreoglobulin-, Natrium-Jodid-Symporter-, TSH-Rezeptor- und Thyreoperoxidase- mRNA) differenzieren. Von besonderer Bedeutung war der Befund, dass nach Einbetten in Kollagen diese Zellen eine spezifische <sup>125</sup>Jodid-Aufnahme aufwiesen, was ein Kennzeichen für differenzierte Schilddrüsenzellen ist.

Diese Arbeit zeigt, dass menschliche Schilddrüsenknotengewebe Stammzellen enthalten, die einen SP-Phänotyp besitzen und ein Potenzial zur klonalen Expansion nach intensiver Wachstumsstimulation aufweisen. Diese Stammzellen exprimieren typische Genprofile und besitzen unter charakteristischen Kulturbedingungen ein Eigenvermehrungs- und Differenzierungspotenzial. Im Vergleich zu Thyreozyten, weisen diese Stammzellen nach akuter Wachstumsstimulation eine erhöhte Proliferationsrate auf. Dies könnte auf eine mögliche Rolle von Stamm- bzw. Progenitorzellen bei der durch chronische Wachstumsstimulation induzierten, nodulären Transformation der Schilddrüse hindeuten.

Weitere Studien sind erforderlich, um die Bedeutung des Stammzellwachstums für die Entstehung neoplastischen Schilddrüsenwachstums zu analysieren. Zukünftige Forschung sollte dabei auf die molekularen und zellulären Aberrationen fokussieren, die während des langen Weges von adulten Stammzellen bis zur differenzierten Thyreozyten *in vivo* auftreten können und Ausgangspunkt der Schilddrüsentumorgenese sein könnten.

## 6. Reference

- 1 Vander JB, Gaston EA, Dawber TR. The significance of nontoxic thyroid nodules. Final report of a 15-year study of the incidence of thyroid malignancy. *Ann Intern Med* 1968;69:537-40.
- 2 Ezzat S, Sarti DA, Cain DR, et al. Thyroid incidentalomas. Prevalence by palpation and ultrasonography. *Arch Intern Med* 1994;154:1838-40.
- 3 Derwahl M, Studer H. Hyperplasia versus adenoma in endocrine tissues: are they different? *Trends Endocrinol Metab* 2002;13:23-8.
- 4 Aghini-Lombardi F, Antonangeli L, Martino E, et al. The spectrum of thyroid disorders in an iodine-deficient community: the Pescopagano survey. *J Clin Endocrinol Metab* 1999;84:561-6.
- 5 Studer H, Derwahl M. Mechanisms of nonneoplastic endocrine hyperplasia--a changing concept: a review focused on the thyroid gland. *Endocr Rev* 1995;16:411-26.
- 6 Krohn K, Fuhrer D, Bayer Y, et al. Molecular pathogenesis of euthyroid and toxic multinodular goiter. *Endocr Rev* 2005;26:504-24.
- 7 Coclet J, Foureau F, Ketelbant P, et al. Cell population kinetics in dog and human adult thyroid. *Clin Endocrinol (Oxf)* 1989;31:655-65.
- 8 Maier J, van Steeg H, van Oostrom C, et al. Deoxyribonucleic acid damage and spontaneous mutagenesis in the thyroid gland of rats and mice. *Endocrinology* 2006;147:3391-7.
- 9 Reya T, Morrison SJ, Clarke MF, et al. Stem cells, cancer, and cancer stem cells. *Nature* 2001;414:105-11.
- 10 Molofsky AV, Pardal R, Morrison SJ. Diverse mechanisms regulate stem cell self-renewal. *Curr Opin Cell Biol* 2004;16:700-7.

11 Raff M. Adult stem cell plasticity: fact or artifact? *Annu Rev Cell Dev Biol* 2003;19:1-22.

12 Yatabe Y, Tavare S, Shibata D. Investigating stem cells in human colon by using methylation patterns. *Proc Natl Acad Sci U S A* 2001;98:10839-44.

13 Tai MH, Chang CC, Kiupel M, et al. Oct4 expression in adult human stem cells: evidence in support of the stem cell theory of carcinogenesis. *Carcinogenesis* 2005;26:495-502.

14 Brill S, Zvibel I, Reid LM. Expansion conditions for early hepatic progenitor cells from embryonal and neonatal rat livers. *Dig Dis Sci* 1999;44:364-71.

15 Zulewski H, Abraham EJ, Gerlach MJ, et al. Multipotential nestin-positive stem cells isolated from adult pancreatic islets differentiate ex vivo into pancreatic endocrine, exocrine, and hepatic phenotypes. *Diabetes* 2001;50:521-33.

16 Stemple DL, Anderson DJ. Isolation of a stem cell for neurons and glia from the mammalian neural crest. *Cell* 1992;71:973-85.

17 Pardal R, Clarke MF, Morrison SJ. Applying the principles of stem-cell biology to cancer. *Nat Rev Cancer* 2003;3:895-902.

18 Polyak K, Hahn WC. Roots and stems: stem cells in cancer. *Nat Med* 2006;12:296-300.

19 Gudjonsson T, Magnusson MK. Stem cell biology and the cellular pathways of carcinogenesis. *Apmis* 2005;113:922-9.

20 Takano T, Amino N. Fetal cell carcinogenesis: a new hypothesis for better understanding of thyroid carcinoma. *Thyroid* 2005;15:432-8.

21 Zhang P, Zuo H, Ozaki T, et al. Cancer stem cell hypothesis in thyroid cancer. *Pathol Int* 2006;56:485-9.

## 6. Reference

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22 De Felice M, Di Lauro R. Thyroid development and its disorders: genetics and molecular mechanisms. *Endocr Rev* 2004;25:722-46.

23 Suzuki K, Lavaroni S, Mori A, et al. Autoregulation of thyroid-specific gene transcription by thyroglobulin. *Proc Natl Acad Sci U S A* 1998;95:8251-6.

24 Seto P, Hirayu H, Magnusson RP, et al. Isolation of a complementary DNA clone for thyroid microsomal antigen. Homology with the gene for thyroid peroxidase. *J Clin Invest* 1987;80:1205-8.

25 Civitareale D, Castelli MP, Falasca P, et al. Thyroid transcription factor 1 activates the promoter of the thyrotropin receptor gene. *Mol Endocrinol* 1993;7:1589-95.

26 Miccadei S, De Leo R, Zammarchi E, et al. The synergistic activity of thyroid transcription factor 1 and Pax 8 relies on the promoter/enhancer interplay. *Mol Endocrinol* 2002;16:837-46.

27 Mansouri A, Chowdhury K, Gruss P. Follicular cells of the thyroid gland require Pax8 gene function. *Nat Genet* 1998;19:87-90.

28 Damante G, Tell G, Di Lauro R. A unique combination of transcription factors controls differentiation of thyroid cells. *Prog Nucleic Acid Res Mol Biol* 2001;66:307-56.

29 Ambesi-Impiombato FS, Parks LA, Coon HG. Culture of hormone-dependent functional epithelial cells from rat thyroids. *Proc Natl Acad Sci U S A* 1980;77:3455-9.

30 Dai G, Levy O, Carrasco N. Cloning and characterization of the thyroid iodide transporter. *Nature* 1996;379:458-60.

31 Weiss SJ, Philp NJ, Ambesi-Impiombato FS, et al. Thyrotropin-stimulated iodide transport mediated by adenosine 3',5'-monophosphate and dependent on protein synthesis. *Endocrinology* 1984;114:1099-107.

## 6. Reference

---

32 Davies T, Marians R, Latif R. The TSH receptor reveals itself. *J Clin Invest* 2002;110:161-4.

33 Stem cells: Scientific Progress and Future Research Directions. National Institute of Health. June 2001. (<http://stemcells.nih.gov/info/scireport/2001report.htm>)

34 Thomas T, Nowka K, Lan L, et al. Expression of endoderm stem cell markers: evidence for the presence of adult stem cells in human thyroid glands. *Thyroid* 2006;16:537-44.

35 Pesce M, Scholer HR. Oct-4: gatekeeper in the beginnings of mammalian development. *Stem Cells* 2001;19:271-8.

36 Dyce PW, Zhu H, Craig J, et al. Stem cells with multilineage potential derived from porcine skin. *Biochem Biophys Res Commun* 2004;316:651-8.

37 Chen J, Hersmus N, Van Duppen V, et al. The adult pituitary contains a cell population displaying stem/progenitor cell and early embryonic characteristics. *Endocrinology* 2005;146:3985-98.

38 Arceci RJ, King AA, Simon MC, et al. Mouse GATA-4: a retinoic acid-inducible GATA-binding transcription factor expressed in endodermally derived tissues and heart. *Mol Cell Biol* 1993;13:2235-46.

39 Molkentin JD. The zinc finger-containing transcription factors GATA-4, -5, and -6. Ubiquitously expressed regulators of tissue-specific gene expression. *J Biol Chem* 2000;275:38949-52.

40 Kiiveri S, Liu J, Westerholm-Ormio M, et al. Differential expression of GATA-4 and GATA-6 in fetal and adult mouse and human adrenal tissue. *Endocrinology* 2002;143:3136-43.

41 Ketola I, Pentikainen V, Vaskivuo T, et al. Expression of transcription factor GATA-4 during human testicular development and disease. *J Clin Endocrinol Metab* 2000;85:3925-31.



42 LaVoie HA. The role of GATA in mammalian reproduction. *Exp Biol Med* (Maywood) 2003;228:1282-90.

43 Heikinheimo M, Ermolaeva M, Bielinska M, et al. Expression and hormonal regulation of transcription factors GATA-4 and GATA-6 in the mouse ovary. *Endocrinology* 1997;138:3505-14.

44 Duncan SA, Manova K, Chen WS, et al. Expression of transcription factor HNF-4 in the extraembryonic endoderm, gut, and nephrogenic tissue of the developing mouse embryo: HNF-4 is a marker for primary endoderm in the implanting blastocyst. *Proc Natl Acad Sci U S A* 1994;91:7598-602.

45 Levinson-Dushnik M, Benvenisty N. Involvement of hepatocyte nuclear factor 3 in endoderm differentiation of embryonic stem cells. *Mol Cell Biol* 1997;17:3817-22.

46 Rocchi E, Khodjakov A, Volk EL, et al. The product of the ABC half-transporter gene ABCG2 (BCRP/MXR/ABCP) is expressed in the plasma membrane. *Biochem Biophys Res Commun* 2000;271:42-6.

47 Bunting KD. ABC transporters as phenotypic markers and functional regulators of stem cells. *Stem Cells* 2002;20:11-20.

48 Alison MR. Tissue-based stem cells: ABC transporter proteins take centre stage. *J Pathol* 2003;200:547-50.

49 Doyle LA, Ross DD. Multidrug resistance mediated by the breast cancer resistance protein BCRP (ABCG2). *Oncogene* 2003;22:7340-58.

50 Doyle LA, Yang W, Abruzzo LV, et al. A multidrug resistance transporter from human MCF-7 breast cancer cells. *Proc Natl Acad Sci U S A* 1998;95:15665-70.

51 Bhattacharya S, Jackson JD, Das AV, et al. Direct identification and enrichment of retinal stem cells/progenitors by Hoechst dye efflux assay. *Invest Ophthalmol Vis Sci* 2003;44:2764-73.

52 Martin CM, Meeson AP, Robertson SM, et al. Persistent expression of the ATP-binding cassette transporter, *Abcg2*, identifies cardiac SP cells in the developing and adult heart. *Dev Biol* 2004;265:262-75.

53 Zhou S, Schuetz JD, Bunting KD, et al. The ABC transporter *Bcrp1/ABCG2* is expressed in a wide variety of stem cells and is a molecular determinant of the side-population phenotype. *Nat Med* 2001;7:1028-34.

54 Goodell MA, Brose K, Paradis G, et al. Isolation and functional properties of murine hematopoietic stem cells that are replicating in vivo. *J Exp Med* 1996;183:1797-806.

55 Goodell MA, McKinney-Freeman S, Camargo FD. Isolation and characterization of side population cells. *Methods Mol Biol* 2005;290:343-52.

56 Plachov D, Chowdhury K, Walther C, et al. *Pax8*, a murine paired box gene expressed in the developing excretory system and thyroid gland. *Development* 1990;110:643-51.

57 Espinoza CR, Schmitt TL, Loos U. Thyroid transcription factor 1 and *Pax8* synergistically activate the promoter of the human thyroglobulin gene. *J Mol Endocrinol* 2001;27:59-67.

58 Schmitt TL, Espinoza CR, Loos U. Transcriptional regulation of the human sodium/iodide symporter gene by *Pax8* and TTF-1. *Exp Clin Endocrinol Diabetes* 2001;109:27-31.

59 Komatsu M, Takahashi T, Takahashi I, et al. Thyroid dysgenesis caused by *PAX8* mutation: the hypermutability with CpG dinucleotides at codon 31. *J Pediatr* 2001;139:597-9.

60 Flamant F, Pogue AL, Plateroti M, et al. Congenital hypothyroid *Pax8*(*-/-*) mutant mice can be rescued by inactivating the *TRalpha* gene. *Mol Endocrinol* 2002;16:24-32.

61 Pasca di Magliano M, Di Lauro R, Zannini M. *Pax8* has a key role in thyroid cell differentiation. *Proc Natl Acad Sci U S A* 2000;97:13144-9.

62 Bonner WA, Hulett HR, Sweet RG, et al. Fluorescence activated cell sorting. *Rev Sci Instrum* 1972;43:404-9.

63 Herzenberg LA, De Rosa SC. Monoclonal antibodies and the FACS: complementary tools for immunobiology and medicine. *Immunol Today* 2000;21:383-90.

64 Julius MH, Masuda T, Herzenberg LA. Demonstration that antigen-binding cells are precursors of antibody-producing cells after purification with a fluorescence-activated cell sorter. *Proc Natl Acad Sci U S A* 1972;69:1934-8.

65 Larderet G, Fortunel NO, Vaigot P, et al. Human side population keratinocytes exhibit long-term proliferative potential and a specific gene expression profile and can form a pluristratified epidermis. *Stem Cells* 2006;24:965-74.

66 Decraene C, Benchaour R, Dillies MA, et al. Global transcriptional characterization of SP and MP cells from the myogenic C2C12 cell line: effect of FGF6. *Physiol Genomics* 2005;23:132-49.

67 Meeson AP, Hawke TJ, Graham S, et al. Cellular and molecular regulation of skeletal muscle side population cells. *Stem Cells* 2004;22:1305-20.

68 Summer R, Kotton DN, Sun X, et al. Side population cells and Bcrp1 expression in lung. *Am J Physiol Lung Cell Mol Physiol* 2003;285:L97-104.

69 Majka SM, Beutz MA, Hagen M, et al. Identification of novel resident pulmonary stem cells: form and function of the lung side population. *Stem Cells* 2005;23:1073-81.

70 Chen ZS, Robey RW, Belinsky MG, et al. Transport of methotrexate, methotrexate polyglutamates, and 17beta-estradiol 17-(beta-D-glucuronide) by ABCG2: effects of acquired mutations at R482 on methotrexate transport. *Cancer Res* 2003;63:4048-54.

71 Shimano K, Satake M, Okaya A, et al. Hepatic oval cells have the side population phenotype defined by expression of ATP-binding cassette transporter ABCG2/BCRP1. *Am J Pathol* 2003;163:3-9.

72 Lassalle B, Bastos H, Louis JP, et al. 'Side Population' cells in adult mouse testis express Bcrp1 gene and are enriched in spermatogonia and germinal stem cells. *Development* 2004;131:479-87.

73 Falciatori I, Borsellino G, Haliassos N, et al. Identification and enrichment of spermatogonial stem cells displaying side-population phenotype in immature mouse testis. *Faseb J* 2004;18:376-8.

74 Iwatani H, Ito T, Imai E, et al. Hematopoietic and nonhematopoietic potentials of Hoechst(low)/side population cells isolated from adult rat kidney. *Kidney Int* 2004;65:1604-14.

75 Yano S, Ito Y, Fujimoto M, et al. Characterization and localization of side population cells in mouse skin. *Stem Cells* 2005;23:834-41.

76 Kim M, Morshead CM. Distinct populations of forebrain neural stem and progenitor cells can be isolated using side-population analysis. *J Neurosci* 2003;23:10703-9.

77 Umemoto T, Yamato M, Nishida K, et al. Limbal epithelial side-population cells have stem cell-like properties, including quiescent state. *Stem Cells* 2006;24:86-94.

78 Dontu G, Abdallah WM, Foley JM, et al. In vitro propagation and transcriptional profiling of human mammary stem/progenitor cells. *Genes Dev* 2003;17:1253-70.

79 Behbod F, Xian W, Shaw CA, et al. Transcriptional profiling of mammary gland side population cells. *Stem Cells* 2006;24:1065-74.

80 Alvi AJ, Clayton H, Joshi C, et al. Functional and molecular characterisation of mammary side population cells. *Breast Cancer Res* 2003;5:R1-8.

## 6. Reference

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81 Watanabe K, Nishida K, Yamato M, et al. Human limbal epithelium contains side population cells expressing the ATP-binding cassette transporter ABCG2. *FEBS Lett* 2004;565:6-10.

82 Bhatt RI, Brown MD, Hart CA, et al. Novel method for the isolation and characterisation of the putative prostatic stem cell. *Cytometry A* 2003;54:89-99.

83 Forbes SJ, Alison MR. Side population (SP) cells: taking center stage in regeneration and liver cancer? *Hepatology* 2006;44:23-6.

84 Reynolds BA, Tetzlaff W, Weiss S. A multipotent EGF-responsive striatal embryonic progenitor cell produces neurons and astrocytes. *J Neurosci* 1992;12:4565-74.

85 Reynolds BA, Weiss S. Clonal and population analyses demonstrate that an EGF-responsive mammalian embryonic CNS precursor is a stem cell. *Dev Biol* 1996;175:1-13.

86 Morshead CM, Reynolds BA, Craig CG, et al. Neural stem cells in the adult mammalian forebrain: a relatively quiescent subpopulation of subependymal cells. *Neuron* 1994;13:1071-82.

87 Uchida N, Buck DW, He D, et al. Direct isolation of human central nervous system stem cells. *Proc Natl Acad Sci U S A* 2000;97:14720-5.

88 Rietze RL, Valcanis H, Brooker GF, et al. Purification of a pluripotent neural stem cell from the adult mouse brain. *Nature* 2001;412:736-9.

89 Johansson CB, Momma S, Clarke DL, et al. Identification of a neural stem cell in the adult mammalian central nervous system. *Cell* 1999;96:25-34.

90 Dontu G, Wicha MS. Survival of mammary stem cells in suspension culture: implications for stem cell biology and neoplasia. *J Mammary Gland Biol Neoplasia* 2005;10:75-86.

91 Toma JG, Akhavan M, Fernandes KJ, et al. Isolation of multipotent adult stem cells from the dermis of mammalian skin. *Nat Cell Biol* 2001;3:778-84.

92 Beltrami AP, Barlucchi L, Torella D, et al. Adult cardiac stem cells are multipotent and support myocardial regeneration. *Cell* 2003;114:763-76.

93 Li H, Liu H, Heller S. Pluripotent stem cells from the adult mouse inner ear. *Nat Med* 2003;9:1293-9.

94 Tropepe V, Coles BL, Chiasson BJ, et al. Retinal stem cells in the adult mammalian eye. *Science* 2000;287:2032-6.

95 Seaberg RM, Smukler SR, Kieffer TJ, et al. Clonal identification of multipotent precursors from adult mouse pancreas that generate neural and pancreatic lineages. *Nat Biotechnol* 2004;22:1115-24.

96 Derwahl M, Manole D, Sobke A, et al. Pathogenesis of toxic thyroid adenomas and nodules: relevance of activating mutations in the TSH-receptor and Gs-alpha gene, the possible role of iodine deficiency and secondary and TSH-independent molecular mechanisms. *Exp Clin Endocrinol Diabetes* 1998;106 Suppl 4:S6-9.

97 Jones HW, Jr., McKusick VA, Harper PS, et al. George Otto Gey. (1899-1970). The HeLa cell and a reappraisal of its origin. *Obstet Gynecol* 1971;38:945-9.

98 Abbott BL, Colapietro AM, Barnes Y, et al. Low levels of ABCG2 expression in adult AML blast samples. *Blood* 2002;100:4594-601.

99 Robey RW, Steadman K, Polgar O, et al. Pheophorbide a is a specific probe for ABCG2 function and inhibition. *Cancer Res* 2004;64:1242-6.

100 de Paiva CS, Chen Z, Corrales RM, et al. ABCG2 transporter identifies a population of clonogenic human limbal epithelial cells. *Stem Cells* 2005;23:63-73.

101 Epstein SP, Wolosin JM, Asbell PA. P63 expression levels in side population and low light scattering ocular surface epithelial cells. *Trans Am Ophthalmol Soc* 2005;103:187-99.

102 Chambard M, Gabrion J, Mauchamp J. Influence of collagen gel on the orientation of epithelial cell polarity: follicle formation from isolated thyroid cells and from preformed monolayers. *J Cell Biol* 1981;91:157-66.

103 Kraiem Z, Sadeh O, Yosef M. Iodide uptake and organification, tri-iodothyronine secretion, cyclic AMP accumulation and cell proliferation in an optimized system of human thyroid follicles cultured in collagen gel suspended in serum-free medium. *J Endocrinol* 1991;131:499-506.

104 Pan GJ, Chang ZY, Scholer HR, et al. Stem cell pluripotency and transcription factor Oct4. *Cell Res* 2002;12:321-9.

105 Moore KA, Lemischka IR. Stem cells and their niches. *Science* 2006;311:1880-5.

106 Hawke TJ, Garry DJ. Myogenic satellite cells: physiology to molecular biology. *J Appl Physiol* 2001;91:534-51.

107 Murayama A, Matsuzaki Y, Kawaguchi A, et al. Flow cytometric analysis of neural stem cells in the developing and adult mouse brain. *J Neurosci Res* 2002;69:837-47.

108 Romano AC, Espana EM, Yoo SH, et al. Different cell sizes in human limbal and central corneal basal epithelia measured by confocal microscopy and flow cytometry. *Invest Ophthalmol Vis Sci* 2003;44:5125-9.

109 Kukekov VG, Laywell ED, Thomas LB, et al. A nestin-negative precursor cell from the adult mouse brain gives rise to neurons and glia. *Glia* 1997;21:399-407.

110 Kukekov VG, Laywell ED, Suslov O, et al. Multipotent stem/progenitor cells with similar properties arise from two neurogenic regions of adult human brain. *Exp Neurol* 1999;156:333-44.

111 Al-Hajj M, Becker MW, Wicha M, et al. Therapeutic implications of cancer stem cells. *Curr Opin Genet Dev* 2004;14:43-7.

## 6. Reference

---

112 Al-Hajj M, Wicha MS, Benito-Hernandez A, et al. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci U S A* 2003;100:3983-8.

113 Brabletz T, Jung A, Spaderna S, et al. Opinion: migrating cancer stem cells - an integrated concept of malignant tumour progression. *Nat Rev Cancer* 2005;5:744-9.

114 Schuldiner M, Yanuka O, Itskovitz-Eldor J, et al. Effects of eight growth factors on the differentiation of cells derived from human embryonic stem cells. *Proc Natl Acad Sci U S A* 2000;97:11307-12.

115 Fujikura J, Yamato E, Yonemura S, et al. Differentiation of embryonic stem cells is induced by GATA factors. *Genes Dev* 2002;16:784-9.

116 Di Palma T, Nitsch R, Mascia A, et al. The paired domain-containing factor Pax8 and the homeodomain-containing factor TTF-1 directly interact and synergistically activate transcription. *J Biol Chem* 2003;278:3395-402.

117 Staud F, Pavek P. Breast cancer resistance protein (BCRP/ABCG2). *Int J Biochem Cell Biol* 2005;37:720-5.

118 Krishnamurthy P, Ross DD, Nakanishi T, et al. The stem cell marker Bcrp/ABCG2 enhances hypoxic cell survival through interactions with heme. *J Biol Chem* 2004;279:24218-25.

119 Hadnagy A, Gaboury L, Beaulieu R, et al. SP analysis may be used to identify cancer stem cell populations. *Exp Cell Res* 2006;312:3701-10.

120 Scharenberg CW, Harkey MA, Torok-Storb B. The ABCG2 transporter is an efficient Hoechst 33342 efflux pump and is preferentially expressed by immature human hematopoietic progenitors. *Blood* 2002;99:507-12.

121 Dekaney CM, Rodriguez JM, Graul MC, et al. Isolation and characterization of a putative intestinal stem cell fraction from mouse jejunum. *Gastroenterology* 2005;129:1567-80.



## 6. Reference

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122 Parmar K, Sauk-Schubert C, Burdick D, et al. Sca+CD34- murine side population cells are highly enriched for primitive stem cells. *Exp Hematol* 2003;31:244-50.

123 Matsuzaki Y, Kinjo K, Mulligan RC, et al. Unexpectedly efficient homing capacity of purified murine hematopoietic stem cells. *Immunity* 2004;20:87-93.

124 Camargo FD, Chambers SM, Drew E, et al. Hematopoietic stem cells do not engraft with absolute efficiencies. *Blood* 2006;107:501-7.

125 De Paiva CS, Pflugfelder SC, Li DQ. Cell size correlates with phenotype and proliferative capacity in human corneal epithelial cells. *Stem Cells* 2006;24:368-75.

126 Benchaouir R, Rameau P, Decraene C, et al. Evidence for a resident subset of cells with SP phenotype in the C2C12 myogenic line: a tool to explore muscle stem cell biology. *Exp Cell Res* 2004;294:254-68.

127 Woodward WA, Chen MS, Behbod F, et al. On mammary stem cells. *J Cell Sci* 2005;118:3585-94.

128 Bechtner G, Schopohl D, Rafferteder M, et al. Stimulation of thyroid cell proliferation by epidermal growth factor is different from cell growth induced by thyrotropin or insulin-like growth factor I. *Eur J Endocrinol* 1996;134:639-48.

129 Trosko JE, Chang CC, Wilson MR, et al. Gap junctions and the regulation of cellular functions of stem cells during development and differentiation. *Methods* 2000;20:245-64.

130 Chang CC, Trosko JE, el-Fouly MH, et al. Contact insensitivity of a subpopulation of normal human fetal kidney epithelial cells and of human carcinoma cell lines. *Cancer Res* 1987;47:1634-45.

131 Frisch SM, Francis H. Disruption of epithelial cell-matrix interactions induces apoptosis. *J Cell Biol* 1994;124:619-26.

## 6. Reference

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132 Schofield R. The relationship between the spleen colony-forming cell and the haemopoietic stem cell. *Blood Cells* 1978;4:7-25.

133 Spradling A, Drummond-Barbosa D, Kai T. Stem cells find their niche. *Nature* 2001;414:98-104.

134 Conti L, Pollard SM, Gorba T, et al. Niche-independent symmetrical self-renewal of a mammalian tissue stem cell. *PLoS Biol* 2005;3:e283.

135 Thompson SD, Franklyn JA, Watkinson JC, et al. Fibroblast growth factors 1 and 2 and fibroblast growth factor receptor 1 are elevated in thyroid hyperplasia. *J Clin Endocrinol Metab* 1998;83:1336-41.

136 Bidey SP, Hill DJ, Eggo MC. Growth factors and goitrogenesis. *J Endocrinol* 1999;160:321-32.

137 Delange F. The disorders induced by iodine deficiency. *Thyroid* 1994;4:107-28.

138 Ollis CA, Davies R, Munro DS, et al. Relationship between growth and function of human thyroid cells in culture. *J Endocrinol* 1986;108:393-8.

139 Eszlinger M, Krohn K, Kratzsch J, et al. Growth factor expression in cold and hot thyroid nodules. *Thyroid* 2001;11:125-35.

140 Dremier S, Golstein J, Mosselmans R, et al. Apoptosis in dog thyroid cells. *Biochem Biophys Res Commun* 1994;200:52-8.

141 di Jeso B, Ulianich L, Racioppi L, et al. Serum withdrawal induces apoptotic cell death in Ki-ras transformed but not in normal differentiated thyroid cells. *Biochem Biophys Res Commun* 1995;214:819-24.

142 Di Matola T, Mueller F, Fenzi G, et al. Serum withdrawal-induced apoptosis in thyroid cells is caused by loss of fibronectin-integrin interaction. *J Clin Endocrinol Metab* 2000;85:1188-93.

143 Kawakami A, Eguchi K, Matsuoka N, et al. Thyroid-stimulating hormone inhibits Fas antigen-mediated apoptosis of human thyrocytes in vitro. *Endocrinology* 1996;137:3163-9.

144 Lin RY, Kubo A, Keller GM, et al. Committing embryonic stem cells to differentiate into thyrocyte-like cells in vitro. *Endocrinology* 2003;144:2644-9.

145 Marians RC, Ng L, Blair HC, et al. Defining thyrotropin-dependent and -independent steps of thyroid hormone synthesis by using thyrotropin receptor-null mice. *Proc Natl Acad Sci U S A* 2002;99:15776-81.

146 Parmentier M, Libert F, Maenhaut C, et al. Molecular cloning of the thyrotropin receptor. *Science* 1989;246:1620-2.

147 Huber GK, Davies TF. Human fetal thyroid cell growth in vitro: system characterization and cytokine inhibition. *Endocrinology* 1990;126:869-75.

148 Davies TF, Platzer M, Schwartz AE, et al. Short- and long-term evaluation of normal and abnormal human thyroid cells in monolayer culture. *Clin Endocrinol (Oxf)* 1985;23:469-79.

149 Davies TF, Platzer M, Schwartz AE, et al. Thyroglobulin secretion by human thyroid cells after monolayer culture--comparison of normal and adenomatous cells. *Clin Endocrinol (Oxf)* 1984;21:239-46.

150 Postiglione MP, Parlato R, Rodriguez-Mallon A, et al. Role of the thyroid-stimulating hormone receptor signaling in development and differentiation of the thyroid gland. *Proc Natl Acad Sci U S A* 2002;99:15462-7.

151 Arufe MC, Lu M, Kubo A, et al. Directed differentiation of mouse embryonic stem cells into thyroid follicular cells. *Endocrinology* 2006;147:3007-15.

152 Peter HJ, Gerber H, Studer H, et al. Pathogenesis of heterogeneity in human multinodular goiter. A study on growth and function of thyroid tissue transplanted onto nude mice. *J Clin Invest* 1985;76:1992-2002.

153 Peter HJ, Studer H, Groscurth P. Autonomous growth, but not autonomous function, in embryonic human thyroids: a clue to understanding autonomous goiter growth? *J Clin Endocrinol Metab* 1988;66:968-73.

154 Berghout A, Wiersinga WM, Smits NJ, et al. Interrelationships between age, thyroid volume, thyroid nodularity, and thyroid function in patients with sporadic nontoxic goiter. *Am J Med* 1990;89:602-8.

155 Vanderpump MP, Tunbridge WM, French JM, et al. The incidence of thyroid disorders in the community: a twenty-year follow-up of the Wickham Survey. *Clin Endocrinol (Oxf)* 1995;43:55-68.

156 Tamura M, Kimura H, Koji T, et al. Role of apoptosis of thyrocytes in a rat model of goiter. A possible involvement of Fas system. *Endocrinology* 1998;139:3646-53.